



# How molecular imaging will enable robotic precision surgery: the role of artificial intelligence, augmented reality, and navigation

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#### **REVIEW ARTICLE**



# How molecular imaging will enable robotic precision surgery

The role of artificial intelligence, augmented reality, and navigation

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#### **Abstract**

Molecular imaging is one of the pillars of precision surgery. Its applications range from early diagnostics to therapy planning, execution, and the accurate assessment of outcomes. In particular, molecular imaging solutions are in high demand in minimally invasive surgical strategies, such as the substantially increasing field of robotic surgery. This review aims at connecting the molecular imaging and nuclear medicine community to the rapidly expanding armory of surgical medical devices. Such devices entail technologies ranging from artificial intelligence and computer-aided visualization technologies (software) to innovative molecular imaging modalities and surgical navigation (hardware). We discuss technologies based on their role at different steps of the surgical workflow, i.e., from surgical decision and planning, over to target localization and excision guidance, all the way to (back table) surgical verification. This provides a glimpse of how innovations from the technology fields can realize an exciting future for the molecular imaging and surgery communities.

**Keywords** Robotic surgery · Molecular imaging · Image-guided surgery · Precision surgery · Artificial intelligence · Augmented reality

To optimize patient outcomes and accommodate public demand, surgical approaches are becoming more and more tailored to the individual's needs. This personalization is pursued to the extent that precision surgery is one of the leading trends in current medicine [1]. Not only is there a drive towards increasing the resection accuracy, but there also is an increasing focus on balancing cure and side

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effects, requiring precise (intraoperative) target definition and control. These trends have driven the growth of minimally invasive surgery, particularly the implementation of robotic surgery and various image-guided surgery technologies.

In the past decades, the availability of robotic telemanipulators or robotic master-slave systems has facilitated robot-assisted surgery. In particular, the da Vinci platform (Intuitive Surgical, Sunnyvale (CA), USA), as the prime example, has become the new standard for the management of prostatic cancer (i.e., prostatectomy and lymphatic dissections) [2]. An indication where patients are increasingly being staged based on molecular imaging (i.e., PSMA PET [3]). The first generation of telemanipulator systems has sparked the dissemination of robotic surgery to other indications, e.g., lymphatic dissections [4], partial nephrectomy [5], hysterectomy [6], pulmonary and esophageal surgery [7], hepatobiliary surgery [8], colorectal cancers [9], and even head and neck surgery [10]. With the routine inclusion of a fluorescence laparoscope (since the da Vinci Si model) and the TilePro function that facilitates the multi-input display of preoperative scans (e.g., PET or SPECT) and intraoperative imaging directly within the surgical console,



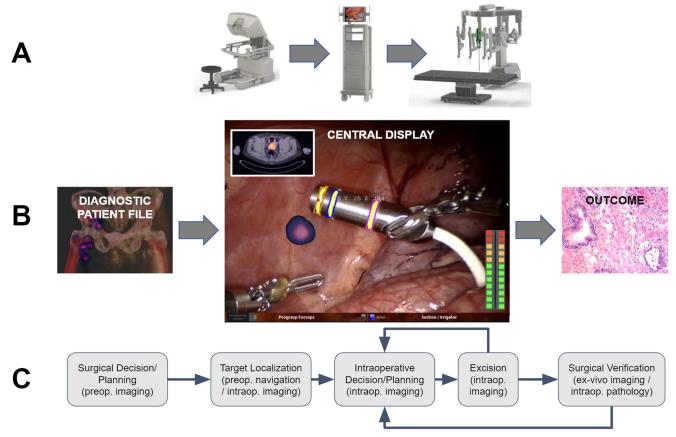


Fig. 1 A Components in a robotic telemanipulator system. The surgeon operates the robot from a console that connects to the robotic arms over a central data processing unit where also the video signal of the laparoscope is processed. B Molecular images, like PET, SPECT, or scintigraphy, and so-called metadata of the patient are fed to the data processing unit. There, intraoperative information is merged and shown in a single central display. There, augmented reality (AR)

image overlays can be shown with the instruments and signals from the surgery (theoretical possibilities indicated). As a result of the procedure, the diseased tissue is removed. This results in an outcome, e.g., the resection borders' status. C A molecular imaging-enhanced robotic surgery can be abstracted as surgical decision/planning, target localization, intraoperative decision/planning, excision, and surgical verification, all of which are interconnected

robotic surgery has well geared up. These upgrades facilitate efficient integration of all the data streams associated with the use of image-guided surgery (Fig. 1).

Beyond the da Vinci system, more robotics platforms are now making their way into clinical care. The Versius (CMR Surgical, Cambridge, UK) and the Senhance surgical system (ransEnterix, Morrisville (NC), USA) follow a similar approach to Intuitive Surgical with a versatile model applicable to several anatomies. However, other surgical robots have entered niche "organ" markets like neurosurgery and orthopedics, e.g., the ROSA brain and knee robots (Zimmer Biomet, Warsaw (IN), USA), the Mazor X spine robot (Medtronic, Dublin, Ireland), the Mako knee surgery assistant (Stryker, Kalamazoo (MI), USA) and the Navio knee robot (Smith & Nephew, Watford, UK). The field also extends into more complex indications like natural orifice total endoscopic surgery, e.g., the Flex robotic system (Medrobotics, Raynham (MA), USA), peripheral lung biopsies, e.g., the Ion (Intuitive Surgical,

Sunnyvale (CA), USA), or the Monarch system (Auris Health, Redwood City (CA), USA), or catheter guidance, e.g., the CorPath GRX (Corindus, Waltham (MA), USA) or the Vdrive system (Stereotaxis, St. Louis (MO), USA). The expansion of robotics will go further in the direction of microrobotics, as needed in eye or brain surgery [11, 12].

Image guidance (and progressively molecular image guidance) is probably one of the most critical levers in realizing precision surgery. Three aspects are strongly contributing to its evolution. First, advances made in radiology and nuclear medicine have made it possible to identify diseased tissue early and with superior accuracy and molecular precision [13]. In particular, molecular imaging plays a significant role in this improvement; Molecular imaging refers to the in vivo imaging of molecule concentrations based on the presence of an endogenous molecule or the injection of an exogenous imaging agent (i.e., tracer Table 1). In vivo detection succeeds utilizing



Table 1 Robotic indications where tracer-based molecular imaging has already clinically demonstrated value in surgical planning, intraoperative guidance, and postoperative control

Indication	Tracer	Preop. imaging	Intraop./ex vivo detection	Reference
Lymphatics				
(Sentinel) Lymph node biopsy	<sup>99m</sup> Tc-radiocolloids	Sz, SPECT/CT	DROP-IN $\gamma$ Counting, ioSz, fhSPECT	[21]
	ICG- <sup>99m</sup> Tc-HSA nanocolloid	Sz, SPECT/CT	DROP-IN γ Counting, ioSz, fhSPECT, FluoLap	[22]
	ICG	-	FluoLap	[23]
	Fluorescein	-	FluoLap	[24]
	Methlyene blue	-	FluoLap	[25]
High blood perfusion				
Tumors (e.g., liver, adrenal glands)	ICG	-	FluoLap	[26]
Vessels (e.g., ureters, anastomosis, vasculature)	ICG	-	FluoLap	[27]
Receptor-targeted				
Prostate cancer	<sup>99m</sup> T c-PSMA I&S	Sz, SPECT/CT	DROP-IN $\gamma$ Counting, ioSz, fhSPECT	[28]
	<sup>68</sup> Ga-PSMA-914	PET/CT, PET/MR	DROP-IN β Counting (experimental), FluoLap, Cerenkov Imaging	[29]
Clear cell renal cell carcinoma (ccRCC)	<sup>111</sup> <i>In-</i> DOTA- girentuximab IRDye800	Sz, SPECT/CT	DROP-IN γ Counting, ioSz, fhSPECT, FluoLap	[30]
	OTL38	-	FluoLap	[31]
Pulmonary nodules	OTL38	-	FluoLap	[32]

io, intraoperative; Sz, scintigraphy; fhSPECT, freehand SPECT; FluoLap, fluorescence laparoscopy

a radioactive, fluorescent, or magnetic label (exogenous tracers, Table 1) or by an optical or electrical signature of the target molecule in a non-contrasted approach [14, 15]. Second, the rise of interventional molecular imaging strategies, in the form of tracers and surgical molecular imaging modalities, increasingly allows surgeons to target and resect tissues based on molecular features [16]. This can be, for example, using radio-guided surgery [17, 18] or fluorescence-guided surgery [19]. Third, in the surgical arena, significant strides have been made concerning image guidance modalities (hardware) and visualization concepts (software) [20].

Since the robotic setting is somewhat different from a traditional operating room setting, many current image guidance modalities have to be specifically adapted. However, since most robotic platforms harbor a plural of highly maneuverable instruments, this does open up many new advantages that could lead to a better performance of the image guidance modalities themselves [33]. Furthermore, since the surgeon is operating behind a video console or "central display" (see Fig. 1A and B), the robotic platform seems to be an ideal system to

integrate new developments in display technology, directly visualizing all kinds of patient information (e.g., [34]). Such patient datasets could entail pre- or intraoperative imaging, surgical planning, navigation, patient monitoring, and (post-resection) lesion confirmation. With the high amounts of computational power available with current-day technology, such datasets' formation is increasingly assisted with artificial intelligence (AI) methods, including machine learning, and in particular deep neural networks. Thus, the establishment of robotic surgery requires the necessary adjustments but provides many exciting opportunities for molecular image-guided surgery.

This review aims at connecting the molecular imaging and nuclear medicine community, which provides a wide range of radiotracers for image-guided surgery, to current hardware and software developments. This entails technologies ranging from artificial intelligence and advanced visualization to robot-tailored imaging modalities. We will review works based on the robotic surgery workflow (see Fig. 1). To provide insight into the direction that the field is moving in, we have mainly focused on technological possibilities that are becoming available or



are expected to provide future impact for robot-assisted surgery.

To guarantee a thorough literature review, here we employed two approaches of research. First, we performed an exhaustive search on the highest impact Molecular Imaging and Medical Robotics journals to identify the most relevant articles and reviews of the field over the last 10 years. Secondly, a systematic and comprehensive keyword-based search was conducted on molecular imaging in a minimally invasive or robotic setup in PubMed, 1 Google Scholar,<sup>2</sup> and Semantic Scholar<sup>3</sup> combining the keywords Molecular Imaging, Nuclear Imaging, Gamma Imaging, Beta Imaging, Fluorescence Imaging with the terms Image-guided surgery, Laparoscopic surgery, Minimally-invasive surgery, and Robotic surgery. All titles and abstracts were scanned manually for relevance, deleting duplicates and selecting the ones that best depicted the techniques searched in the view of the authors, prioritizing in-human trials while only including the most promising non-human trials.

## Preoperative planning and navigation

Preoperative molecular images, like SPECT and PET, nowadays almost always in combination with CT or MRI, are the starting point of many robotic surgery procedures. They serve as means to select patients, plan entry paths, trocar placement, and then even, in some cases, guide the surgeon grossly to the area of the target.

# Preoperative molecular imaging for surgical selection, planning, and prediction

The selection of patients for surgery has been a task of the surgeon or a multidisciplinary group of experts. Such a process often involves considering the patient metadata (health state, age, occupation, family history), available know-how, and infrastructure, as well as imaging material to assess if surgery is the most recommendable option. The surgical risk can also be estimated by looking at organs at risk close to the surgical site and potential access paths.

Routine lymphatic mapping has made treating surgeons aware of the role that preoperative imaging plays in image guidance. Moreover, they have shown that the type of preoperative imaging defines the value created in the OR [35]. For example, complementing 2D scintigraphic images with 3D SPECT/CT increases the number of identified sentinel nodes. Equally important, the SPECT-registered

<sup>&</sup>lt;sup>3</sup>https://www.semanticscholar.org/



(low dose) CT-based anatomical image (also in 3D) helps provide anatomical context, visualize specific surgical landmarks, and locate organs at risk. This allows surgeons to plan their surgical procedure by pinpointing the area of interest more accurately and make a risk estimation concerning the induction of damage to vital structures.

Artificial intelligence for segmentation In the whole preoperative route of patient selection, surgical planning, and prediction, AI methods are starting to play an increasingly relevant role, assisting the physicians in delivering their best performance. So far, such AI methods have mostly focussed on the aspect of surgical planning, providing an automatic segmentation of tumors and organs at risk using both patient imaging and metadata. For such segmentation models, deep learning methods have predominantly taken over, often based on convolutional neural networks (CNNs). For image processing, such CNNs have been shown to overperform humans in classification and detection methods (e.g., [36, 37]). They are increasingly being used for tasks like multi-organ segmentation [38]. When dealing with imaging data, a CNN consists of stacked nonlinear convolutional layers that can extract features of high-level (shapes, content) and low-level (details, texture) from images. The network architectures reduce dimensionality first to extract highlevel features in a so-called bottleneck representation of the image (see Fig. 2). From there, these features are further processed, where so-called skip connections contribute lowlevel information from early layers. In the end, the final layers convert the features into probabilities, for example, in the scenario of organ segmentation. AI organ segmentation can even be further optimized by feeding the neural network with relevant patient metadata, such as patient age, weight, clinical parameters, and previous treatment strategies (e.g., [39]).

