

Personalized Clinical Decision Making Through Implementation of a Molecular Tumor Board: A German Single-Center Experience

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Purpose Dramatic advances in our understanding of the molecular pathophysiology of cancer, along with a rapidly expanding portfolio of molecular targeted drugs, have led to a paradigm shift toward personalized, biomarker-driven cancer treatment. Here, we report the 2-year experience of the Comprehensive Cancer Center Freiburg Molecular Tumor Board (MTB), one of the first interdisciplinary molecular tumor conferences established in Europe. The role of the MTB is to recommend personalized therapy for patients with cancer beyond standard-of-care treatment.

Methods This retrospective case series includes 198 patients discussed from March 2015 through February 2017. The MTB guided individual molecular diagnostics, assessed evidence of actionability of molecular alterations, and provided therapy recommendations, including approved and off-label treatments as well as available matched clinical trials.

Results The majority of patients had metastatic solid tumors (73.7%), mostly progressive (77.3%) after a mean of 2.0 lines of standard treatment. Diagnostic recommendations resulted in 867 molecular diagnostic tests for 172 patients (five per case), including exome analysis in 36 cases (18.2%). With a median turnaround time of 28 days, treatment recommendations were given to 104 patients (52.5%). These included single-agent targeted therapies (42.3%), checkpoint inhibitors (37.5%), and combination therapies (18.3%). Treatment recommendations were implemented in 33 of 104 patients (31.7%), of whom 19 (57.6%) showed stable disease or partial response, including 14 patients (7.1% of the entire population) receiving off-label treatments.

Conclusion Personalized extended molecular-guided patient care is effective for a small but clinically meaningful proportion of patients in challenging clinical situations. Limited access to targeted drugs, lack of trials, and submission at late disease stage prevents broader applicability, whereas genome-wide analyses are not a strict requirement for predictive molecular testing.

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INTRODUCTION

Personalized cancer medicine uses molecular biomarkers for standard-of-care treatment stratification, such as activating BRAF mutations for the treatment of melanoma with BRAF inhibitors.1 In parallel, it has become evident that therapeutic strategies with targeted drugs are no longer specific for the treatment of distinct entities but rather for particular molecular profiles across different cancers.2-4 Thus, testing for single-drug targets can provide therapeutic information, but its predictive value may vary between entities. Although an activating BRAF V600E mutation will predict response to BRAF inhibitors in melanoma,1 it may not do so in colorectal cancers because of epidermal growth factor receptor (EGFR) feedback activation with requirement of additional EGFR targeting.^{5,6} Moreover, non-V600 BRAF mutations might not be responsive to BRAF inhibition at all.7 Thus, one-mutation-one-drug approaches may be ineffective, especially in heavily pretreated patients with cancer. Underlying causes include

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the challenge to discriminate relevant mutations and pathway aberrations from background and passenger mutations,8 the clonal molecular intra- and intertumoral heterogeneity,9,10 and dynamic changes in the molecular composition of cancer, especially if treatment leads to selection of resistant subclones. Examples include the selection of RAS mutant clones in colorectal cancer treated with EGFR antibodies, such as cetuximab or panitumumab, 11 or the acquisition of a secondary EGFR T790M kinase domain mutation mediating resistance to EGFR kinase inhibitors, such as gefitinib or erlotinib in nonsmall-cell lung cancer. 12,13

This increasing amount of complexity requires tools to translate individual information into personalized treatment concepts. A molecular tumor board (MTB) represents a platform that integrates clinical and molecular parameters for clinical decision making. Here, we report the 2-year experience of the Comprehensive Cancer Center Freiburg MTB that provides personalized treatment recommendations on the basis of individual molecular diagnostics. We hereby present detailed data on patient characteristics, treatment recommendations, clinical adherence to recommendations, and outcomes of treated patients.

METHODS

MTB Implementation and Organization

The MTB is run by an interdisciplinary team of medical and scientific experts with a focus on clinical and translational oncology and computational and molecular biology. Cases are submitted using an online registration and documentation system (Appendix Fig A1). Each case is assigned to a clinician scientist with expertise in the specific cancer type (entity expert), who reviews the literature and available clinical trials. In parallel, the molecular pathology team reviews the individual tumor pathology and sets up a presentation of already performed and suggested diagnostic tests. The initial discussion includes a clinical case presentation, review of the pathology data and the tumor-specific genetic landscape, known molecular predictive or prognostic markers, active clinical trials, and potential in- and off-label molecular targeted treatments. The molecular diagnostic requests are performed using certified and standard operating procedure (SOP)-driven processes.

Diagnostic results are presented to the MTB by the molecular pathology and/or the computational biologist team. After discussion, treatment recommendations are given and are supported by levels of evidence (Data Supplement). These are based on published molecular biomarker recommendations.14

Patients and Patient Informed Consent

All patients discussed (n = 198) were included in this retrospective single-center case series. All molecular diagnostic tests were conducted in accordance with the medical treatment contract signed by each patient. Patient tissue was stored in the local biobank and required a signed informed consent, approved by the University of Freiburg institutional review board. Patients with individual or family history indicative of germline disease-causing mutations were referred to the Institute of Human Genetics for counseling and possibly germline genetic analyses.