To indicate organs at risk, such automatic multi-organ segmentations have been successfully tackled using CT scans [40-42] or MR scans [43], even in case they were cropped [44]. The higher the accuracy of these segmentation models (often expressed as a higher "Dice score"), the lower the need for an expert physician to correct the automatic segmentation. Interestingly, combining such anatomy-based segmentations with molecular imaging (i.e., PET or SPECT) can further improve the AI results. A network having both inputs can learn the physiological uptake of different organs and enhance the segmentation (e.g., improved definition of (metastatic) tumor nodules). First approaches that use both anatomical and molecular images to enhance segmentation have been undertaken with PET/CT in automatic tumor segmentation in lung cancer [45–47], head and neck tumors [48, 49], and non-Hodgkin lymphoma [50]. Here, a combination of the tumor detection task and the multi-organ segmentation can be advantageous

<sup>&</sup>lt;sup>1</sup>https://www.ncbi.nlm.nih.gov

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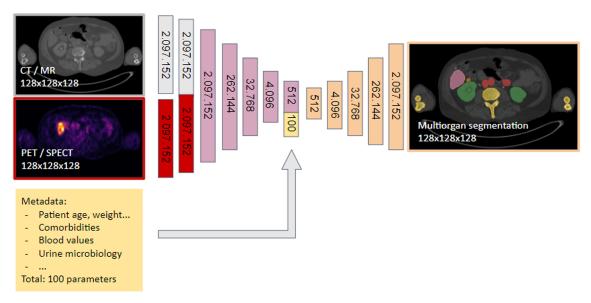


Fig. 2 Example of a convolutional deep neural network (CNN), a standard AI algorithm. CNNs can be applied for multiorgan image segmentation using molecular imaging and metadata. Here, the CNN reads anatomical images (CT or MR, gray) and molecular images (PET/SPECT/scintigraphy, red). After initial processing, it concatenates their features (pink). The network then reduces the input's dimensionality (i.e., brings the images from a size of, e.g.,  $128 \times 128 \times 128 = 2,097,152$  to only values 512 representing both of

them). These 512 parameters get further concatenated with the 100 metadata parameters (yellow) fed into the network's bottleneck. The 512+100=612 parameters contain a compressed high-level representation of the image and patient information. Based on the image and metadata, the network then solves the target tasks (here organ segmentation, orange) while increasing dimensionality (i.e., upscaling from 612 parameters back to  $128 \times 128 \times 128 = 2,097,152$ )

as this mimics both nuclear medicine physicians and radiologists [51].

Artificial intelligence for outcome prediction Next to assistance in surgical planning, AI methods such as deep learning might improve initial patient selection and even enable predicting treatment outcomes. The latter could entail the detection of abnormal anatomy during surgery, the occurrence of side effects, the duration of the procedure, or the length of the postoperative stay. Initial works have been published for the treatment of esophageal [52] and oropharyngeal cancer [53] using PET images as input. Despite the prediction being here the response to chemotherapy or radiotherapy, technically, the same approach can be used to train a system for surgical prediction by changing the training dataset to include surgical outcome information.

# Preoperative molecular imaging as intraoperative roadmaps for target localization

Preoperative imaging techniques are used routinely to create detailed roadmaps, based on which a surgeon can plan and execute its resection. Initially, surgeons study the diagnostic images together with their nuclear medicine colleagues. Based on these discussions, the surgeon would then create a mental roadmap of the procedure. While expert surgeons are surprisingly effective in doing such translations, this

approach's failure rate is relatively high (e.g., [54–57]). This has been the motivation behind initiatives that physically bring preoperative imaging information into the surgical theatre. We do not refer to the old-fashioned printout of, e.g., a 2D scintigraphic image, but rather the use of fully annotated digital images displayed on a dedicated dashboard in the OR. Such displays have been used since the beginning of medical imaging and, mainly, since PACS's introduction in the 1990s [58]. The increasing complexity of 3D scans such as SPECT/CT or PET/CT has driven a growth in the use of such display options. The TilePro extension of the da Vinci surgical console now even makes it possible to directly input preoperative images (or other datasets) as windows in the surgeon's display [59, 60]. Despite the availability of such displays, it remains challenging to translate such preoperative imaging information to successful surgical execution. This happens since the patient is drastically repositioned, covered by the surgical robot, insufflated, and visualized through the robotic laparoscope's vision.

The direct integration of the preoperative images into the surgeon's laparoscopic view is a promising development field. This can be enabled with either an augmented reality (AR) overlay on the laparoscopic video feed or a virtual reality (VR) visualization. Both allow navigating towards tissue targets as marked in the preoperative images [61]. For these approaches to work, it is instrumental



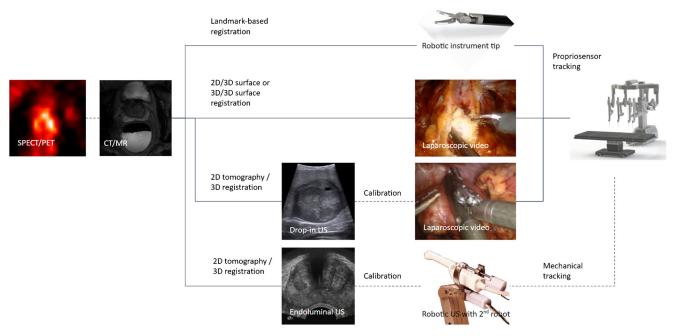


Fig. 3 Possible registration options to bring molecular images (SPECT/PET or MRI) over anatomical images (CT/MR) to the robot's coordinate system

that the preoperative images are geometrically registered to the patient's position and orientation (pose) in the operating room. In robotic surgery, such registration relies on tracking solutions, such as optical or mechanical tracking systems calibrated to the robot, or the robotic arms' propriosensory information, to define the patient's relative pose, laparoscopic instruments, and camera (Fig. 3). Several registration approaches can be followed to bring the preoperative images into the tracking system's coordinate system.

Landmark-based registration This is the most intuitive and straightforward option already proposed at the end of the 1990s [62]. First, particular landmarks are selected on the preoperative images and subsequently tipped with a calibrated tracked instrument in situ. The resulting point correspondences between preoperative images and tracking coordinates of the anatomical landmarks can be processed to yield the desired registration. The more anatomic landmarks used, the better the quality of registration. In practical terms, this is challenging and, unfortunately, often results in poor registrations. On the other hand, in robotic surgery, the robotic instruments are tracked using the propriosensory and can be used to tip the landmarks making an external tracking system unnecessary.

Laparoscopic video-based registration Several groups have proposed methods for registering the laparoscopic video with renderings of the preoperatively acquired images [63, 64], either using a 2D or 3D laparoscope. Such

approaches rely on the fact that preoperative CT or MRI depicts the anatomy at the time of surgery reasonably well (i.e., no significant resections or strong tissue deformations have taken place). Besides, the lack of clearly distinguishable landmarks in the laparoscopic images makes this registration approach challenging. However, since the robot's propriosensors track the laparoscope's pose, no additional tracking hardware is required here. Also, with the continuous development of AI-supported surgical scene recognition, it is expected that there is a lot to be won in the accuracy of this method (e.g., [65, 66]).

Intraoperative ultrasound-based registration While registering organ surfaces to preoperative imaging is feasible using the laparoscopic video feed, deep-lying structures may not be properly registered. In a robotic setup, this could be achieved with a positionally tracked ultrasound (US) probe. More specifically, the probe can be either endoluminal ultrasound (i.e., transrectal or transvaginal ultrasound, tracked via mechanics or a second robot [67]) or DROP-IN ultrasound (tracked via laparoscopic-video [68]). The most important advantage is the possibility of using indepth information to improve registration results. However, it is limited to specific applications and organs where US is feasible (such as the prostate, kidney, liver).

From the methods mentioned above, we believe that evolutions of laparoscopic video-based registration, and to a lesser extent, ultrasound registration, combined with mechanical tracking of the robotic platform, will be the method of choice in robotic surgery in the years to come.



This is because they can be integrated into the robotic suite and thus require less external hardware.

With the proper registrations in place, surgical navigation has the potential to translate the wealth of preoperative information (e.g., multi-modal scans, target definition, most efficient route, critical organ segmentations) to the operating room, directly visualized in the surgical robot console. Using molecular imaging modalities (i.e., PET and SPECT), this has primarily been oncological data, providing guidance towards, e.g., (metastatic) lymph nodes defined before surgery [18].

Unfortunately, the use of preoperative images for surgical navigation is challenging in a soft tissue environment, where organs deform and where surgery itself changes the anatomy dramatically [69]. Efforts have been made to cope with these inaccuracies by regularly reapplying registration of the preoperative images to the current surgical situation using C-arms and ultrasound (e.g., [70, 71]), as well as tracked gamma probes [72] or gamma cameras [73].

Alternatively, to cope with soft tissue–induced deformation, it has also been proposed to navigate intraoperative molecular detection modalities, basically using them as pointers within the navigation workflow. In this way, navigation is useful for orientation and rough localization in the patient. Simultaneously, the real-time feedback of the molecular detection modality allows for correction and confirmation of the actual tissue targets once close enough. This concept has been shown in vivo using gamma probes (e.g., [74]) and fluorescence cameras [75–77], and preclinically even using robotic DROP-IN gamma probes [78] and robotic fluorescence cameras [33]. In our view, with the

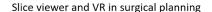
current status of this field, intraoperative imaging (e.g., ultrasound, freehand SPECT, fluorescence laparoscopy) remains indispensable in soft tissue environments.

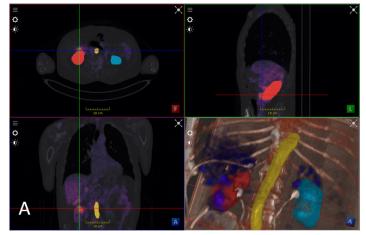
# Advanced visualization and navigation strategies for preoperative molecular images

Before going to intraoperative molecular imaging approaches, it is relevant to discuss visualization techniques in more detail. To advance the integration between preoperative imaging and the surgical experience once the registration is solved, advanced medical image visualization strategies have been put forward.

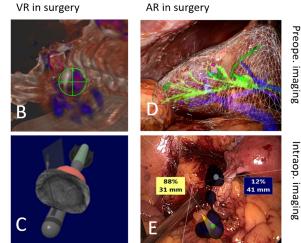
Virtual reality Since the 2000s, it has been possible to stream a digital full image viewer showing either 2D images or 3D images in three planes, often visualized as a 3D render in a virtual environment (i.e., virtual reality (VR); e.g., [79]) where a priori segmented organs and structures can be highlighted (Fig. 4A). This applies not only to preoperative images but also to intraoperative information, such as the read-out of intraoperative ultrasound (US), displayed in relation to the surgical instruments [67], giving the surgeon a virtual context during surgery (Fig. 4B).

Augmented reality The next relevant improvement in this field came with the introduction of augmented reality (AR) into the operating room. Unlike the virtual environment provided with VR, AR is defined as an approach of visualization where "invisible" information (e.g., pre- or intraoperatively obtained imaging findings) are overlaid on





**Fig. 4** Different VR/AR visualization options applicable for robotic surgery. **A** Visualization of PET/CT in 3 planes (axial, coronal, and sagittal) and 3D render, including an overlay of segmented organs. **B** VR view of PET/CT image as used for guidance on PSMA-guided surgery. **C** VR view of intraoperative TRUS in the context of the



TRUS probe and the robot instruments. Image courtesy of Tim Salcudean, UBC, Canada. **D** Segmented organs overlaid as AR patients' body for port placement planning [80]. **E** AR visualization of free-hand SPECT images showing sentinel lymph nodes in an endometrium cancer surgery



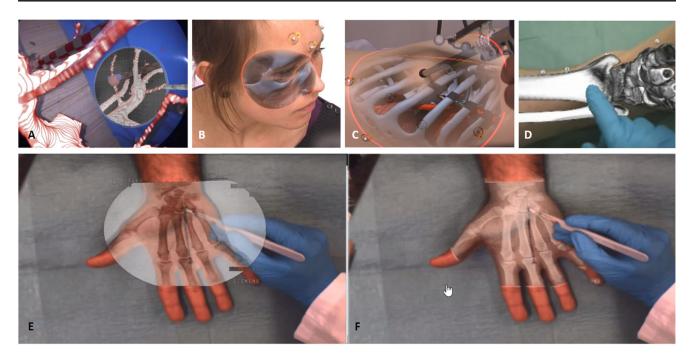


Fig. 5 Depth perception improvement methods for AR: A virtual mirror, B curvature-dependent transparency, C virtual shadows, D object-subtraction; E, F example of a new paradigm of AR visualization where only relevant information is overlaid on the real image versus standard AR

the view of the real environment, providing immediate context with the actual patient (Fig. 4D and E). This concept was introduced in orthopedic and brain surgery as early as the 1990s [81] using CT/MRI and even in combination with microscopic imaging [82]. However, it took until 2000 for AR to make it to clinical applications [61].

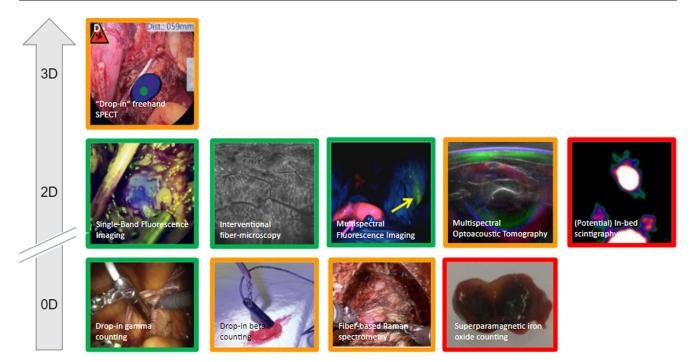
A key challenge in AR is achieving a visualization that provides additional information without diminishing the camera's information or giving a wrong perception. This limitation of AR has strongly limited its adoption. Different approaches have been proposed to improve depth perception (Fig. 5A–D), like the use of virtual mirrors [83], curvature-dependent transparency [84], virtual shadows [85], or object subtraction [86]. With the advances in machine learning, in particular, object subtraction has shown excellent performance for the detection of instruments in laparoscopy videos [87], opening a path towards visualization approaches where only the relevant information is overlaid to the surgeon [88] (Fig. 5E and F).