Diagnostic Molecular Pathology

Appropriate tissues were subjected to molecular analyses as recommended by the MTB (Fig 1). All analyses were carried out according to routine pathology laboratory testing procedures, with assays being nationally validated and certified. Targeted next-generation sequencing (tNGS) included a custom-designed hotspot eight-gene panel (designed by S.L. and produced by Illumina, San Diego, CA), a BRCA1/2 panel (produced by Illumina), a hotspot 48-gene panel (TruSeq Amplicon Cancer Panel, Illumina), and a 54-gene myeloid panel (TruSight Myeloid Sequencing Panel, Illumina).15-17

Investigational Genetic Tumor Characterization

Whole-exome sequencing (WES) and RNA sequencing (RNA-Seq) were performed on tumor tissue. Complementary germline DNA was obtained from peripheral blood or healthy tissue. Only nonsynonymous mutations detected with a variant allele frequency > 10% and listed with a minor allele frequency < 0.001% by the Exome Aggregation Consortium¹⁸ were reported. Single nucleotide variations were classified according to ClinVar,19 COSMIC,20 dbSNP,21,22

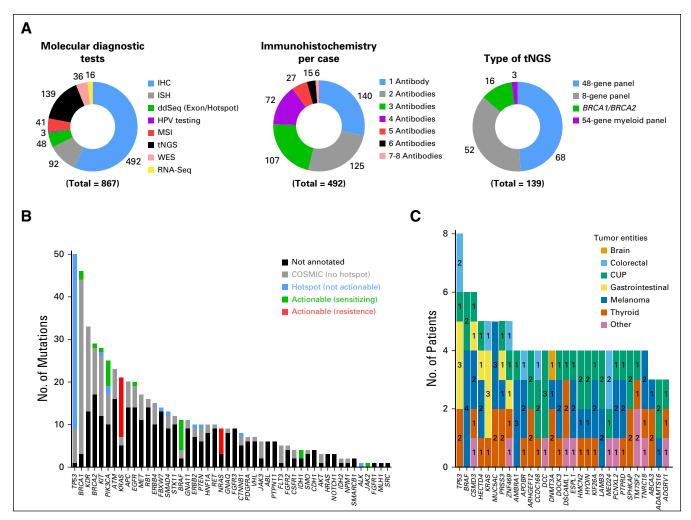


Fig 1. Molecular diagnostic testing. (A) The panels depict the type of molecular diagnostic testing performed (left panel) and specify the number of immunohistochemical stains (one to eight antibodies) per case (middle panel) as well as the type of targeted next-generation sequencing (tNGS) library sequenced (right panel). tNGS was performed either by a custom panel (eight-gene panel), a 48-gene panel (TruSeq Amplicon Cancer Panel, Illumina, San Diego, CA), a 54-gene myeloid panel (TruSight Myeloid Sequencing Panel, Illumina) or a custom BRCA1/2 consortium panel. (B) The bar plot depicts the number of sequence variants detected in tumor DNA of 139 patients using tNGS. The bars indicate the numbers of mutations in a given gene (black) and sequence variants that are annotated in COSMIC (gray). The numbers of actionable mutations is shown in green (drug sensitizing) and red (drug resistance). (C) The bar plot depicts the 30 most frequently somatic mutated genes of 36 patients analyzed by whole-exome sequencing (WES). The colors indicate different tumor entities. Mutations with a variant allele frequency > 10% and a minor allele frequency < 0.001 were considered. The GI tumor category includes liver, pancreas, stomach, and esophagus. CUP, carcinoma of unknown primary; HPV, human papillomavirus; IHC, immunohistochemistry; ISH, in situ hybridization; MSI, microsatellite instability.

hotspot mutation^{23,24} (http://cancerhotspots.org/ #/home), TARGET db (http://archive.broadinstitute.org/cancer/cga/target), drug-gene interaction (DGIdb; http://www.dgidb.org),25 and CADD (http://cadd.gs.washington.edu), and categorized according to the predicted impact on protein function by Condel.²⁶⁻³¹ Copy number alteration analysis was performed using Control-FREEC.32 The STAR33 aligner was used to align and infer the gene expression level. FusionCatcher (https://doi.org/10.1101/011650) was used to predict gene fusions. Differentially expressed genes were identified using the limmavoom package from R/Bioconductor.34,35

RESULTS

From March 2015 through February 2017, 49 MTB meetings were attended by a median of 16 physicians and scientists, ensuring continuous interdisciplinary data interpretation and discussions with diagnostic and therapeutic decision making. The workflow of the MTB included a case and literature review, molecular diagnostic recommendations, and follow-up discussions of the molecular diagnostic results, including treatment recommendations (Appendix Fig A1). Thus, a total of 385 case discussions were held for 198 patients (1.9 discussions per patient;

Table 1. Results

Recommendations	No. (%)
Meetings, No.	49
Case discussions, total No. (per-patient average)	385 (1.9)
Recommendations, No.	505
Diagnostic	305 (60.4)
Therapeutic	104 (20.6)
No treatment recommendation	77 (15.2)
Conditional recommendation	14 (2.8)
Genetic counseling	4 (0.8)
Referral to organ board	1 (0.2)
Diagnostic recommendations, total No. (per-patient average)	305 (1.5)
Patients with diagnostic recommendations	172 (86.9)
Routine pathology	153 (89.0)
Extended genetic analysis*	69 (40.1)
Rebiopsy	15 (8.7)
Other	6 (3.5)
Diagnostic recommendations, not implemented	71 (23.3)
Patients with treatment recommendations	104 (52.5)
Implemented	33 (31.7)
Partial response	11 (33.3)
Stable disease	8 (24.2)
No. of treatment recommendations	104
Treatment recommendations, not implemented	71 (68.3)

NOTE: Data presented as No. (%) unless otherwise noted. See Figure 2 and Data Supplement for details of treatment recommendations and treatment recommendations not implemented. *Whole-exome sequencing, RNA-seq.

Table 1). In total, 505 structured recommendations were given (2.5 per patient; Table 1). These included 305 diagnostic and 104 treatment recommendations.

Patient Characteristics

The average patient age at the time of the initial MTB presentation was 58 years (range, 1 to 85 years). Detailed patient characteristics are listed in Table 2. One hundred ninety-one of 198 patients (96.4%) had an underlying malignant condition. Patients with solid tumors largely outbalanced hematopoietic malignancies (95.5% v 4.5%). Soft tissue tumors (12.6%), CNS tumors (11.1%), and carcinoma of unknown primary (CUP; 10.1%) were the most frequent tumor entities. The majority of patients (n = 146; 73.7%) suffered from metastatic disease, and 77.3% (n = 153) showed disease progression while receiving the standard treatment (Table 2). The mean time interval from diagnosis to first

MTB discussion was 33.6 months (range, 1 to 541 months). Patients with treatment-refractory metastatic disease had undergone a mean of 2.0 (range, one to 11) lines of systemic pretreatments. A minority of the patients was referred to the board with rare tumors (n = 33; 16.7%) or because of young age (n = 3; 1.5%).

Molecular Diagnostic Testing

The distribution of molecular diagnostic recommendations is shown in Table 1. For 172 patients (86.9%), 305 recommendations were given and included routine molecular tests in 153 (89%), extended genetic analysis in 69 (40.1%), and both in 53 (30.8%) patients. Rebiopsies were recommended in 15 cases, mostly because of lack of adequate tissue. Of all diagnostic recommendations, 234 (76.7%) were implemented, resulting in 867 single diagnostic tests (mean, five per patient), including 815 routine molecular tests and 52 extended genetic analyses (Fig 1A, left panel).

Routine molecular diagnostics included immunohistochemical (IHC) staining for biomarkers (n = 492; Fig 1A, middle panel), such as programmed death-ligand 1 (PD-L1) and mismatch repair proteins, in situ hybridizations (ISH) for gene copy number analyses (n = 92), and testing for microsatellite instability and/or gene hotspot variations (n = 89) and tNGS (n = 139; Fig 1A). The latter included libraries of different gene panels (Fig 1A; right panel). The most frequent COSMIC annotated sequence variants detected by tNGS occurred in TP53, BRCA1, KDR, KIT, KRAS, PIK3CA, BRCA2, and BRAF (Fig 1B; Data Supplement). Therapeutically relevant mutations in hotspot regions were identified in 41 of 139 patients (29.5%), including drug-sensitizing variants in BRAF, PIK3CA, IDH1, EGFR, and KIT, as well as drug resistance variants in KRAS and NRAS.

Extended genetic analyses including exome and transcriptome assays were performed for 36 patients (18.2%; WES and RNA-Seq: n = 35; RNA-Seq only: n = 1). In those, we identified a total of 5,335 variants, including 18 COSMIC annotated hotspot mutations (Data Supplement). Sixteen were classified as therapy relevant according to the DGI and the TARGET databases. Among the remaining non-hotspot mutations, 1,518 were annotated in COSMIC, including 288 and 28 mutations annotated in

Table 2. Patient Characteristics

Characteristic

Total, No.	198
Sex	
Female	84 (42.4)
Male	114 (57.6)
Age, years, median (range)	58 (1-85)
Time interval from diagnosis to first MTB, months, median (range)	33.6 (1-541)
Tumor type	
Soft tissue	25 (12.6)
CNS	22 (11.1)
Unknown primary site	20 (10.1)
Colorectal	19 (9.6)
Urogenital	15 (7.5)
Thyroid	14 (7.0)
Breast	10 (5.1)
Lung	9 (4.5)
Hepatobiliary	8 (4.0)
Skin	8 (4.0)
Upper GI tract	8 (4.0)
Hematologic	8 (4.0)
Neuroendocrine	6 (3.0)
Pediatric	5 (2.5)
Head and neck	1 (0.5)
Others	13 (6.6)
Solid tumors (n = 189): stage at presentation	
Complete remission	1 (0.5)
Localized disease	42 (22.2)
Metastatic disease	146 (77.3)
No. of previous lines of therapy, mean (range)	2.0 (0-11)
0	25 (12.6)
1	66 (33.3)
2 to 3	80 (40.4)
> 3	27 (13.6)
Reason for referral	
Progressive disease after standard treatment	153 (77.3)
Rare tumor	33 (16.7)
Others	12 (6.1)

NOTE: Data presented as No. (%) unless otherwise noted. Abbreviation: MTB, Molecular Tumor Board.

> DGI and TARGET databases, respectively (Data Supplement). A total of 3,799 mutations were unknown to COSMIC (Data Supplement). The disease impact of non-hotspot mutations is more difficult to evaluate; however, it can lead to

additional therapy-relevant insights. For example, the ERBB2 S656F mutation might, according to TARGET and DGI databases, constitute an activating mutation, therefore targetable by trastuzumab or lapatinib. The most frequently mutated genes were TP53 and BRAF (Fig 1C).

Overall, 71 of 305 diagnostic recommendations (23.3%) were not pursued. As shown in the Data Supplement, reasons for nonadherence included technical reasons (53.5%; mostly lack of sufficient tissue or DNA/RNA), patient death (12.7%), loss to follow-up (11.3%), medical reasons (9.9%), or patient will (9.9%).

Treatment Recommendations

Specific treatment recommendations were given to 104 patients (Table 1; Fig 2) and mainly included off-label immune checkpoint inhibitor (CPI; n = 36; 34.6%), off-label targeted therapy (n = 19; 18.3%) with tyrosine kinase inhibitors, small molecules or antibodies that were not CPI (AB), trial inclusions (n = 13; 12.5%), and offlabel combination treatments (n = 18; 17.3%; Data Supplement; Fig 2). Ninety of 104 treatment recommendations (86.5%) were either off-label therapies (n = 77) or trial inclusions (n = 13).

The implementation rate of treatment recommendations was 31.7% (33 of 104). In-label recommendations were pursued in nine of 14 cases (64.3%), whereas off-label recommendations and trial inclusions were implemented in only 28.6% (22 of 77) and 15.4% (two of 13) of the cases, respectively. Intended trial inclusion in 11 patients failed because of poor performance status or patient death (n = 5), closed trial arm (n = 4), or patient will (n = 2). Main reasons for nonimplementation of treatment recommendations included loss to follow-up (22.5%), recommendation in the future (19.7%), patient death (16.9%), patient will (14.1%), and medical reasons (14.1%; Data Supplement). Of note, evidence level of individual off-label recommendations did not affect implementation rates (data not shown).