# Intraoperative planning, decision, and excision assistance

In addition to the planning and guidance delivered with preoperative scans, intraoperative imaging plays an important role in intraoperative lesion localization, decision making, and subsequent confirmation. Intraoperative imaging was first introduced in the context of X-rays at the beginning of the twentieth century and has since then evolved to provide a direct anatomical context within the operating room using, for example, a C-arm during orthopedic surgery [89]. Other forms of intraoperative anatomical imaging included the often used intraoperative US (e.g., to evaluate the extent of a tumor lesion during liver surgery [90]), or even intraoperative MRI (e.g., surgical management of glioblastoma [91]). However, molecular imaging has also played an essential role in image-guided surgery, especially using radio-guided surgery. Current intraoperative molecular imaging approaches mainly focus on radioactive, fluorescent, magnetic, or hybrid tracers, but also non-contrasted approaches are flourishing like multispectral optoacoustic tomography (MSOT; [92]), fiber-based microscopy [93], Raman spectrometry [94], among others (Fig. 6).

Before diving into the different intraoperative detection and imaging modalities, it is worth mentioning that navigation and advanced visualization concepts for preoperative images can be applied similarly using intraoperative molecular images. Techniques like freehand SPECT already included AR and pointer navigation means from its first publications [95–97]. The advantage of intraoperative imaging is that updated images, even after lesion removal, can be used to (partially) cope with tissue deformation during navigation [69]. Combined with real-time feedback from an intraoperative modality (e.g., radio- or fluorescence guidance) further confirms successful lesion localization [77].





**Fig. 6** Non-exhaustive overview of some current and possible future technologies for robotic intraoperative molecular imaging separated by their dimensions and development status (green, commercially available; orange, research prototypes available; red, potential

developments). Non-imaging devices are defined as zero-dimensional as they are a single pixel detector and not a line detector which would be one-dimensional

# Radio-guided surgery

Being routinely applied for over several decades, radio-guided surgery is one of the most used types of (molecular) image-guided surgery [17, 18]. RGS focuses on the intraoperative detection and imaging of lesions or processes targeted with radiopharmaceuticals. The available detection modalities help realize in situ localization and provide an intraoperative control of successful resection. Being applied for a great amount of open and laparoscopic indications, the most used routine application has been the SLNB procedure (see Table 1). That said, many of the modalities developed to facilitate SLNB readily translate to, for example, receptor-targeted procedures. In receptor-targeted procedures, the main success story has been the radio-guided PSMA-targeted salvage surgery in prostate cancer, currently already applied in >250 patients [98].

A relevant aspect of RGS is the fact that the patient is radioactive during surgery. This is less of a radiation protection issue for the patient: RGS is only performed if the patient's benefits (i.e., removal of a tumor, minimally invasive lymphatic status, etc.) outweigh the potential risks (i.e., late radiation-induced cancer). However, it is for the surgical staff, who would else not receive any dose. This concern is interestingly less relevant in robotic surgery than in open or laparoscopic surgery, as fewer people stand around the patient. For procedures using low- to middle-energy gamma emitters (less than 250 keV;

e.g., <sup>99m</sup>Tc or <sup>111</sup>In) or beta minus emitters (e.g., <sup>90</sup>Y), the personnel's radiation burden remains acceptable if all protective measures are taken [99]. However, this is not the case for RGS based on PET (e.g., <sup>18</sup>F or <sup>68</sup>Ga) or high-energy gamma emitters (e.g., <sup>131</sup>I). In this case, beyond radioprotection means, it is recommended for surgeons and nurses to carry personal dosimeters and restrict the number of procedures per year depending on the isotope and the dose range used [100]. This downside of RGS has to be put in perspective with the wide variety of tracers available and the significantly higher penetration of gamma rays in tissue, allowing for in-depth detection possibilities.

Technologies that support the implementation of RGS concepts in the (robot-assisted) laparoscopic setting are discussed below.

Gamma counting Gamma counting has been the most used surgical modality within RGS [17, 101]. Moving towards minimal-invasive procedures, long "laparoscopic" gamma probes are routinely used in sentinel lymph node biopsies (SLNB) in prostate cancer [102] and cervical cancer [103], but also in several other indications. Unfortunately, in the laparoscopic setting, their use is complicated by the limited movability. Being a rigid instrument, placement of laparoscopic gamma probes during tracing is restricted due to the limited range of motion available when working through a trocar [33]. This is further complicated in the robotic setting since the surgeon is no longer located in the



sterile field. To improve positioning and at the same time regain autonomy for the surgeon and increase the range of motion available during tracing, a small-sized and tethered drop-in gamma probe was introduced [21, 33], a technology that is in line with the DROP-IN ultrasound technology that is used in robotic surgery [104]. Evaluations in prostate cancer SLNB indeed indicate an improved sentinel node detection rate for the DROP-IN (100%) versus laparoscopic gamma probe (76%) [22]. Recent studies indicate the DROP-IN also facilitates receptor-targeted surgery in the robotic setting (e.g., PSMA RGS [28]). These DROP-IN concepts applied to gamma probes in robotic surgery can be readily transferred to beta-probes [105] and as such allow for intraoperative detection of typical "PET-isotopes" (beta plus, e.g., <sup>18</sup>F or <sup>68</sup>Ga) and even opens the door for "therapeutic isotopes" (beta minus, e.g., <sup>90</sup>Y).

Intraoperative scintigraphy Portable gamma cameras can form a 2D image of the tracer distribution directly in the operating room [106-108] and probably even precede the use of gamma probes [109]. Initially developed for open surgery, their application has been shown in laparoscopic surgery as well, including prostate cancer, renal cell carcinoma, and testicular cancer SLNB procedures [110]. Although the gamma camera placement is more complicated during laparoscopy, its use helped verify the successful removal of the surgical targets. Providing only a 2D image, depth estimation is challenging, requiring frequent repositioning of the camera with different angles around the patient. To correlate the gamma image and the instruments in the field, radioactive markers on the instrument, e.g., a <sup>125</sup>I-seed on the tip of them and a dual-isotope mode on the camera, have been proposed with improved usability [111]. We, ourselves, have tried to use such portable gamma cameras during robot-assisted surgery. Unfortunately, this proved complex, resulting in frequent collisions between the robot and the camera. Usage of such cameras during robotic surgery is most likely only valuable when directly integrated into one of the robotic arms, or possibly the patient bed [109]—a solution that unfortunately has not made it to product but is technically feasible. Alternatively, truly laparoscopic gamma camera prototypes have been evaluated in phantoms [112]. In a similar preclinical setting, other research groups have demonstrated a beta-cameras' potential as a future application in (robot-assisted) laparoscopic RGS [18].

Freehand emission tomography 3D freehand imaging of radiopharmaceuticals is a method whereby a tracking system is used to determine the position and orientation of a nuclear detector (e.g., gamma probe or gamma camera) within the operating room, allowing for a 3D reconstruction based on the signal collected at these various positions

[113]. The most established RGS form of this, and the only one being applied in vivo yet, is freehand SPECT [97]. Combining this technology with a tracked US device, intraoperative freehand SPECT/US has also made it to patients [114-117]. Preclinical research has also been performed to investigate freehand imaging of beta emissions [95, 118], high-energy gamma emissions [119], or even freehand PET imaging using a coincidence or time-offlight principle with multiple detectors [120, 121]. While freehand SPECT is mainly used for open surgery procedures (e.g., SLNB or even receptor-targeted procedures [18]), this technology has also been translated to laparoscopic procedures, e.g., SLNB in gynecology and urology [77, 122] as well as radio-guided occult lesion localization in the lung [123]. However, the laparoscopic setting application is more challenging, where the limited movement of a laparoscopic probe restricts signal acquisition, and with that, reduces the quality of the freehand SPECT scan. Therefore, first steps are taken to translate the freehand imaging method towards robotic surgery using a tracked DROP-IN gamma probe [78, 124, 125]. Robotic SPECT has also been performed in a slightly different setting, using a robotic arm to autonomously create scans [126, 127]. These works have even extended to simultaneously acquire a cone-beam CT towards intraoperative SPECT/CT [128, 129]. Although the latter has not yet been tested in the surgical setting, one needs little imagination to hypothesize that such scans could be performed using robotic devices in the future. Interestingly, studies are being conducted to investigate if freehand SPECT can provide a reliable surgical roadmap (e.g., [130]). Some even suggest that freehand SPECT could potentially wholly replace the preoperative SPECT/CT. In line with this, no difference in performance was observed for 50 oral cancer SN procedures, where the surgeons were blinded for SPECT/CT [131].

# Fluorescence-guided and hybrid fluorescence-radio-guided surgery

Second to the application of RGS, fluorescence has seen a strong renewal of interest in the past decade [19, 132, 133]. Since an integrated laparoscopic camera is a crucial part of many robotic platforms, the interest in fluorescence imaging is even more emphasized by the fact that most surgical robots currently come with a laparoscope capable of fluorescence imaging.

Single-band fluorescence imaging The most common fluorescence imaging type is single-band, meaning that it uses a single band-pass filter to depict one sole fluorescent "color." In the clinic, fluorescence detection of the visible dye fluorescein and near-infrared (NIR) dye



indocyanine green (ICG) is currently a routinely applied tool for angiographic purposes. An increasing number of trials are being conducted in the area of lymphatic mapping (e.g., [134]), perfusion-based tumor resection (e.g., [135]), and receptor-targeted imaging (e.g., [136]). The main advantage of fluorescence is that it provides realtime visual feedback concerning the tracer uptake during surgery, especially useful to confirm successful lesion localization during excision. Interestingly, while fluorescence images satisfy the surgeon's demand for optical feedback, being based on light, it is inherently unsuited to visualize lesions located more than a couple of millimeters in tissue (i.e., < 1 cm depth [137]), something that is possible with for example nuclear medicine or radiological techniques. This was recently confirmed by Meershoek et al., demonstrating that in 52% of the patients, lesions were missed during robotic SLN procedures in prostate cancer if fluorescence imaging was used alone, underlining the need for additional technologies such as RGS [21]. This renders fluorescence imaging mostly useful for superficial applications and visual confirmation of successful target localization.

Multi-wavelength fluorescence imaging Every fluorescent dye has its absorption and emission spectrum. By wisely choosing non-conflicting spectra of the individual dyes, several (targeted) fluorescent tracers can be depicted simultaneously using so-called multi-wavelength (also known as multispectral or multicolor) fluorescence imaging. First in-human studies illustrate this has the potential to separate different anatomical structures [138], a concept that could be especially interesting to improve the balance between surgically induced cure and side effects (e.g., decrease damage to healthy nerves and lymphatics [139, 140]). While most proof-of-concept studies have evaluated the use in microscopic neurosurgery (e.g., in glioblastoma [141]), there are also examples in laparoscopic surgery (e.g., parathyroid surgery [142], bladder cancer [143], liver cancer [144], and gynecology [145], or even robot-assisted laparoscopic surgery (i.e., prostate cancer [24])). Contrary to popular belief that only NIR-I (700-900 nm) is useful for fluorescence imaging, based on the relatively low tissueinduced absorption of light at these wavelengths, these studies do not visualize that much of a difference in intensity when used in a surgical environment, justifying the use of different wavelengths as well [146]. Even deeper NIR (e.g., 1000 to 1700 nm) has been suggested, theoretically providing higher resolution due to lower light scattering at these wavelengths [147]. However, limiting to the whole approach of multi-wavelength fluorescence is that only a small amount of fluorescent tracers is currently clinically approved (i.e., ICG, fluorescein, methylene blue, PpIX<sup>5</sup>-ALA/HAL [138]).

Hybrid imaging Since detection techniques have their strengths and limitations, hybrid imaging (also referred to as dual-modal or bimodal imaging) combines the imaging properties of different modalities into a single technique, providing "best-of-both-worlds". This is not only relevant for modalities that combine anatomical information with molecular information, but even for modalities that combine two types of molecular imaging, where these might provide guidance during different parts of the image-guided surgery process. This is the case for the currently trending topic of hybrid fluorescence-radio-guided surgery that combines both fluorescent and nuclear signatures into a single tracer [15, 148]. This concept was first introduced in the clinic in 2010 using ICG-99mTc-HSA nanocolloid [149] for SLNB in various forms of cancer (e.g., prostate [150], penile [151], head-and-neck [152], cervical [153], and breast cancer [154], as well as melanoma [155]). Currently, it has been applied in > 1500 patients, of which some were already treated in the robotic setting. Following the success in SLNB, this hybrid concept is now increasingly adopted for research in receptor-targeted tracers (e.g., neuroendocrine tumors using Cy5-111 In-DTPA-Tyr3-octreotate [156], prostate cancer using PSMA I&F [157] and breast cancer using <sup>111</sup>In-DTPA-trastuzumab IRDye800 (Wang2015)). There, some have recently even entered first-in-human studies (e.g., clear cell renal cell carcinoma using <sup>111</sup>In-DOTA-girentuximab IRDye800 [30] or prostate cancer using <sup>64</sup>Ga-PSMA-914 [29]).

## Other intraoperative molecular guidance methods

Given the penetration constraints of fluorescence and the fear patients, in particular, have concerning radioactivity, several research groups have turned to alternative physical events. In the following, we will describe three trending methods that are either already applicable to robotic surgery or where we foresee an application in robotic surgery shortly.

Superparamagnetic iron-oxide nanoparticle detection Superparamagnetic iron-oxide nanoparticles (SPIONs) were introduced in 2013 as an alternative to radiolabelled colloids and blue dye in sLNB [158]. Clinically, their use has been restricted to SLNB (e.g., breast [159], vulva [160], prostate [161], penis cancer [162], and also melanoma [163]), or occult-lesion localization even in a laparoscopic setup [164]. As with radioactive tracers, SPIONs do allow for preoperative imaging and mapping of the sentinel nodes using MRI, where they usually show a black taint such that a degree of bare-eye guidance is possible. So far, the limitations in our view are in the lack of miniaturized detectors that can work in combination with surgical robots and slow tracer clearance from the body [165]. Interestingly,



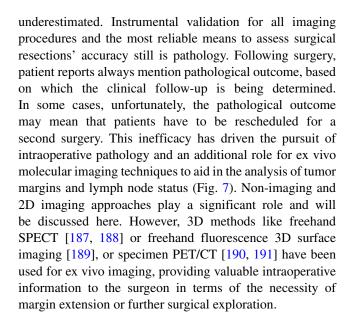
more and more groups are nevertheless actively working on bringing up tumor-specific SPIONs [166], which might offer alternatives for the future.

Multispectral optoacoustic tomography The concept of using pulsed light in tissue while imaging the resulting ultrasonic vibrations to discern differences in tissueabsorption was introduced in 2007 for diagnosing breast cancer [167, 168]. Along with improvements of the technology itself, a significant breakthrough was achieved by applying fluorophores to increase tissue contrast in 2012 [169]. This technique, currently known as multispectral optoacoustic tomography (MSOT), has the main advantage of an improved tissue-penetration (up to 5 cm [92]) with comparison to fluorescence imaging and the fact that the resulting images are tomographic. Applications have been reported for lymphatic imaging [170], breast cancer detection [171], characterization of non-melanoma skin cancers [172], among others. Current research focuses on miniaturizing the devices to make them suitable for laparoscopic surgery and developing AI-powered signal processing algorithms to improve the interpretability generated images (e.g., [173, 174]).

Raman spectrometry One of the promising technologies in the molecular imaging realm is Raman spectrometry, a technique that is based on the fact that different molecules, and with that different tissue types, have different Raman scattering spectra. This modality does not necessarily require a tracer. Its first applications in the operating room were as early as 2006 to evaluate resection margins in breast cancer [175]. Raman spectrometry can be applied to various indications like the detection of parathyroid adenoma [176] and the evaluation of lymph node metastasis in breast cancer [177], as well as the definition of resection margins in oral cancer [178, 179], glioma [180, 181], follicular thyroid cancer [182], and prostate cancer [94]. Given that Raman spectrometers can be easily miniaturized, porting them to a robotic setup is easy, as reported in the last reference. Like in MSOT, AI also has contributed significantly in particular by coping for illumination interference [183, 184] opening thus alternatives to generalize the use of this technology [185] and even beat fluorescence imaging in particular indications [186]. Using similar optical settings as with fluorescence imaging, tissue penetration is expected to be low (< 1 cm) making this a superficial technology.

## (Back table) Surgical verification

While preoperative and intraoperative molecular imaging have captured the spotlight over the last decades, the possibilities of ex vivo molecular imaging are not to be

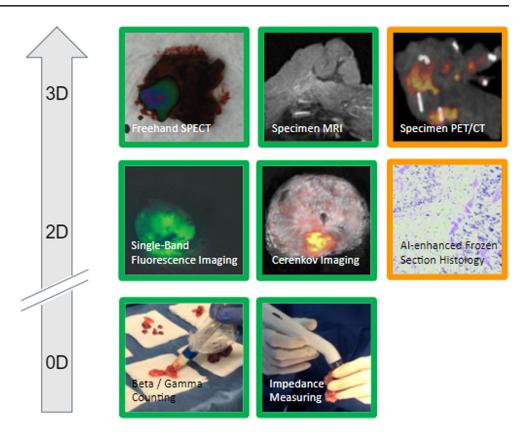


Intraoperative pathology The introduction of frozen section histology has been a significant step towards highspeed pathological tissue analysis, which can be performed within the time frame of the surgery itself. Not only has this been widely applied during lymphatic mapping, where it is recommended in almost all indications besides breast cancer and melanoma [192], it is also routinely used in primary tumor resection wherever big margins are not possible to avoid functional or aesthetic side effects, e.g., in skin cancer [193], breast cancer [194], head and neck cancer [195], and prostate cancer [196, 197]. However, having a pathologist available for intraoperative assessments is a luxury most hospitals can not afford. The waiting time until results are reported must be minimized (extension of anesthesia, blocking of the operating room, lost time for OR personnel, etc.). As such, methods for faster surgical verification have been proposed. Here, AI has made its first steps into the field, providing preliminary evaluations or highlighting suspicious areas, thus supporting the pathologist's efforts (e.g., [198, 199]). Such AI methods have great potential to accelerate the pathological process even further and might make intraoperative pathology logistically feasible for a lot more hospitals. One relevant alternative non-imaging approach is the intraoperative biochemical analyses of lymph nodes, like one-step nucleic acid amplification (OSNA). OSNA has made it to clinical routine in several countries and has shown diagnostic accuracy not only in breast cancer [200], but also in other tumor entities [201].

**Molecular non-imaging methods** Labeled tissue on the margins of specimens can be detected with non-imaging probes. This can be, for example, prostate tumors that are marked with PSMA [202] or sugar-avid tumors that are marked with FDG [203]. These methods can also



Fig. 7 Non-exhaustive overview of some current and possible future technologies for back table specimen analysis/intraoperative pathological evaluation separated by dimensionality and development status (green, commercially available; orange, research prototypes available). Non-imaging devices are defined as zero-dimensional as they are a single pixel detector and not a line detector which would be one-dimensional



verify if labeled structures were removed, like hyperactive parathyroids, or sentinel or metastatic lymph nodes. These control approaches are highly specific and are strongly recommended in guidelines for RGS. Alternatively, the tissue properties can be analyzed on-site using, for example, impedance/electromagnetic spectrometry [204], Raman spectrometry [186], or diffuse reflectance spectrometry [205], among others. Here, results are more controversial (e.g., [15]) and are not well spread in indications of robotic surgery.

Molecular imaging methods Of particular interest for robotic surgery are methods providing images, which can be played back in the surgeon's console and thus, for example, letting him or her see which margins to extend. Intraoperative imaging modalities like portable gamma camera scintigraphy and single-band fluorescence imaging have been shown to provide valuable information not only in radio-guided occult lesion localization setups, for instance [206, 207], but also in more challenging applications like PSMA-guided surgery [29]. An interesting approach that has made it through the regulatory path into the market is Cerenkov luminesce. Where beta probes directly detect the emitted beta plus or beta minus particles of a radiopharmaceutical, it is also possible to detect the Cerenkov light with specialized, highly sensitive cameras [208-210]. Since most of this light is generated in the ultraviolet spectrum, the penetration depth is estimated at a couple of millimeters, similar to beta particle detection [211, 212]. In practical terms, specimens of patients given beta-emitters before surgery, such as 68Ga-PSMA or 18F-FDG, can be imaged in a light-tight device next to the OR table, providing an image in 1–5 min [213, 214]. This is needed as the amount of emitted luminescence is only in the range of a few photons per radioactive decay [215]. Non-tracer approaches are also available, being optical coherence tomography (OCT), one option that is picking up more and more momentum (e.g., [216]).

## **Discussion and future perspectives**

By connecting advances made in the field of medical devices (hardware and software) with tracer-based molecular imaging strategies, it becomes possible to provide a more comprehensive view of the direction that precision surgery is moving to. The same technologies are likely to disseminate to other surgical approaches and indications where robotic surgery is leading the way in this technical (r)evolution. One thing is clear, though: The future of precision surgery will rely on an interplay between pre- and intraoperative imaging, surgical hardware, and advanced visualization strategies enhanced by AI [34]. Based on the above-reviewed literature, below we summarize the trends

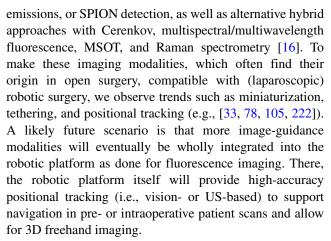


and future perspectives that we have derived from the main steps relevant to molecular image-guided surgery.

For non-invasive (total-body) molecular imaging, nuclear medicine remains the golden standard with an extensive range of radiotracers available for various indications, mostly using either SPECT or PET imaging, combined with anatomical imaging like CT or MRI (e.g., [217, 218]). With the uprising of AI strategies, the first studies underline a great potential in molecular imaging to accelerate and optimize the process of disease diagnosis (e.g., [45]). But this is not where it stops. Since an AI algorithm has the potential to quickly compare current patient information with gigantic databases of previously treated patients (including patient scans, but importantly also metadata such as age, health state, comorbidities, blood values), such technology could also be used to suggest the most optimal treatment strategies (i.e., patient selection) and predict treatment outcome (e.g., [53]).

In the framework of surgical navigation based on preoperative imaging, AI also has the potential to assist in surgical planning by providing (semi-)automatic segmentation of the structures of importance [219]. Various technologies are already used to register these roadmaps to the patient in the operating room, but to not overly complicate logistics, we expect that in the robotic setting, these registration technologies will eventually converge to such which are directly integrated into the robotic platforms (e.g., laparoscopic video-based and US-based registrations) [67, 78]. However, due to patient deformation and repositioning, accurate registration of preoperatively acquired roadmaps remains challenging in soft-tissue anatomies. This will remain a big topic of research for the upcoming years. Nonetheless, since the robotic approach requires the surgeon to operate behind a (video) console, there is significant potential to integrate these navigated approaches using intuitive augmented reality visualizations directly (see Fig. 1B) that include AI for an optimized perception (see Fig. 5F) [88].

Intraoperative planning, decision-making, and assistance during the excision are perhaps the area that receives the most focus from the image-guided surgery point of view. In particular, the use of tracers to illuminate specific structures using radioactive or fluorescent signals is popular [69, 220]. Both strategies have clear advantages and disadvantages, which can be overcome using hybrid tracers. As these tracers provide both imaging signatures, they are compatible with different imaging modalities. Concerning the radioactive imaging signatures used, most radioactive detection is currently established around SPECT-based signals (i.e., low-to-mid-energy gamma emissions from isotopes such as  $^{99m}$ Tc and  $^{111}$ In) [221]. This still leaves a lot of room to investigate the value of alternative approaches using, for example, PET-based signals (i.e., beta plus or high-energy gamma emissions), beta minus



Surgical verification is another area where molecular imaging is playing an emerging role. Imaging and counting modalities already enable confirming ex vivo the removal of structures seen both in preoperative images and detected intraoperatively (e.g., [202, 203, 207]). Dedicated specimen analysis methods will still play a role, in particular, there where intraoperative imaging cannot provide high image resolution ([208] vs. [119]). The more information becomes available and the better the registration, the more tasks will be taken over semi-autonomously by the robot [223]. From there, the step to step to autonomous robotic system is not far, and (intraoperative) image guidance will be instrumental. Several research groups have already shown that robots can already take actions based on previously defined trajectories derived from preoperative images (e.g., [224, 225]) or be guided by intraoperative imaging (e.g., [226]). More developments in that direction are to be expected.

## **Conclusions**

With the uprising of minimal-invasive robotic surgery, more and more instruments are becoming available to optimize surgical actions. Integration of molecular imaging might be the key to bringing precision surgery to the next level. The robotic platform seems to be ideal for the direct integration of image-guided surgery technologies. For most technologies, the way thereto is still long. Still, the research and engineering community and the exponential growth of AI are pushing towards precision surgery solutions that fully integrate preoperative, intraoperative, and postoperative imaging modalities to achieve an optimal patient outcome. Nuclear medicine plays a crucial role in this evolution, facilitating computer-assisted diagnosis, planning, (robotic) navigation and detection, and (back table) verification throughout the complete surgical route.

**Author contribution** All the authors defined the structure of the paper jointly and its goal. TW, FvL, and MvO performed the literature review, prepared the figures, and drafted the paper. NN reviewed the



paper and provided critical comments and recommendations during the complete drafting process.

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#### **Declarations**

Conflict of interest TW is a consultant for technology developments for the medical device companies SurgicEye and Crystal Photonics. FvL is a consultant for the medical device company Hamamatsu Photonics. The remaining authors do not have any conflict of interest to disclose.

**Ethical approval** This article does not contain any studies with human participants or animals performed by any of the authors. It is a review of the literature.

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## References

- Lidsky ME, D'Angelica MI. An outlook on precision surgery. Eur J Surg Oncol. 2017;43(5):853–855. https://doi.org/10.1016/j. ejso.2016.11.014.
- Liu S, Hemal A. Techniques of robotic radical prostatectomy for the management of prostate cancer: which one, when and why. Transl Androl Urol. 2020;9(2):906–918. https://doi.org/10.21037/tau.2019.09.13.
- 3. Petersen LJ, Zacho HD. PSMA PET for primary lymph node staging of intermediate and high-risk prostate cancer: an expedited systematic review. Cancer Imaging. 2020;20(1):10. https://doi.org/10.1186/s40644-020-0290-9.
- Harbin AC, Eun DD. The role of extended pelvic lymphadenectomy with radical prostatectomy for high-risk prostate cancer. Urol Oncol. 2015;33(5):208–216. https://doi.org/10.1016/j.urolonc.2014.11.011.
- Tsai S-H, Tseng P-T, Sherer BA, Lai Y-C, Lin P-Y, Wu C-K, Stoller ML. Open versus robotic partial nephrectomy: Systematic review and meta-analysis of contemporary studies. Int J Med Robot. 2019;15(1):e1963. https://doi.org/10.1002/rcs.1963.
- Wang J, Li X, Wu H, Zhang Y, Wang F. A Meta-Analysis of Robotic Surgery in Endometrial Cancer: Comparison with Laparoscopy and Laparotomy. Dis Markers. 2020;2020:2503753. https://doi.org/10.1155/2020/2503753.
- Schwartz G, Sancheti M, Blasberg J. Robotic Thoracic Surgery. Surg Clin North Am. 2020;100(2):237–248. https://doi.org/10.1016/j.suc.2019.12.001.