Clinical Outcome

In 33 patients with implemented treatment recommendations, partial remissions (PR) and stable diseases (SD) were seen in 11 (33.3%) and eight patients (24.2%; Table 1), respectively. After excluding in-label therapies, nine patients

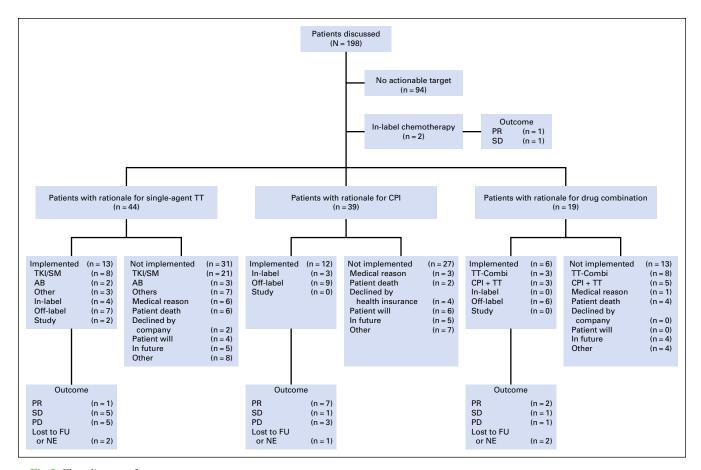


Fig 2. Flow diagram of patients discussed at the Molecular Tumor Board. Responses were determined according to Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1. AB, antibody; Combi, combination; CPI, checkpoint inhibitor; FU, follow-up; NE, not evaluable; PD, progressive disease; PR, partial remission; SD, stable TKI, tyrosine kinase inhibitor; TT, targeted therapy.

achieved PR and five patients SD, resulting in an overall response rate of 4.6% (nine of 198 patients) and a disease control rate (DCR) of 7.1% (14 of 198 patients). Of note, all five patients experiencing SD experienced disease progression while receiving the previous treatment. Of 14 responders receiving off-label therapies, eight (57.1%) showed a progression-free survival (PFS) ratio (PFS2/PFS1; PFSr) > 1.3, disease; SM, small molecule; supporting the impact of the recommended therapies.³⁶ Three patients had a PFSr < 1.3 with ongoing responses, meaning that their PFSr is still increasing. Details about the outcome of responding patients are shown in Table 3. Two individual cases are shown in the Data Supplement. Adherence to recommendations and outcome according to type of treatment is shown in Fig 2. To assess whether implementation of treatment recommendations affected overall survival from first MTB discussion, we analyzed all patients with stage IV malignancies according to three subgroups (n = 148; Fig 3). The median survival was not reached for patients with implemented treatment recommendations (n = 33 recommendations pursued; 95% CI, 9

months to not reached), 8 months for patients for whom treatment recommendations were not implemented (n = 43 recommendations not pursued; 95% CI, 3 to 10 months), and 10 months for patients who did not receive a treatment recommendation (n = 72 no recommendations; 95% CI, 7 to 17 months). Patients who did not receive the recommended therapy because of death before treatment initiation (n = 12) were excluded from analysis.

DISCUSSION

In a cohort of 198 patients with mostly advanced malignancies beyond standard-of-care treatment, the Comprehensive Cancer Center Freiburg MTB identified actionable targets in 52.5% of the cases. Thirty-two percent received the recommended treatment. In 33 patients with implemented treatment recommendations the disease-control rate was 57.6%; it was 9.6% (19 of 198 patients) for the entire cohort. Because the primary goal of an MTB is to give treatment recommendations beyond standard of care, we excluded five responders who received in-label

Table 3. Patients With Tumor Response

Cancer Tyne	Rationale for Treatment Recommendation	Board Recommendation	E	_	~	PFS2 (week)	PFS1 (week)	PFSr	Outcome
Adeno-CUP	Reclassification: histopathological most likely from GI tract	Chemotherapy with FOLFIRI	A1 1		PR	36	NE NE	SE E	PR for 9 months, then PD and death after 14 months
	Immunogenic tumor (PD-L1 positive in cancer cells, abundant TILs, high mutational burden with 358 SNVs and 7 LoH)	Checkpoint inhibition with nivolumab	B3 (Off.	PR	> 76	70	> 1.1	PR since 17 months and ongoing
	Reclassification: histopathological most likely pancreatic cancer	Second-line treatment with gemeitabine and NAB-paclitaxel	A1 1	In	SD	61	17	3.6	For 14 months, then PD
ALL	Combination with JAK inhibitor for synergistic antistromal effect 37	Combination of ruxolitinib and nilotinib analog to CoRNea trial (ClinicalTrials.gov identifier: NCT02253277)	CI	Off	PR	22	39	9.0	Marked decrease of BCR - ABL transcript ratio for 4 months, then PD
Ependymoma	Activation of PI3K/mTOR pathway suggested by positive pS6K staining (70% of tumor cells)	Targeted therapy with sirolimus	C3 (C3	Э	SD	92	11	5.9	For 16 months, then PD
CRC	Immunogenic tumor (PD-L1 positive in 5% of cancer cells, abundant TILs, MSI deficiency)	Checkpoint inhibition with pembrolizumab	A3 (Off	PR	66 <	17	> 5.8	Since 22 months and ongoing
GIST	In-label first-line treatment	Imatinib	A1]	In (SD	NE.	154	ZE	Since 35 months and ongoing
GBM	Immunogenic tumor (PD-L1 positive in 20% of cancer cells, TILs in tumor periphery)	Combination of nivolumab and bevacizumab	B1 (Off]	PD + CI	12	35	0.3	Significant CI after 2 months. Imaging at month 4 demonstrated PD
Lymphoma plus NBCCS	After chemotherapy (Hodgkin lymphoma) the patient is under risk to develop basalioma or other secondary neoplasia because of the underlying NBCCS.	Prophylaxis with hedgehog inhibitor cholecalciferol	3	Off]	Z E	ZE	NE	NE NE	No basaliomas since start of chemotherapy for 13 months and ongoing
Melanoma	BRAF exon 15 mutation D594G described as inhibitory mutation ³⁸	Combination of targeted therapy with sorafenib and trametinib	C3 (Э	SD	11	11	1.0	SD at week 11 with regressive liver metastases. Clinical progression at week 21. Patient died at month 7.
NSCLC	Mutation in EGFR exon 19	Targeted therapy with gefitinib	A1 1	In	PR	NE	32	NE	Initially good PR, then PD after 8 months
Ovary	No molecular rationale but trial availability	Trial (anti-VEGF/ANG2 nanobody BI 836880; ClinicalTrials.gov identifier: NCT02674152)	CI	Э	SD	69	11	6.3	For 16 months, then PD
Squamous-CUP	Immunogenic tumor (51 SNVs and 13 LoH)	Checkpoint inhibition with nivolumab	B3 (Э	SD	13	22	9.0	Stable disease and significant clinical improvement for 4 months. Then PD and death.

(Continued on following page)

Table 3. Patients With Tumor Response (Continued)

E	Rationale for Treatment	Board	į	,	f	PFS2	PFS1		
SCLC	Results from Checkmate-032 demonstrating responses to nivolumab independent of PD-L1 expression (PD-L1 < 1% on cancer cells)	Recommendation Checkpoint inhibition with nivolumab	A3	J JO	PR PR	(week)	(week)	4.3	Since 16 months and ongoing
PNET	Study availability for solid tumors	Trial (anti-VEGF/ANG2 nanobody BI 836880; Clinical Trials.gov identifier: NCT02674152)	CI	ЭО	SD	> 61	169	> 0.4	Since 14 months and ongoing
Sarcoma	Despite nonimmunogenic tumor (15 SNVs, 1 LoH, no PD-L1 on cancer cells, few TILs), recent data show response to checkpoint inhibition (SARC028 study)	Checkpoint inhibition with pembrolizumab	A3	ЭЭO	PR	78	228	> 0.3	Since 18 months and ongoing
Thyroid (medullary)	RET M918T	Targeted therapy with vandetanib	A1	In	SD	ZE	78	R	For 18 months, then PD
Thyroid (anaplastic)	Thyroid (anaplastic) Immunogenic tumor (PD-L1 positive in 80% of cancer cells, abundant TILs, 412 SNVs, and 112 LoH, suggesting genetic instability)	Checkpoint inhibition with pembrolizumab	A3	ЭO	PR	86	8	19.6	PR at month 2 and ongoing since 22 months. Normalization of thyroglobulin at month 5 and ongoing. Discontinuation of pembrolizumab at month 8
Thyroid (anaplastic)	Thyroid (anaplastic) Immunogenic tumor (PD-L1 positive in 60% of cancer cells, 1,234 SNVs, and 16 LoH, suggesting genetic instability)	Combination of lenvatinib and pembrolizumab	B3	ĴЮ	PR	96	16	0.9	Since 22 months and ongoing
Thyroid (anaplastic)	Immunogenic tumor (PD-L1 positive in 50% of cancer cells, moderate infiltration of TILs, high mutational burden, with 99 SNVs and 12 LoH), SNVs and CNVs indicating enrichment in MAPK-PI3K signaling	Combination of lenvatinib and pembrolizumab	B3	ЭO	PR	70	9	11.7	For 16 months, then PD
Thyroid (papillary)	Immunogenic tumor (PD-L1 positive in 40% of cancer cells, abundant TILs); results of Keynore-012 trial	Checkpoint inhibition with pembrolizumab	C3	ЭЭO	PR	25	8	3.1	PR for 5 months, then PD of the primary tumor

uable. Listed are the diagnostic results that constituted the rationale for treatment recommendation, the board recommendation including level of evidence (EL), in- or off-label status (L), treatment response (R), PFS of the NOTE. Twenty-one of 198 patients received the recommended therapy and showed at least SD (19 patients) and/or significant CI (one patient). One patient was treated pre-emptively; therefore, the response was not evalrecommended treatment (PFS2; PFS1 if first line), PFS of the previous systemic treatment (PFS1), PFS ratio (PFS1, PFS2/PFS1), and outcome.

1; PFS, progression-free survival; PI3K, phosphatidylinositide 3-kinase; PNET, primitive neuroectodermal tumor; PR, partial response; SCLC, small-cell lung cancer; SD, stable disease; SNV, single-nucleotide variant; TIL, NAB-paclitaxel, nanoparticle albumin-bound paclitaxel; NBCCS, nevoid basal cell carcinoma syndrome; NE, not evaluable; NSCLC, non-small-cell lung cancer; PD, progressive disease; PD-L1, programmed death-ligand eucovorin, and irinotecan; GBM, glioblastoma; GIST, GI stromal tumor; LOH, loss of heteroxygosity; MAPK, mitogen-activated protein kinase; MSI, microsatellite instability; mTOR, mammalian target of rapamycin; Abbreviations: ALL, acute lymphoblastic leukemia; ANG2, angiopoietin-2; CI, clinical improvement; CNV, copy number variation; CRC, colorectal cancer; CUP, cancer of unknown primary; FOLFIRI, fluorouracil, umor-infiltrating lymphocyte; VEGF, vascular endothelial growth factor.

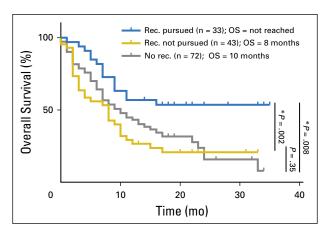


Fig 3. Survival analysis. The Kaplan-Meier curve shows the survival of the following three subgroups of patients with stage IV malignancies (n = 148): patients who implemented the treatment recommendation (Rec. pursued, n = 33), patients who did not implement the treatment recommendation (Rec. not pursued, n = 43; of note: patients who did not receive the recommended therapy because of death before treatment initiation [n = 12] were excluded from analysis), and patients who did not receive a treatment recommendation (n = 72). The curve comparison with the log-rank (Mantel-Cox) test revealed statistical significant differences as shown on graph. OS, overall survival. (*) P < .01.

therapies resulting in a DCR of 7.1% (14 of 198 patients). Other MTB case series reported DCRs in 3.2%, 7.8%, 9%, and 23.3% of the patients,³⁸⁻⁴¹ suggesting that approximately 10% of patients might benefit from advanced personalized decision making.