- Guerra F, Di Marino M, Coratti A. Robotic Surgery of the Liver and Biliary Tract. J Laparoendosc Adv Surg Tech A. 2019;29(2):141–146. https://doi.org/10.1089/lap.2017.0628.
- Addison P, Agnew JL, Martz J. Robotic Colorectal Surgery. Surg Clin North Am. 2020;100(2):337–360. https://doi.org/10. 1016/j.suc.2019.12.012.
- Finegersh A, Holsinger FC, Gross ND, Orosco RK. Robotic Head and Neck Surgery. Surg Oncol Clin N Am. 2019;28(1):115–128. https://doi.org/10.1016/j.soc.2018.07.008.
- Zhou M, Hamad M, Weiss J, Eslami A, Huang K, Maier M, Lohmann CP, Navab N, Knoll A, Nasseri MA. Towards Robotic Eye Surgery: Marker-Free, Online Hand-Eye Calibration Using Optical Coherence Tomography Images. IEEE Robot Autom Lett. 2018;3(4):3944–3951. https://doi.org/10.1109/LRA.2018. 2858744.
- Nuzzi R, Brusasco L. State of the art of robotic surgery related to vision: brain and eye applications of newly available devices. Eye Brain. 2018;10:13–24. https://doi.org/10.2147/EB.S148644.
- Liu Z, Wang S, Dong D, Wei J, Fang C, Zhou X, Sun K, Li L, Li B, Wang M, Tian J. The Applications of Radiomics in Precision Diagnosis and Treatment of Oncology: Opportunities and Challenges. Theranostics. 2019;9(5):1303–1322. https://doi.org/10.7150/thno.30309.
- Weissleder R. Molecular imaging: exploring the next frontier. Radiology. 1999;212(3):609–614. https://doi.org/10.1148/radiology.212.3.r99se18609.
- van Leeuwen FWB, Schottelius M, Brouwer OR, Vidal-Sicart S, Achilefu S, Klode J, Wester H-J, Buckle T. Trending: Radioactive and Fluorescent Bimodal/Hybrid Tracers as Multiplexing Solutions for Surgical Guidance. J Nucl Med. 2020;61(1):13–19. https://doi.org/10.2967/jnumed.119.228684.
- Mondal SB, O'Brien CM, Bishop K, Fields RC, Margenthaler JA, Achilefu S. Repurposing Molecular Imaging and Sensing for Cancer Image-Guided Surgery. J Nucl Med. 2020;61(8):1113– 1122. https://doi.org/10.2967/jnumed.118.220426.
- Povoski SP, Neff RL, Mojzisik CM, O'Malley DM, Hinkle GH, Hall NC, Murrey DA, Knopp MV, Martin EW. A comprehensive overview of radioguided surgery using gamma detection probe technology. World J Surg Oncol. 2009;7:11. https://doi.org/10.1186/1477-7819-7-11.
- van Oosterom MN, Rietbergen DDD, Welling MM, Poel HGVD, Maurer T, van Leeuwen FWB. Recent advances in nuclear and hybrid detection modalities for imageguided surgery. Expert Rev Med Dev. 2019;16(8):711–734. https://doi.org/10.1080/17434440.2019.1642104.
- van Leeuwen FWB, Hardwick JCH, van Erkel AR. Luminescence-based Imaging Approaches in the Field of Interventional Molecular Imaging. Radiology. 2015;276(1):12– 29. https://doi.org/10.1148/radiol.2015132698.
- Qian L, Wu JY, DiMaio SP, Navab N, Kazanzides P. A Review of Augmented Reality in Robotic-Assisted Surgery. IEEE Trans Med Robot Bion. 2020;2(1):1–16. https://doi.org/10.1109/TMRB.2019.2957061.
- Meershoek P, van Oosterom MN, Simon H, Mengus L, Maurer T, van Leeuwen PJ, Wit EMK, van der Poel HG, van Leeuwen FWB. Robot-assisted laparoscopic surgery using DROP-IN radioguidance: first-in-human translation. Eur J Nucl Med Mol Imaging. 2019;46(1):49–53. https://doi.org/10. 1007/s00259-018-4095-z.
- Dell'Oglio P, Meershoek P, Maurer T, Wit EMK, van Leeuwen PJ, van der Poel HG, van Leeuwen FWB, van Oosterom MN. A DROP-IN Gamma Probe for Robot-assisted Radioguided Surgery of Lymph Nodes During Radical Prostatectomy. Eur Urol. 2021;79(1):124–132. https://doi.org/10.1016/j. eururo.2020.10.031.



- 23. Harke NN, Godes M, Wagner C, Addali M, Fangmeyer B, Urbanova K, Hadaschik B, Witt JH. Fluorescence-supported lymphography and extended pelvic lymph node dissection in robot-assisted radical prostatectomy: a prospective, randomized trial. World J Urol. 2018;36(11):1817–1823. https://doi.org/10.1007/s00345-018-2330-7.
- 24. van den Berg NS, Buckle T, KleinJan GH, van der Poel HG, van Leeuwen FWB. Multispectral Fluorescence Imaging During Robot-assisted Laparoscopic Sentinel Node Biopsy: A First Step Towards a Fluorescence-based Anatomic Roadmap. Eur Urol. 2017;72(1):110–117. https://doi.org/10.1016/j.eururo.2016.06.012.
- Rozenholc A, Samouelian V, Warkus T, Gauthier P, Provencher D, Sauthier P, Gauthier F, Drakopoulos P, Cormier B. Green versus blue: Randomized controlled trial comparing indocyanine green with methylene blue for sentinel lymph node detection in endometrial cancer. Gynecol Oncol. 2019;153(3):500–504. https://doi.org/10.1016/j.ygyno.2019.03. 103.
- Marino MV, Di Saverio S, Podda M, Gomez Ruiz M, Gomez Fleitas M. The Application of Indocyanine Green Fluorescence Imaging During Robotic Liver Resection: A Case-Matched Study. World J Surg. 2019;43(10):2595–2606. https://doi.org/10.1007/s00268-019-05055-2.
- Jafari MD, Lee KH, Halabi WJ, Mills SD, Carmichael JC, Stamos MJ, Pigazzi A. The use of indocyanine green fluorescence to assess anastomotic perfusion during robotic assisted laparoscopic rectal surgery. Surg Endosc. 2013;27(8):3003– 3008. https://doi.org/10.1007/s00464-013-2832-8.
- van Leeuwen FWB, van Oosterom MN, Meershoek P, van Leeuwen PJ, Berliner C, van der Poel HG, Graefen M, Maurer T. Minimal-Invasive Robot-Assisted Image-Guided Resection of Prostate-Specific Membrane Antigen-Positive Lymph Nodes in Recurrent Prostate Cancer. Clin Nucl Med. 2019;44(7):580–581. https://doi.org/10.1097/RLU.0000000000002600.
- 29. Eder A.-C., Omrane MA, Stadlbauer S, Roscher M, Khoder WY, Gratzke C, Kopka K, Eder M, Meyer PT, Jilg CA, Ruf J. The PSMA-11-derived hybrid molecule PSMA-914 specifically identifies prostate cancer by preoperative PET/CT and intraoperative fluorescence imaging. Eur J Nucl Med Mol Imaging. 2021. https://doi.org/10.1007/s00259-020-05184-0.
- Hekman MC, Rijpkema M, Muselaers CH, Oosterwijk E, Hulsbergen-Van de Kaa CA, Boerman OC, Oyen WJ, Langenhuijsen JF, Mulders PF. Tumor-targeted Dual-modality Imaging to Improve Intraoperative Visualization of Clear Cell Renal Cell Carcinoma: A First in Man Study. Theranostics. 2018;8(8):2161–2170. https://doi.org/10.7150/thno.23335.
- Shum CF, Bahler CD, Low PS, Ratliff TL, Kheyfets SV, Natarajan JP, Sandusky GE, Sundaram CP. Novel Use of Folate-Targeted Intraoperative Fluorescence, OTL38, in Robot-Assisted Laparoscopic Partial Nephrectomy: Report of the First Three Cases. J Endourol Case Rep. 2016;2(1):189–197. https://doi.org/10.1089/cren.2016.0104.
- Predina JD, Newton AD, Keating J, Barbosa EM, Okusanya O, Xia L, Dunbar A, Connolly C, Baldassari MP, Mizelle J, Delikatny EJ, Kucharczuk JC, Deshpande C, Kularatne SA, Low P, Drebin J, Singhal S. Intraoperative Molecular Imaging Combined With Positron Emission Tomography Improves Surgical Management of Peripheral Malignant Pulmonary Nodules. Ann Surg. 2017;266(3):479–488. https://doi.org/10.1097/SLA.00000000000002382.
- van Oosterom MN, Simon H, Mengus L, Welling MM, van der Poel HG, van den Berg NS, van Leeuwen FW. Revolutionizing (robot-assisted) laparoscopic gamma tracing

- using a drop-in gamma probe technology. Am J Nucl Med Mol Imaging. 2016;6(1):1–17.
- 34. Andras I, Mazzone E, van Leeuwen FWB, De Naeyer G, van Oosterom MN, Beato S, Buckle T, O'Sullivan S, van Leeuwen PJ, Beulens A, Crisan N, D'Hondt F, Schatteman P, van Der Poel H, Dell'Oglio P, Mottrie A. Artificial intelligence and robotics: a combination that is changing the operating room. World J Urol. 2020;38(10):2359–2366. https://doi.org/10.1007/s00345-019-03037-6.
- 35. Valdés Olmos RA, Rietbergen DD, Vidal-Sicart S, Manca G, Giammarile F, Mariani G. Contribution of SPECT/CT imaging to radioguided sentinel lymph node biopsy in breast cancer, melanoma, and other solid cancers: from "open and see" to "see and open". Q J Nucl Med Mol Imaging. 2014;58(2): 127–139.
- 36. Kido S, Hirano Y, Mabu S. Deep Learning for Pulmonary Image Analysis: Classification, Detection, and Segmentation. Adv Exp Med Biol. 2020;1213:47–58. https://doi.org/10.1007/978-3-030-33128-3\_3.
- 37. Currie G, Rohren E. Intelligent Imaging in Nuclear Medicine: the Principles of Artificial Intelligence, Machine Learning and Deep Learning. Semin Nucl Med. 2021;51(2):102–111. https://doi.org/10.1053/j.semnuclmed.2020.08.002.
- Zhou X. Automatic Segmentation of Multiple Organs on 3D CT Images by Using Deep Learning Approaches. Adv Exp Med Biol. 2020;1213:135–147. https://doi.org/10.1007/978-3-030-33128-3-9.
- 39. Burwinkel H, Kazi A, Vivar G, Albarqouni S, Zahnd G, Navab N, Ahmadi S-A. Adaptive Image-Feature Learning for Disease Classification Using Inductive Graph Networks. Medical Image Computing and Computer Assisted Intervention MICCAI 2019, Lecture Notes in Computer Science. In: Shen D, Liu T, Peters TM, Staib LH, Essert C, Zhou S, Yap P-T, and Khan A, editors. Cham: Springer International Publishing; 2019. p. 640–648.
- Dong X, Lei Y, Wang T, Thomas M, Tang L, Curran WJ, Liu T, Yang X. Automatic multiorgan segmentation in thorax CT images using U-net-GAN. Med Phys. 2019;46(5):2157–2168. https://doi.org/10.1002/mp.13458.
- 41. Han M, Yao G, Zhang W, Mu G, Zhan Y, Zhou X, Gao Y. Segmentation of CT thoracic organs by multi-resolution vbnets. Proceedings of the 2019 Challenge on Segmentation of THoracic Organs at Risk in CT Images, SegTHOR@ISBI 2019, April 8, 2019. In: Petitjean C, Ruan S, Lambert Z, and Dubray B, editors, CEUR Workshop Proceedings. CEUR-WS.org; 2019. http://ceur-ws.org/Vol-2349/SegTHOR2019\_paper\_1.pdf.
- 42. He T, Hu J, Song Y, Guo J, Yi Z. Multi-task learning for the segmentation of organs at risk with label dependence. Med Image Anal. 2020;61:101666. https://doi.org/10.1016/j.media.2020.101666.
- 43. Lavdas I, Glocker B, Kamnitsas K, Rueckert D, Mair H, Sandhu A, Taylor SA, Aboagye EO, Rockall AG. Fully automatic, multiorgan segmentation in normal whole body magnetic resonance imaging (MRI), using classification forests (CFs), convolutional neural networks (CNNs), and a multi-atlas (MA) approach. Med Phys. 2017;44(10):5210–5220. https://doi.org/10.1002/mp.12492.
- 44. Feng X, Qing K, Tustison NJ, Meyer CH, Chen Q. Deep convolutional neural network for segmentation of thoracic organs-at-risk using cropped 3D images. Med Phys. 2019;46(5):2169–2180. https://doi.org/10.1002/mp.13466.
- 45. Zhao Y, Gafita A, Vollnberg B, Tetteh G, Haupt F, Afshar-Oromieh A, Menze B, Eiber M, Rominger A, Shi K. Deep neural network for automatic characterization of lesions on 68Ga-PSMA-11 PET/CT. Eur J Nucl Med Mol Imaging. 2020; 47(3):603–613. https://doi.org/10.1007/s00259-019-04606-y.