Although molecular heterogeneity will limit the effect of therapeutic kinase inhibitors, higher nonsynonymous mutational burden can create more neoantigens and therefore improve response rates to CPI.42,43 In our series, eight of 11 patients (72.7%) showing PR received CPI, including seven off-label uses. Predictive biomarkers for individualized immunotherapies are emerging and changing rapidly, with strong differences between entities.44 Here, we used IHC for programmed cell death protein 1 (PD-1)/ PD-L1, tumor-infiltrating lymphocytes, microsatellite instability testing, and mutational burden assessment as predictive biomarkers. In the near future, identifying individual cancer neoantigens might allow a more precise prediction of responses to immunotherapies.⁴⁵ This highlights the importance of an interdisciplinary MTB team that analyzes and interprets biomarkers to identify patients who might benefit from offlabel immuno-oncology treatments.

In an MTB workflow, the portfolio of molecular diagnostic tests, as well as criteria to match and prioritize targeted therapies to molecular biomarkers, affects the probability to identify patients with actionable targets. Here, we used customized molecular diagnostics, including IHC/ISH and tNGS, in 153 out of 198 patients (77.3%) We implemented WES or RNA-Seg analyses for patients with carcinomas of unknown primary and rare cancers and with diseases in which routine molecular diagnostics

did not reveal any actionable target (18.2% of patients).

Multidimensional data have not been implemented successfully to clinical routine, partly because of the complexity of developing and evaluating mathematical predictive models. 46,47 A recent analysis showed that an MTB workflow including WES/whole-genome sequencing, RNA-Seq, and data interpretation by a multidisciplinary board required a turnaround time of 6 weeks.48 Using high-dimensional molecular data, the Molecular Screening for Cancer Treatment Optimization (MOSCATO-01) trial reported actionable mutations in less than half of the patients with advanced solid tumors, 49 and in the National Cancer Institute Molecular Analysis for Therapy Choice (NCI-MATCH) trial, only 9% of the patients could be assigned to one of the prespecified treatment arms.⁵⁰ In contrast, our approach of customized molecular diagnostic testing with restricted use of extended genetic analyses (WES, RNA-Seq) allows a faster turnover with comparable rates of genetically matched treatment recommendations. Therefore, average costs per case can be reduced at least by half when compared with performing extended molecular analysis for each patient. We identified actionable targets in 52.5% of cases and provided treatment recommendations with a median turnaround time of 28 days. To improve standardization and turnaround time, we recently implemented SOPs for diagnostic workups (Data Supplement). Our approach shares similarities with Memorial Sloan Kettering-Integrated Mutation Profiling of Actionable Cancer Targets (MSK-IMPACT), focusing on therapeutically targetable biomarkers for fast clinical decision making and referral of patients to available clinical trials.⁵¹

Targeted drug combinations might offer better DCR over single-agent therapies. 52-55 In part, this is due to crosstalk between signaling pathways as well as spatial and temporal clonal heterogeneity, especially in patients with advanced cancer who failed standard-of-care treatment. 56,57 Most current programs for precision oncology use prespecified, genetically matched, single-agent treatments (NCI-MATCH, ClinicalTrials.gov identifier: NCT02465060; or Targeted Agent and Profiling Utilization Registry [TAPUR], ClinicalTrials.gov identifier: NCT02693535). In our series, three of 19 treatments that successfully controlled disease (15.8%) included molecular combination treatments (Fig 2). These patients did not suffer from grade 3 to 4 adverse effects, although treatment combinations may bear a higher risk of toxicity.⁵⁸

Earlier referral to an MTB (eg, after failure of first-line treatment) might prevent the institution of ineffective treatments, improve the implementation rate, and increase the likelihood of success of molecular biomarker-matched treatments. In our series, patient death, patient preference, or medical reasons precluded implementation in 23.3% of diagnostic and 68.3% of treatment recommendations. The survival analysis revealed a significant overall survival advantage for patients with implemented MTB treatment recommendations (median overall survival not reached; 95% CI, 9 months to not reached) compared with patients where recommendations were not pursued (8 months; 95% CI, 3 to 10 months; P = .002) as well as for patients without treatment recommendation (10 months; 95% CI, 7 to 17 months; P = .008). Because of the low sample size and the heterogeneous composition of patients in the cohorts, the validity of this survival analysis is limited.

Access to molecular biomarker–matched, offlabel agents for cancer treatment is limited. In a recent single-center study, only 5% of molecular biomarker–matched treatment recommendations were implemented, mainly because of limited access to clinical trials or to restricted use of drugs outside their marketed label.⁵⁹ Thus, it is crucial to build up platforms for patients and treating physicians to link individual molecular information of the tumor to appropriate nonapproved drugs and available clinical trials. To this end, MTB networks might implement SOPs for diagnostic work-ups and data interpretation and build alliances to governmental institutions and insurance companies to generate criteria for the financial coverage of molecular analyses and off-label treatments. Finally, an MTB is predestined to generate knowledge and evidence in oncology via single-person trials instead of large, time- and cost-intensive clinical trials. In case of sequence variants with undetermined significance, precision oncology workflows should allow fast reverse translation of sequence variants into informative preclinical models. In a patient with melanoma, we identified a kinase-inactivating BRAF mutation (Data Supplement). In vitro characterization indicated antitumor activity of combined pan-RAF and mitogen-activated protein kinase kinase inhibition and guided successful treatment with sorafenib and trametinib. In rare entities, and especially in the setting of treatment-refractory cancers, precision oncology networks should allow hypothesis-driven in vitro studies and validation in small sets of individuals. Thus, within the concept of patient-centric, biomarker-driven trial designs,60 an MTB might constitute a critical tool to identify informative patients for clinical trials of targeted therapies in rare molecular subgroups.