- Zhong Z, Kim Y, Zhou L, Plichta K, Allen B, Buatti J, Wu X. 3D fully convolutional networks for co-segmentation of tumors on PET-CT images. In: 2018 IEEE 15th International Symposium on Biomedical Imaging (ISBI 2018); 2018. p. 228– 231.
- Li L, Zhao X, Lu W, Tan S. Deep Learning for Variational Multimodality Tumor Segmentation in PET/CT. Neurocomputing. 2020;392:277–295. https://doi.org/10.1016/j.neucom.2018. 10.099.
- 48. Chen H, Chen H, Wang L. Iteratively Refine the Segmentation of Head and Neck Tumor in FDG-PET and CT Images. Head and Neck Tumor Segmentation, Lecture Notes in Computer Science. In: Andrearczyk V, Oreiller V, and Depeursinge A, editors. Cham: Springer International Publishing; 2021. p. 53–58.
- 49. Huang B, Chen Z, Wu P-M, Ye Y, Feng S-T, Wong C-YO, Zheng L, Liu Y, Wang T, Li Q, Huang B. Fully Automated Delineation of Gross Tumor Volume for Head and Neck Cancer on PET-CT Using Deep Learning: A Dual-Center Study. Contrast Media Mol Imaging. 2018;2018:8923028. https://doi.org/10.1155/2018/8923028.
- Jemaa S, Fredrickson J, Carano RAD, Nielsen T, de Crespigny A, Bengtsson T. Tumor Segmentation and Feature Extraction from Whole-Body FDG-PET/CT Using Cascaded 2D and 3D Convolutional Neural Networks. J Digit Imaging. 2020;33(4):888–894. https://doi.org/10.1007/s10278-020-00341-1.
- Liu L, Zhang B, Wang H. Organ Localization in PET/CT Images using Hierarchical Conditional Faster R-CNN Method. In: Proceedings of the Third International Symposium on Image Computing and Digital Medicine, ISICDM 2019. New York: Association for Computing Machinery; 2019. p. 249–253.
- 52. Amyar A, Ruan S, Gardin I, Chatelain C, Decazes P, Modzelewski R. 3-D RPET-NET: Development of a 3-D PET Imaging Convolutional Neural Network for Radiomics Analysis and Outcome Prediction. IEEE Trans Radiat Plasma Med Sci. 2019;3(2):225–231. https://doi.org/10.1109/TRPMS.2019.2896399.
- Wang C, Liu C, Chang Y, Lafata K, Cui Y, Zhang J, Sheng Y, Mowery Y, Brizel D, Yin F-F. Dose-Distribution-Driven PET Image-Based Outcome Prediction (DDD-PIOP): A Deep Learning Study for Oropharyngeal Cancer IMRT Application. Front Oncol. 2020;10:1592. https://doi.org/10.3389/fonc.2020.01592.
- 54. Testori A, Rastrelli M, De Fiori E, Soteldo J, Della Vigna P, Trifir G, Mazzarol G, Travaini LL, Verrecchia F, Ratto EL, Bellomi M. Radio-guided ultrasound lymph node localization: feasibility of a new technique for localizing and excising nonpalpable lymph nodes ultrasound suspicious for melanoma metastases. Melanoma Res. 2010;20(3):197–202. https://doi.org/10.1097/CMR.0b013e3283350527.
- Vilar Tabanera A, Ajuria O, Rioja ME, Caba ns Montero J. Selective Neck Dissection Guided by a Radioactive I125 Seed for Papillary Thyroid Carcinoma Recurrence. Cir Esp. 2020; 98(8):478–481. https://doi.org/10.1016/j.ciresp.2020.04.018.
- Einspieler I, Novotny A, Okur A, Essler M, Martignoni ME. First experience with image-guided resection of paraganglioma. Clin Nucl Med. 2014;39(8):e379–381. https://doi.org/10.1097/RLU.0000000000000239.
- 57. Badenes-Romero A, Orozco-Cortés J, Balaguer-Mu noz D, Abreu SÁnchez P, Mut Dólera T, Gómez-Abril SA, Dolz-Gaitón R, Cabellero Calabuig E, Cueto Caadas B, Latorre Agraz I, Reyes Ojeda MD, Plancha Mansanet C, Esteban Hurtado A. Detección radioguiada de lesión oculta no palpable (ROLL) en un caso de metÁstasis abdominal de tumor neuroendocrino. Rev Esp Enferm Dig. 2020;112(10):768–771. https://doi.org/10.17235/reed.2020.6926/2020.

- Reiner BI, Siegel EL, Hooper F, Pomerantz SM, Protopapas Z, Pickar E, Killewich L. Picture archiving and communication systems and vascular surgery: clinical impressions and suggestions for improvement. J Digit Imaging. 1996;9(4):167–171. https://doi.org/10.1007/BF03168613.
- Bhayani SB, Snow DC. Novel dynamic information integration during da Vinci robotic partial nephrectomy and radical nephrectomy. J Robot Surg. 2008;2(2):67–69. https://doi.org/10.1007/s11701-008-0083-9.
- 60. Porpiglia F, Fiori C, Checcucci E, Amparore D, Bertolo R. Hyperaccuracy Three-dimensional Reconstruction Is Able to Maximize the Efficacy of Selective Clamping During Robot-assisted Partial Nephrectomy for Complex Renal Masses. Eur Urol. 2018;74(5):651–660. https://doi.org/10.1016/j.eururo.2017.12.027.
- Navab N, Blum T, Wang L, Okur A, Wendler T. First Deployments of Augmented Reality in Operating Rooms. Computer. 2012;45(7):48–55. https://doi.org/10.1109/MC.2012.75.
- 62. Vorbeck F, Cartellieri M, Ehrenberger K, Imhof H. Intraoperative navigation in paranasal sinus surgery with the Philips "Neuroguide" system. Radiologe. 2000;40(3):227–232. https://doi.org/10.1007/s001170050661.
- 63. Su L-M, Vagvolgyi BP, Agarwal R, Reiley CE, Taylor RH, Hager GD. Augmented Reality During Robot-assisted Laparoscopic Partial Nephrectomy: Toward Real-Time 3D-CT to Stereoscopic Video Registration. Urol. 2009;73(4):896–900. https://doi.org/10.1016/j.urology.2008.11.040.
- 64. Robu MR, Ramalhinho J, Thompson S, Gurusamy K, Davidson B, Hawkes D, Stoyanov D, Clarkson MJ. Global rigid registration of CT to video in laparoscopic liver surgery. Int J Comput Assist Radiol Surg. 2018;13(6):947–956. https://doi.org/10.1007/s11548-018-1781-z.
- 65. Hattab G, Arnold M, Strenger L, Allan M, Arsentjeva D, Gold O, Simpfendörfer T, Maier-Hein L, Speidel S. Kidney edge detection in laparoscopic image data for computer-assisted surgery: Kidney edge detection. Int J Comput Assist Radiol Surg. 2020;15(3):379–387. https://doi.org/10.1007/s11548-019-02102-0.
- 66. Luo H, Yin D, Zhang S, Xiao D, He B, Meng F, Zhang Y, Cai W, He S, Zhang W, Hu Q, Guo H, Liang S, Zhou S, Liu S, Sun L, Guo X, Fang C, Liu L, Jia F. Augmented reality navigation for liver resection with a stereoscopic laparoscope. Comput Methods Programs Biomed. 2020;187:105099. https://doi.org/10.1016/j.cmpb.2019.105099.
- 67. Samei G, Tsang K, Kesch C, Lobo J, Hor S, Mohareri O, Chang S, Goldenberg SL, Black PC, Salcudean S. A partial augmented reality system with live ultrasound and registered preoperative MRI for guiding robot-assisted radical prostatectomy. Med Image Anal. 2020;60:101588. https://doi.org/10.1016/j.media.2019.101588.
- 68. Hughes-Hallett A, Pratt P, Mayer E, Marco AD, Yang G-Z, Vale J, Darzi A. Intraoperative Ultrasound Overlay in Robot-assisted Partial Nephrectomy: First Clinical Experience. Eur Urol. 2014;65(3):671–672. https://doi.org/10.1016/j.eururo. 2013.11.001.
- 69. van Oosterom MN, van der Poel HG, Navab N, van de Velde CJH, van Leeuwen FWB. Computer-assisted surgery: virtual- and augmented-reality displays for navigation during urological interventions. Curr Opin Urol. 2018;28(2):205–213. https://doi.org/10.1097/MOU.00000000000000478.
- Shekhar R, Dandekar O, Bhat V, Philip M, Lei P, Godinez C, Sutton E, George I, Kavic S, Mezrich R, Park A. Live augmented reality: a new visualization method for laparoscopic surgery using continuous volumetric computed tomography. Surg Endosc. 2010;24(8):1976–1985. https://doi.org/10.1007/s00464-010-0890-8.



- Bernhardt S, Nicolau SA, Agnus V, Soler L, Doignon C, Marescaux J. Automatic localization of endoscope in intraoperative CT image: A simple approach to augmented reality guidance in laparoscopic surgery. Med Image Anal. 2016;30:130–143. https://doi.org/10.1016/j.media.2016.01.008.
- Vetter C, Lasser T, Wendler T, Navab N. 1D-3D registration for functional nuclear imaging. Med Image Comput Comput Assist Interv. 2011;14(Pt 1):227–234.
- Pinto F, Fuerst B, Frisch B, Navab N. Radiopositive Tissue Displacement Compensation for SPECT-guided Surgery. Medical Image Computing and Computer-Assisted Intervention – MICCAI 2015, Lecture Notes in Computer Science. In: Navab N, Hornegger J, Wells WM, and Frangi A, editors. Cham: Springer International Publishing; 2015. p. 536–543.
- 74. Brouwer OR, van den Berg NS, Mathéron HM, Wendler T, van der Poel HG, Horenblas S, Valdés Olmos RA, van Leeuwen FWB. Feasibility of Intraoperative Navigation to the Sentinel Node in the Groin Using Preoperatively Acquired Single Photon Emission Computerized Tomography Data: Transferring Functional Imaging to the Operating Room. J Urol. 2014;192(6):1810–1816. https://doi.org/10.1016/j.juro.2014.03.127.
- Brouwer OR, Buckle T, Bunschoten A, Kuil J, Vahrmeijer AL, Wendler T, Valdés-Olmos RA, van der Poel HG, van Leeuwen FWB. Image navigation as a means to expand the boundaries of fluorescence-guided surgery. Phys Med Biol. 2012;57(10):3123– 3136. https://doi.org/10.1088/0031-9155/57/10/3123.
- KleinJan GH, van den Berg NS, van Oosterom MN, Wendler T, Miwa M, Bex A, Hendricksen K, Horenblas S, van Leeuwen FWB. Toward (Hybrid) Navigation of a Fluorescence Camera in an Open Surgery Setting. J Nucl Med. 2016;57(10):1650–1653. https://doi.org/10.2967/jnumed.115.171645.
- 77. van Oosterom MN, Meershoek P, KleinJan GH, Hendricksen K, Navab N, van de Velde CJH, van der Poel HG, van Leeuwen FWB. Navigation of Fluorescence Cameras during Soft Tissue Surgery-Is it Possible to Use a Single Navigation Setup for Various Open and Laparoscopic Urological Surgery Applications?. J Urol. 2018;199(4):1061–1068. https://doi.org/10.1016/j.juro.2017.09.160.
- 78. Azargoshasb S, Houwing KHM, Roos PR, van Leeuwen SI, Boonekamp M, Mazzone E, Bauwens K, Dell'Oglio P, van Leeuwen F, van Oosterom MN. Optical navigation of a DROP-IN gamma probe as a means to strengthen the connection between robot-assisted and radioguided surgery. J Nucl Med. 2021 https://doi.org/10.2967/jnumed.120.259796.
- 79. Volonté F, Buchs NC, Pugin F, Spaltenstein J, Schiltz B, Jung M, Hagen M, Ratib O, Morel P. Augmented reality to the rescue of the minimally invasive surgeon. The usefulness of the interposition of stereoscopic images in the Da Vinci robotic console. Int J Med Robot. 2013;9(3):e34–38. https://doi.org/10.1002/rcs.1471.
- López-Mir F, Naranjo V, Fuertes JJ, Alcaiz M, Bueno J, Pareja E. Design and Validation of an Augmented Reality System for Laparoscopic Surgery in a Real Environment. BioMed Res Int. 2013;2013:758491. https://doi.org/10.1155/2013/758491.
- 81. Wagner A, Ploder O, Enislidis G, Truppe M, Ewers R. Virtual image guided navigation in tumor surgery-technical innovation. J Craniomaxillofac Surg. 1995;23(5):217–213. https://doi.org/10.1016/s1010-5182(05)80155-6.
- 82. King AP, Edwards PJ, Maurer CR, de Cunha DA, Hawkes DJ, Hill DL, Gaston RP, Fenlon MR, Strong AJ, Chandler CL, Richards A, Gleeson MJ. A system for microscope-assisted guided interventions. Stereotact Funct Neurosurg. 1999;72(2-4):107–111. https://doi.org/10.1159/000029708.

- Bichlmeier C, Heining SM, Feuerstein M, Navab N. The virtual mirror: a new interaction paradigm for augmented reality environments. IEEE Trans Med Imaging. 2009;28(9):1498– 1510. https://doi.org/10.1109/TMI.2009.2018622.
- 84. Bichlmeier C, Sandro Michael H, Mohammad R, Nassir N. Laparoscopic Virtual Mirror for Understanding Vessel Structure: Evaluation Study by Twelve Surgeons. In: Proceedings of the 6th International Symposium on Mixed and Augmented Reality (ISMAR). Nara, Japan; 2007. p. 125–128.
- 85. Bichlmeier C, Kipot M, Holdstock S, Heining SM, Euler E, Navab N. A Practical Approach for Intraoperative Contextual In-Situ Visualization. In: International Workshop on Augmented environments for Medical Imaging including Augmented Reality in Computer-aided Surgery (AMI-ARCS 2009). New York: MICCAI Society; 2009.
- 86. Kutter O, Aichert A, Bichlmeier C, Michael R, Ockert B, Euler E, Navab N. Real-time Volume Rendering for High Quality Visualization. In: in Augmented Reality. International Workshop on Augmented environments for Medical Imaging including Augmented Reality in Computer-aided Surgery (AMI-ARCS 2008; 2008.
- 87. Pakhomov D, Shen W, Navab N. Towards Unsupervised Learning for Instrument Segmentation in Robotic Surgery with Cycle-Consistent Adversarial Networks. In: IEEE/RSJ Int. Conf. on Intelligent Robots and Systems (IROS). Las Vegas: IEEE; October 2020. p. 8499–8504.
- Pauly O, Diotte B, Fallavollita P, Weidert S, Euler E, Navab N. Machine learning-based augmented reality for improved surgical scene understanding. Comput Med Imaging Graph. 2015;41:55– 60. https://doi.org/10.1016/j.compmedimag.2014.06.007.
- 89. Ojodu I, Ogunsemoyin A, Hopp S, Pohlemann T, Ige O, Akinola O. C-arm fluoroscopy in orthopaedic surgical practice. Eur J Orthop Surg Traumatol. 2018;28(8):1563–1568. https://doi.org/10.1007/s00590-018-2234-7.
- Kamiyama T, Kakisaka T, Orimo T. Current role of intraoperative ultrasonography in hepatectomy. Surg Today. 2021. https://doi.org/10.1007/s00595-020-02219-9.
- 91. Noh T, Mustroph M, Golby AJ. Intraoperative Imaging for High-Grade Glioma Surgery. Neurosurg Clin N Am. 2021;32(1):47–54. https://doi.org/10.1016/j.nec.2020.09.003.
- 92. MacCuaig WM, Jones MA, Abeyakoon O, McNally LR. Development of Multispectral Optoacoustic Tomography as a Clinically Translatable Modality for Cancer Imaging. Radiol Imaging Cancer. 2020;2(6):e200066. https://doi.org/10.1148/rycan.2020200066.
- 93. Ferrer-Roca O. Telepathology and Optical Biopsy. https://doi.org/10.1155/2009/740712. 2010.
- 94. Pinto M, Zorn KC, Tremblay J-P, Desroches J, Dallaire F, Aubertin K, Marple ET, Kent C, Leblond F, Trudel D, Lesage F. Integration of a Raman spectroscopy system to a robotic-assisted surgical system for real-time tissue characterization during radical prostatectomy procedures. JBO. 2019;24(2):025001. https://doi.org/10.1117/1.JBO.24.2.025001.
- Wendler T, Traub J, Ziegler SI, Navab N. Navigated three dimensional beta probe for optimal cancer resection. Med Image Comput Comput Assist Interv. 2006;9(Pt 1):561–569.
- Wendler T, Hartl A, Lasser T, Traub J, Daghighian F, Ziegler SI, Navab N. Towards intra-operative 3D nuclear imaging: reconstruction of 3D radioactive distributions using tracked gamma probes. Med Image Comput Comput Assist Interv. 2007;10(Pt 2):909–917. https://doi.org/10.1007/978-3-540-75759-7\_110.
- Bluemel C, Matthies P, Herrmann K, Povoski SP.
   Scintigraphic Imaging and Navigation in Radioguided Surgery: Freehand SPECT Technology and its



- Clinical Applications. Expert Rev Med Devices. 2016. https://doi.org/10.1586/17434440.2016.1154456.
- 98. Maurer T, Robu S, Schottelius M, Schwamborn K, Rauscher I, van den Berg NS, van Leeuwen FWB, Haller B, Horn T, Heck MM, Gschwend JE, Schwaiger M, Wester H-J, Eiber M. 99mTechnetium-based Prostate-specific Membrane Antigenradioguided Surgery in Recurrent Prostate Cancer. Eur Urol. 2019;75(4):659–666. https://doi.org/10.1016/j.eururo.2018.03.013.
- Michel R, Hofer C. Radiation safety precautions for sentinel lymph node procedures. Health Phys. 2004;86(2 Suppl):S35–37. https://doi.org/10.1097/00004032-200402001-00011.
- 100. Piert M, Burian M, Meisetschläger G, Stein HJ, Ziegler S, Nährig J, Picchio M, Buck A, Siewert JR, Schwaiger M. Positron detection for the intraoperative localisation of cancer deposits. Eur J Nucl Med Mol Imaging. 2007;34(10):1534–1544. https://doi.org/10.1007/s00259-007-0430-5.
- Hoffman EJ, Tornai MP, Janecek M, Patt BE, Iwanczyk JS. Intraoperative probes and imaging probes. Eur J Nucl Med. 1999;26(8):913–935. https://doi.org/10.1007/s002590050468.
- 102. van der Poel HG, Meershoek P, Grivas N, KleinJan G, van Leeuwen FWB, Horenblas S. Sentinel node biopsy and lymphatic mapping in penile and prostate cancer. Urologe A. 2017;56(1):13–17. https://doi.org/10.1007/s00120-016-0270-7.
- 103. Rossetti D, Vitale SG, Tropea A, Biondi A, Lagan AS. New procedures for the identification of sentinel lymph node: shaping the horizon of future management in early stage uterine cervical cancer. Updates Surg. 2017;69(3):383–388. https://doi.org/10.1007/s13304-017-0456-6.
- 104. Gunelli R, Fiori M, Salaris C, Salomone U, Urbinati M, Vici A, Zenico T, Bertocco M. The role of intraoperative ultrasound in small renal mass robotic enucleation. Arch Ital Urol Androl. 2016;88(4):311–313. https://doi.org/10.4081/aiua.2016.4.311.
- 105. Collamati F, van Oosterom MN, De Simoni M, Faccini R, Fischetti M, Mancini Terracciano C, Mirabelli R, Moretti R, Heuvel JO, Solfaroli Camillocci E, van Beurden F, van der Poel HG, Valdes Olmos RA, van Leeuwen PJ, van Leeuwen FWB, Morganti S. A DROP-IN beta probe for robot-assisted 68Ga-PSMA radioguided surgery: first ex vivo technology evaluation using prostate cancer specimens. EJNMMI Res. 2020;10(1):92. https://doi.org/10.1186/s13550-020-00682-6.
- 106. Heller S, Zanzonico P. Nuclear probes and intraoperative gamma cameras. Semin Nucl Med. 2011;41(3):166–181. https://doi.org/10.1053/j.semnuclmed.2010.12.004.
- 107. Hellingman D, Vidal-Sicart S Herrmann K, Nieweg OE, Povoski SP, (eds). 2016. The Use of Intraoperative Small and Large Field of View Gamma Cameras for Radioguided Surgery. Cham: Springer International Publishing.
- Tsuchimochi M, Hayama K. Intraoperative gamma cameras for radioguided surgery: technical characteristics, performance parameters, and clinical applications. Phys Med. 2013; 29(2):126–138. https://doi.org/10.1016/j.ejmp.2012.05.002.
- Gitsch E, Philipp K, Pateisky N. Intraoperative lymph scintigraphy during radical surgery for cervical cancer. J Nucl Med. 1984;25(4):486–489.
- 110. Vermeeren L, Valdés Olmos RA, Meinhardt W, Bex A, van der Poel HG, Vogel WV, Sivro F, Hoefnagel CA, Horenblas S. Intraoperative radioguidance with a portable gamma camera: a novel technique for laparoscopic sentinel node localisation in urological malignancies. Eur J Nucl Med Mol Imaging. 2009;36(7):1029–1036. https://doi.org/10.1007/s00259-009-1100-6.
- 111. Vermeeren L, Meinhardt W, Bex A, van der Poel HG, Vogel WV, Hoefnagel CA, Horenblas S, Valdés Olmos RA. Paraaortic sentinel lymph nodes: toward optimal

- detection and intraoperative localization using SPECT/CT and intraoperative real-time imaging. J Nucl Med. 2010;51(3):376–382. https://doi.org/10.2967/jnumed.109.071779.
- 112. Kang HG, Song SH, Han YB, Lee H-Y, Kim KM, Hong SJ. Proof-of-concept of a multimodal laparoscope for simultaneous NIR/gamma/visible imaging using wavelength division multiplexing. Opt Express. 2018;26(7):8325–8339. https://doi.org/10.1364/OE.26.008325.
- 113. Wendler T, Herrmann K, Schnelzer A, Lasser T, Traub J, Kutter O, Ehlerding A, Scheidhauer K, Schuster T, Kiechle M, Schwaiger M, Navab N, Ziegler SI, Buck AK. First demonstration of 3-D lymphatic mapping in breast cancer using freehand SPECT. Eur J Nucl Med Mol Imaging. 2010;37(8):1452–1461. https://doi.org/10.1007/s00259-010-1430-4.
- 114. Freesmeyer M, Opfermann T, Winkens T. Hybrid integration of real-time US and freehand SPECT: proof of concept in patients with thyroid diseases. Radiology. 2014;271(3):856–861. https://doi.org/10.1148/radiol.14132415.
- Freesmeyer M, Winkens T, Opfermann T, Elsner P, Runnebaum I, Darr A. Real-time ultrasound and freehand-SPECT. Experiences with sentinel lymph node mapping. Nuklearmedizin. 2014;53(6):259–264. https://doi.org/10.3413/Nukmed-0680-14-06.
- 116. Bluemel C, Safak G, Cramer A, Wöckel A, Gesierich A, Hartmann E, Schmid J-S, Kaiser F, Buck AK, Herrmann K. Fusion of freehand SPECT and ultrasound: First experience in preoperative localization of sentinel lymph nodes. Eur J Nucl Med Mol Imaging. 2016;43(13):2304–2312. https://doi.org/10.1007/s00259-016-3443-0.
- 118. Monge F, Shakir DI, Lejeune F, Morandi X, Navab N, Jannin P. Acquisition models in intraoperative positron surface imaging. Int J Comput Assist Radiol Surg. 2017;12(4):691–703. https://doi.org/10.1007/s11548-016-1487-z.
- 119. Shakir DI, Okur A, Hart A, Matthies P, Ziegler SI, Essler M, Lasser T, Navab N. Towards intra-operative PET for head and neck cancer: lymph node localization using high-energy probes. Med Image Comput Comput Assist Interv. 2012;15(Pt 1):430–437. https://doi.org/10.1007/978-3-642-33415-3\_53.
- 120. Frisch B. Combining endoscopic ultrasound with Time-Of-Flight PET: The EndoTOFPET-US Project. Nuclear Instrum Methods Phys Res Sect A: Accel Spectrometers, Detect Assoc Equipment. 2013;732:577–580. https://doi.org/10.1016/j.nima.2013.05.027.
- 121. Liyanaarachchi MR, Shimazoe K, Takahashi H, Nakagawa K, Kobayashi E, Sakuma I. Development and evaluation of a prototype detector for an intraoperative laparoscopic coincidence imaging system with PET tracers. Int J CARS. 2021;16(1):29–39. https://doi.org/10.1007/s11548-020-02282-0.
- 122. Markus A, Ray ASC, Bolla D, Müller J, Diener P-A, Wendler T, Hornung R. Sentinel lymph node biopsy in endometrial and cervical cancers using freehand SPECTfirst experiences. Gynecol Surg. 2016;13(4):499–506. https://doi.org/10.1007/s10397-016-0969-x.
- 123. Müller J, Putora PM, Schneider T, Zeisel C, Brutsche M, Baty F, Markus A, Kick J. Handheld single photon emission computed tomography (handheld SPECT) navigated video-assisted thoracoscopic surgery of computer tomography-guided radioactively marked pulmonary lesions. Interact Cardiovasc Thorac Surg. 2016;23(3):345–350. https://doi.org/10.1093/icvts/ivw136.
- 124. Fuerst B, Sprung J, Pinto F, Frisch B, Wendler T, Simon H, Mengus L, van den Berg NS, van der Poel HG, van



- Leeuwen FWB, Navab N. First Robotic SPECT for Minimally Invasive Sentinel Lymph Node Mapping. IEEE Trans Med Imaging. 2016;35(3):830–838. https://doi.org/10.1109/TMI.2015.2498125.
- 125. Huang B, Tsai Y-Y, Cartucho J, Vyas K, Tuch D, Giannarou S, Elson DS. Tracking and visualization of the sensing area for a tethered laparoscopic gamma probe. Int J CARS. 2020;15(8):1389–1397. https://doi.org/10.1007/s11548-020-02205-z.
- 126. Matthies P, Sharma K, Okur A, Gardiazabal J, Vogel J, Lasserl T, Navab N. First use of mini gamma cameras for intra-operative robotic SPECT reconstruction. Med Image Comput Comput Assist Interv;16(Pt 1):163–170. https://doi.org/10.1007/978-3-642-40811-3\_21.
- 127. Matthies P, Gardiazabal J, Okur A, Vogel J, Lasser T, Navab N. Mini gamma cameras for intra-operative nuclear tomographic reconstruction. Med Image Anal. 2014;18(8):1329– 1336. https://doi.org/10.1016/j.media.2014.04.009.
- 128. Gardiazabal J, Matthies P, Vogel J, Frisch B, Navab N, Ziegler S, Lasser T. Flexible mini gamma camera reconstructions of extended sources using step and shoot and list mode. Med Phys. 2016;43(12):6418. https://doi.org/10.1118/1.4966700.
- 129. Gardiazabal J, Esposito M, Matthies P, Okur A, Vogel J, Kraft S, Frisch B, Lasser T, Navab N. Towards personalized interventional SPECT-CT imaging. Med Image Comput Comput Assist Interv. 2014;17(Pt 1):504–511. https://doi.org/10.1007/978-3-319-10404-1\_63.
- 130. Rietbergen DDD, Meershoek P, van Oosterom MN, Roestenberg M, van Erkel AR, Smit F, van der Hage JA, Valdés Olmos RA, van Leeuwen FWB. Freehand-SPECT with 99mTc-HDP as tool to guide percutaneous biopsy of skeletal lesions detected on bone scintigraphy. Rev Esp Med Nucl Imagen Mol. 2019;38(4):218–223. https://doi.org/10.1016/j.remn.2019.01.003.
- Schilling C, Gnansegaran G, Thavaraj S, McGurk M. Intraoperative sentinel node imaging versus SPECT/CT in oral cancer A blinded comparison. Eur J Surg Oncol. 2018;44(12):1901–1907. https://doi.org/10.1016/j.ejso.2018.08.026.
- 132. Jeremiasse B, van den Bosch CH, Wijnen MWHA, Terwisscha van Scheltinga CEJ, Fiocco MF, van der Steeg AFW. Systematic review and meta-analysis concerning near-infrared imaging with fluorescent agents to identify the sentinel lymph node in oncology patients. Eur J Surg Oncol. 2020;46(11):2011–2022. https://doi.org/10.1016/j.ejso.2020.07.012.
- 133. Lee JYK, Cho SS, Stummer W, Tanyi JL, Vahrmeijer AL, Rosenthal E, Smith B, Henderson E, Roberts DW, Lee A, Hadjipanayis CG, Bruce JN, Newman JG, Singhal S. Review of clinical trials in intraoperative molecular imaging during cancer surgery. J Biomed Opt. 2019;24(12):1–8. https://doi.org/10.1117/1.JBO.24.12.120901.
- 134. Lee G-W, Park J-Y, Kim D-Y, Suh D-S, Kim J-H, Kim Y-M, Kim Y-T, Nam J-H. Usefulness of sentinel lymph node mapping using indocyanine green and fluorescent imaging in the diagnosis of lymph node metastasis in endometrial cancer. J Obstet Gynaecol:1–7. 2020. https://doi.org/10.1080/01443615.2020.1787965.
- 135. Mehdorn A-S, Beckmann JH, Braun F, Becker T, Egberts J-H. Usability of Indocyanine Green in Robot-Assisted Hepatic Surgery. J Clin Med. 2021;10(3). https://doi.org/10.3390/jcm10030456.
- 136. Ko YJ, Kim WJ, Kim K, Kwon IC. Advances in the strategies for designing receptor-targeted molecular imaging probes for cancer research. J Control Release. 2019;305:1–17. https://doi.org/10.1016/j.jconrel.2019.04.030.