In summary, this MTB experience illustrates that patient management, on the basis of individual molecular biomarker profiling and analysis, is feasible in patients beyond standard-of-care treatment. We show a high proportion of trial-and off-label treatment recommendations (86.5%) and a DCR for off-label treatments of 7.1%. In cases where no approved treatment is available, an MTB might allow molecular biomarker—matched off-label use of approved drugs across entity barriers or alternatively facilitate the access to therapeutic basket trials.

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REFERENCES

- 1. Chapman PB, Hauschild A, Robert C, et al: Improved survival with vemurafenib in melanoma with BRAF V600E mutation. N Engl J Med 364:2507-2516, 2011
- 2. Hoadley KA, Yau C, Wolf DM, et al: Multiplatform analysis of 12 cancer types reveals molecular classification within and across tissues of origin. Cell 158:929-944, 2014
- 3. Hyman DM, Puzanov I, Subbiah V, et al: Vemurafenib in multiple nonmelanoma cancers with BRAF V600 mutations. N Engl J Med 373:726-736, 2015
- 4. Borghaei H, Paz-Ares L, Horn L, et al: Nivolumab versus docetaxel in advanced nonsquamous non-small-cell lung cancer. N Engl J Med 373:1627-1639, 2015
- 5. Prahallad A, Sun C, Huang S, et al: Unresponsiveness of colon cancer to BRAF(V600E) inhibition through feedback activation of EGFR. Nature 483:100-103, 2012
- Hong DS, Morris VK, El Osta B, et al: Phase IB study of vemurafenib in combination with irinotecan and cetuximab in patients with metastatic colorectal cancer with BRAFV600E mutation. Cancer Discov 6:1352-1365, 2016
- 7. Stones CJ, Kim JE, Joseph WR, et al: Comparison of responses of human melanoma cell lines to MEK and BRAF inhibitors. Front Genet 4:66, 2013
- 8. Lawrence MS, Stojanov P, Polak P, et al: Mutational heterogeneity in cancer and the search for new cancer-associated genes. Nature 499:214-218, 2013

- 9. Gerlinger M, Rowan AJ, Horswell S, et al: Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. N Engl J Med 366:883-892, 2012
- 10. Xu X, Hou Y, Yin X, et al: Single-cell exome sequencing reveals single-nucleotide mutation characteristics of a kidney tumor. Cell 148:886-895, 2012
- 11. Siravegna G, Mussolin B, Buscarino M, et al: Clonal evolution and resistance to EGFR blockade in the blood of colorectal cancer patients. Nat Med 21:795-801, 2015 [Erratum: Nat Med 21:827, 2015]
- 12. Kobayashi S, Boggon TJ, Dayaram T, et al: EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. N Engl J Med 352:786-792, 2005
- 13. Goss G, Tsai CM, Shepherd FA, et al: Osimertinib for pretreated EGFR Thr790Met-positive advanced non-small-cell lung cancer (AURA2): A multicentre, open-label, single-arm, phase 2 study. Lancet Oncol 17:1643-1652, 2016
- 14. Li MM, Datto M, Duncavage EJ, et al: Standards and guidelines for the interpretation and reporting of sequence variants in cancer: A joint consensus recommendation of the Association for Molecular Pathology, American Society of Clinical Oncology, and College of American Pathologists. J Mol Diagn 19:4-23, 2017
- 15. Hirsch B, Endris V, Lassmann S, et al: Multicenter validation of cancer gene panel-based nextgeneration sequencing for translational research and molecular diagnostics. Virchows Arch 472:557-565, 2018
- 16. Geißler AL, Geißler M, Kottmann D, et al: ATM mutations and E-cadherin expression define sensitivity to EGFR-targeted therapy in colorectal cancer. Oncotarget 8:17164-17190, 2017
- 17. Kovaleva V, Geissler AL, Lutz L, et al: Spatio-temporal mutation profiles of case-matched colorectal carcinomas and their metastases reveal unique de novo mutations in metachronous lung metastases by targeted next generation sequencing. Mol Cancer 15:63, 2016
- 18. Lek M, Karczewski KJ, Minikel EV, et al: Analysis of protein-coding genetic variation in 60,706 humans. Nature 536:285-291, 2016
- 19. Landrum MJ, Lee JM, Riley GR, et al: ClinVar: Public archive of relationships among sequence variation and human phenotype. Nucleic Acids Res 42:D980-D985, 2014
- 20. Forbes SA, Beare D, Gunasekaran P, et al: COSMIC: Exploring the world's knowledge of somatic mutations in human cancer. Nucleic Acids Res 43:D805-D811, 2015
- 21. Sherry ST, Ward M, Sirotkin K: dbSNP-database for single nucleotide polymorphisms and other classes of minor genetic variation. Genome Res 9:677-679, 1999
- 22. Sherry ST, Ward MH, Kholodov M, et al: dbSNP: The NCBI database of genetic variation. Nucleic Acids Res 29:308-311, 2001
- 23. Chang MT, Asthana S, Gao SP, et al: Identifying recurrent mutations in cancer reveals widespread lineage diversity and mutational specificity. Nat Biotechnol 34:155-163, 2016
- 24. Chang MT, Bhattarai TS, Schram AM, et al: Accelerating discovery of functional mutant alleles in cancer. Cancer Discov 8:174-183, 2018
- 25. Cotto KC, Wagner AH, Feng YY, et al: DGIdb 3.0: A redesign and expansion of the drug-gene interaction database. Nucleic Acids Res 46:D1068-D1073, 2017
- 26. Shihab HA, Gough J, Cooper DN, et al: Predicting the functional, molecular, and phenotypic consequences of amino acid substitutions using hidden Markov models. Hum Mutat 34:57-65, 2013
- 27. Reva B, Antipin Y, Sander C: Predicting the functional impact of protein mutations: Application to cancer genomics. Nucleic Acids Res 39:e118, 2011
- 28. González-Pérez A, López-Bigas N: Improving the assessment of the outcome of nonsynonymous SNVs with a consensus deleteriousness score, Condel. Am J Hum Genet 88:440-449, 2011

- 29. Chen Y, Cunningham F, Rios D, et al: Ensembl variation resources. BMC Genomics 11:293, 2010
- 30. Adzhubei IA, Schmidt S, Peshkin L, et al: A method and server for predicting damaging missense mutations. Nat Methods 7:248-249, 2010
- 31. Kumar P, Henikoff S, Ng PC: Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. Nat Protoc 4:1073-1081, 2009
- 32. Boeva V, Zinovyev A, Bleakley K, et al: Control-free calling of copy number alterations in deep-sequencing data using GC-content normalization. Bioinformatics 27:268-269, 2011
- 33. Dobin A, Davis CA, Schlesinger F, et al: STAR: Ultrafast universal RNA-seq aligner. Bioinformatics 29:15-21, 2013
- 34. Ritchie ME, Phipson B, Wu D, et al: limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res 43:e47, 2015
- 35. Law CW, Chen Y, Shi W, et al: voom: Precision weights unlock linear model analysis tools for RNA-seq read counts. Genome Biol 15:R29, 2014
- 36. Von Hoff DD, Stephenson JJ Jr, Rosen P, et al: Pilot study using molecular profiling of patients' tumors to find potential targets and select treatments for their refractory cancers. J Clin Oncol 28:4877-4883, 2010
- 37. Traer E, MacKenzie R, Snead J, et al: Blockade of JAK2-mediated extrinsic survival signals restores sensitivity of CML cells to ABL inhibitors. Leukemia 26:1140–1143, 2012
- Larkin J, Ascierto PA, Dréno B, et al: Combined vemurafenib and cobimetinib in BRAF-mutated melanoma. N Engl J Med 371:1867-76, 2014
- 39. Sohal DP, Rini BI, Khorana AA, et al: Prospective clinical study of precision oncology in solid tumors. J Natl Cancer Inst 108:djv332, 2015
- 40. Johnson DB, Dahlman KH, Knol J, et al: Enabling a genetically informed approach to cancer medicine: A retrospective evaluation of the impact of comprehensive tumor profiling using a targeted next-generation sequencing panel. Oncologist 19:616-622, 2014
- Dalton WB, Forde PM, Kang H, et al: Personalized medicine in the oncology clinic: Implementation and outcomes of the Johns Hopkins Molecular Tumor Board. JCO Precision Oncol: 10.1200/PO.16.00046, 2017
- 42. Rizvi NA, Hellmann MD, Snyder A, et al: Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science 348:124-128, 2015
- 43. McGranahan N, Furness AJ, Rosenthal R, et al: Clonal neoantigens elicit T cell immunoreactivity and sensitivity to immune checkpoint blockade. Science 351:1463-1469, 2016
- 44. Gibney GT, Weiner LM, Atkins MB: Predictive biomarkers for checkpoint inhibitor-based immunotherapy. Lancet Oncol 17:e542-e551, 2016
- 45. Gfeller D, Bassani-Sternberg M, Schmidt J, et al: Current tools for predicting cancer-specific T cell immunity. OncoImmunology 5:e1177691, 2016
- 46. McShane LM, Cavenagh MM, Lively TG, et al: Criteria for the use of omics-based predictors in clinical trials. Nature 502:317-320, 2013
- 47. Singer J, Irmisch A, Ruscheweyh HJ, et al: Bioinformatics for precision oncology. Brief Bioinform: bbx143, 2017
- 48. Horak P, Klink B, Heining C, et al: Precision oncology based on omics data: The NCT Heidelberg experience. Int J Cancer 141:877-886, 2017
- 49. Massard C, Michiels S, Ferté C, et al: High-throughput genomics and clinical outcome in hard-to-treat advanced cancers: Results of the MOSCATO 01 trial. Cancer Discov 7:586-595, 2017
- Conley BA, Gray R, Chen A, et al: NCI-molecular analysis for therapy choice (NCI-MATCH) clinical trial: interim analysis. Presented at American Association for Cancer Research 107th Annual Meeting 2016, New Orleans, LA, April 16-20, 2016

- 51. Cheng DT, Mitchell TN, Zehir A, et al: Memorial Sloan Kettering-Integrated Mutation Profiling of Actionable Cancer Targets (MSK-IMPACT): A hybridization capture-based next-generation sequencing clinical assay for solid tumor molecular oncology. J Mol Diagn 17:251-264, 2015
- 52. Tannock IF, Hickman JA: Limits to personalized cancer medicine. N Engl J Med 375:1289-1294,
- 53. Yap TA, Omlin A, de Bono JS: Development of therapeutic combinations targeting major cancer signaling pathways. J Clin Oncol 31:1592-1605, 2013
- 54. Liu S, Nikanjam M, Kurzrock R: Dosing de novo combinations of two targeted drugs: Towards a customized precision medicine approach to advanced cancers. Oncotarget 7:11310-11320, 2016
- 55. Turner NC, Reis-Filho JS: Genetic heterogeneity and cancer drug resistance. Lancet Oncol 13:e178-e185, 2012
- 56. Swanton C, Govindan R: Clinical implications of genomic discoveries in lung cancer. N Engl J Med 374:1864-1873, 2016
- 57. Shi H, Hugo W, Kong X, et al: Acquired resistance and clonal evolution in melanoma during BRAF inhibitor therapy. Cancer Discov 4:80-93, 2014
- 58. Soria JC, Massard C, Izzedine H: From theoretical synergy to clinical supra-additive toxicity. J Clin Oncol 27:1359-1361, 2009
- 59. Beltran H, Eng K, Mosquera JM, et al: Whole-exome sequencing of metastatic cancer and biomarkers of treatment response. JAMA Oncol 1:466-474, 2015
- 60. Biankin AV, Piantadosi S, Hollingsworth SJ: Patient-centric trials for therapeutic development in precision oncology. Nature 526:361-370, 2015

Appendix

Fig A1. Molecular Tumor Board (MTB) workflow. TOS, Tumorboard Online System.