- 137. Stoffels I, Dissemond J, Pöppel T, Schadendorf D, Klode J. Intraoperative Fluorescence Imaging for Sentinel Lymph Node Detection: Prospective Clinical Trial to Compare the Usefulness of Indocyanine Green vs Technetium Tc 99m for Identification of Sentinel Lymph Nodes. JAMA Surg. 2015;150(7):617–623. https://doi.org/10.1001/jamasurg.2014.3502.
- 138. van Beurden F, van Willigen DM, Vojnovic B, van Oosterom MN, Brouwer OR, der Poel HG, Kobayashi H, van Leeuwen FWB, Buckle T. Multi-Wavelength Fluorescence in Image-Guided Surgery, Clinical Feasibility and Future Perspectives. Mol Imaging. 2020;19:1536012120962333. https://doi.org/10.1177/1536012120962333.
- 139. Meershoek P, KleinJan GH, Oosterom MNv, Wit EMK, Willigen DMv, Bauwens KP, Gennep EJv, Mottrie AM, Poel HGvd, Leeuwen FWBv. Multispectral-Fluorescence Imaging as a Tool to Separate Healthy from Disease-Related Lymphatic Anatomy During Robot-Assisted Laparoscopy. J Nucl Med. 2018;59(11):1757–1760. https://doi.org/10.2967/jnumed.118.211888.
- 140. van Willigen DM, van den Berg NS, Buckle T, Klein-Jan GH, Hardwick JC, van der Poel HG, van Leeuwen FWB. Multispectral fluorescence guided surgery; a feasibility study in a phantom using a clinical-grade laparoscopic camera system. Am J Nucl Med Mol Imaging. 2017;7(3):138– 147.
- 141. Francaviglia N, Iacopino DG, Costantino G, Villa A, Impallaria P, Meli F, Maugeri R. Fluorescein for resection of high-grade gliomas: A safety study control in a single center and review of the literature. Surg Neurol Int. 2017;8. https://doi.org/10.4103/sni.sni\_89\_17.
- 142. Ladurner R, Lerchenberger M, Al Arabi N, Gallwas JKS, Stepp H, Hallfeldt KKJ. Parathyroid AutofluorescenceHow Does It Affect Parathyroid and Thyroid Surgery? A 5 Year Experience. Molecules. 2019;24(14):2560. https://doi.org/10.3390/molecules24142560.
- 143. Kriegmair MC, Rother J, Grychtol B, Theuring M, Ritter M, Günes C, Michel MS, Deliolanis NC, Bolenz C. Multi-parametric Cystoscopy for Detection of Bladder Cancer Using Real-time Multispectral Imaging. Eur Urol. 2020;77(2):251–259. https://doi.org/10.1016/j.eururo.2019.08.024.
- 144. Kaibori M, Matsui K, Ishizaki M, Iida H, Okumura T, Sakaguchi T, Inoue K, Ikeura T, Asano H, Kon M. Intraoperative Detection of Superficial Liver Tumors by Fluorescence Imaging Using Indocyanine Green and 5-aminolevulinic Acid. Anticancer Res. 2016;36(4):1841–1849. https://ar.iiarjournals.org/content/36/4/1841.
- 145. Laios A, Volpi D, Tullis IDC, Woodward M, Kennedy S, Pathiraja PNJ, Haldar K, Vojnovic B, Ahmed AA. A prospective pilot study of detection of sentinel lymph nodes in gynaecological cancers using a novel near infrared fluorescence imaging system. BMC Res Notes. 2015;8(1):608. https://doi.org/10.1186/s13104-015-1576-z.
- 146. Meershoek P, KleinJan GH, van Willigen DM, Bauwens KP, Spa SJ, van Beurden F, van Gennep EJ, Mottrie AM, van der Poel HG, Buckle T, van Leeuwen FWB, van Oosterom MN. Multi-wavelength fluorescence imaging with a da Vinci Fireflya technical look behind the scenes. J Robotic Surg. 2020. https://doi.org/10.1007/s11701-020-01170-8.
- 147. Hu Z, Fang C, Li B, Zhang Z, Cao C, Cai M, Su S, Sun X, Shi X, Li C, Zhou T, Zhang Y, Chi C, He P, Xia X, Chen Y, Gambhir SS, Cheng Z, Tian J. First-in-human liver-tumour surgery guided by multispectral fluorescence imaging in the visible and near-infrared-I/II windows. Nat Biomed Eng. 2020;4(3):259–271. https://doi.org/10.1038/s41551-019-0494-0.



- 148. Vargas SH, Ghosh SC, Azhdarinia A. New Developments in Dual-Labeled Molecular Imaging Agents. J Nucl Med. 2019;60(4):459–465. https://doi.org/10.2967/jnumed.118. 213488.
- 149. van der Poel HG, Buckle T, Brouwer OR, Valdés Olmos RA, van Leeuwen FWB. Intraoperative Laparoscopic Fluorescence Guidance to the Sentinel Lymph Node in Prostate Cancer Patients: Clinical Proof of Concept of an Integrated Functional Imaging Approach Using a Multimodal Tracer. Eur Urol. 2011;60(4):826–833. https://doi.org/10.1016/j.eururo.2011.03.024.
- 150. KleinJan GH, van Werkhoven E, van den Berg NS, Karakullukcu MB, Zijlmans HJMAA, van der Hage JA, van de Wiel BA, Buckle T, Klop WMC, Horenblas S, Valdés Olmos RA, van der Poel HG, van Leeuwen FWB. The best of both worlds: a hybrid approach for optimal pre- and intraoperative identification of sentinel lymph nodes. Eur J Nucl Med Mol Imaging. 2018;45(11):1915–1925. https://doi.org/10.1007/s00259-018-4028-x.
- 151. Dell'Oglio P, de Vries HM, Mazzone E, KleinJan GH, Donswijk ML, van der Poel HG, Horenblas S, van Leeuwen FWB, Brouwer OR. Hybrid Indocyanine Green99mTc-nanocolloid for Single-photon Emission Computed Tomography and Combined Radio- and Fluorescence-guided Sentinel Node Biopsy in Penile Cancer: Results of 740 Inguinal Basins Assessed at a Single Institution. Eur Urol. 2020;78(6):865–872. https://doi.org/10.1016/j.eururo.2020.09.007.
- 152. Christensen A, Juhl K, Charabi B, Mortensen J, Kiss K, Kær A, von Buchwald C. Feasibility of Real-Time Near-Infrared Fluorescence Tracer Imaging in Sentinel Node Biopsy for Oral Cavity Cancer Patients. Ann Surg Oncol. 2016;23(2):565–572. https://doi.org/10.1245/s10434-015-4883-7.
- 153. Paredes P, Vidal-Sicart S, Campos F, Tapias A, SÁnchez N, Martínez S, Carballo L, Pahisa J, Torné A, Ordi J, Carmona F, Lomea F. Role of ICG-99mTc-nanocolloid for sentinel lymph node detection in cervical cancer: a pilot study. Eur J Nucl Med Mol Imaging. 2017;44(11):1853–1861. https://doi.org/10.1007/s00259-017-3706-4.
- 154. Schaafsma BE, Verbeek FPR, Rietbergen DDD, van der Hiel B, van der Vorst JR, Liefers G-J, Frangioni JV, van de Velde CJH, van Leeuwen FWB, Vahrmeijer AL. Clinical trial of combined radio- and fluorescence-guided sentinel lymph node biopsy in breast cancer. Br J Surg. 2013;100(8):1037–1044. https://doi.org/10.1002/bjs.9159.
- 155. Stoffels I, Leyh J, Pöppel T, Schadendorf D, Klode J. Evaluation of a radioactive and fluorescent hybrid tracer for sentinel lymph node biopsy in head and neck malignancies: prospective randomized clinical trial to compare ICG-(99m)Tc-nanocolloid hybrid tracer versus (99m)Tc-nanocolloid. Eur J Nucl Med Mol Imaging. 2015;42(11):1631–1638. https://doi.org/10.1007/s00259-015-3093-7.
- 156. Santini C, Kuil J, Bunschoten A, Pool S, Blois E, Ridwan Y, Essers J, Bernsen MR, Leeuwen FWBv, Jong Md. Evaluation of a Fluorescent and Radiolabeled Hybrid Somatostatin Analog In Vitro and in Mice Bearing H69 Neuroendocrine Xenografts. J Nucl Med. 2016;57(8):1289–1295. https://doi.org/10.2967/jnumed.115.164970.
- 157. Schottelius M, Wurzer A, Wissmiller K, Beck R, Koch M, Gorpas D, Notni J, Buckle T, Oosterom MN, Steiger K, Ntziachristos V, Schwaiger M, Leeuwen FWBv, Wester H-J. Synthesis and Preclinical Characterization of the PSMA-Targeted Hybrid Tracer PSMA-I&F for Nuclear and Fluorescence Imaging of Prostate Cancer. J Nucl Med. 2019;60(1):71–78. https://doi.org/10.2967/jnumed.118.212720.

- 158. Johnson L, Pinder SE, Douek M. Deposition of superparamagnetic iron-oxide nanoparticles in axillary sentinel lymph nodes following subcutaneous injection. Histopathology. 2013;62(3):481–486. https://doi.org/10.1111/his.12019.
- 159. Thompson W, ArgÁez C. Magnetic Localization System for Sentinel Lymph Node Biopsy: A Review of the Diagnostic Accuracy, Cost-Effectiveness, and Guidelines, CADTH Rapid Response Reports. Ottawa: Canadian Agency for Drugs and Technologies in Health; 2020. http://www.ncbi.nlm.nih.gov/ books/NBK562944/.
- 160. Jedryka MA, Klimczak P, Kryszpin M, Matkowski R. Superparamagnetic iron oxide: a novel tracer for sentinel lymph node detection in vulvar cancer. Int J Gynecol Cancer. 2020;30(9):1280–1284. https://doi.org/10.1136/ijgc-2020-001458
- 161. Winter A, Kowald T, Paulo TS, Goos P, Engels S, Gerullis H, Schiffmann J, Chavan A, Wawroschek F. Magnetic resonance sentinel lymph node imaging and magnetometer-guided intraoperative detection in prostate cancer using superparamagnetic iron oxide nanoparticles. Int J Nanomedicine. 2018;13:6689–6698. https://doi.org/10.2147/IJN.S173182.
- 162. Winter A, Kowald T, Engels S, Wawroschek F. Magnetic Resonance Sentinel Lymph Node Imaging and Magnetometer-Guided Intraoperative Detection in Penile Cancer, using Superparamagnetic Iron Oxide Nanoparticles: First Results. Urol Int. 2020;104(3-4):177–180. https://doi.org/10.1159/000502017.
- 163. Pi nro-Madrona A, NicolÁs-Ruiz F, Rull-Ortu no R, Vidal-Sicart S, Caba ns-Montero J, Rioja-Martín ME, Rodríguez-FernÁndez R, Gil-Olarte MA, GonzÁlez-García B, SÁnchez JH-G. Correlation between ferromagnetic and isotopic tracers for sentinel lymph node detection in cutaneous melanoma: IMINEM study. J Surg Oncol. 2020. https://doi.org/10.1002/jso.26303.
- 164. Imai K, Kawaharada Y, Ogawa J-I, Saito H, Kudo S, Takashima S, Saito Y, Atari M, Ito A, Terata K, Yoshino K, Sato Y, Motoyama S, Minamiya Y. Development of a New Magnetometer for Sentinel Lymph Node Mapping Designed for Video-Assisted Thoracic Surgery in Non-Small Cell Lung Cancer. Surg Innov. 2015;22(4):401–405. https://doi.org/10.1177/1553350615585421.
- 165. Forte S, Kubik-Huch RA, Leo C. Improvement in breast magnetic resonance imaging after a sentinel procedure for breast cancer with superparamagnetic tracers. Eur J Radiol Open. 2019;6:215–219. https://doi.org/10.1016/j.ejro.2019.05.006.
- 166. Frantellizzi V, Conte M, Pontico M, Pani A, Pani R, De Vincentis G. New Frontiers in Molecular Imaging with Superparamagnetic Iron Oxide Nanoparticles (SPIONs): Efficacy, Toxicity, and Future Applications. Nucl Med Mol Imaging. 2020; 54(2):65–80. https://doi.org/10.1007/s13139-020-00635-w.
- 167. Manohar S, Vaartjes SE, van Hespen JCG, Klaase JM, van den Engh FM, Steenbergen W, van Leeuwen TG. Initial results of in vivo non-invasive cancer imaging in the human breast using near-infrared photoacoustics. Opt Express. 2007;15(19):12277– 12285. https://doi.org/10.1364/oe.15.012277.
- 168. Ermilov SA, Khamapirad T, Conjusteau A, Leonard MH, Lacewell R, Mehta K, Miller T, Oraevsky AA. Laser optoacoustic imaging system for detection of breast cancer. J Biomed Opt. 2009;14(2):024007. https://doi.org/10.1117/1.3086616.
- 169. Razansky D, Harlaar NJ, Hillebrands JL, Taruttis A, Herzog E, Zeebregts CJ, van Dam GM, Ntziachristos V. Multispectral optoacoustic tomography of matrix metalloproteinase activity in vulnerable human carotid plaques. Mol Imaging Biol. 2012;14(3):277–285. https://doi.org/10.1007/s11307-011-0502-6.



- 170. Stoffels I, Jansen P, Petri M, Goerdt L, Brinker TJ, Griewank KG, Poeppel TD, Schadendorf D, Klode J. Assessment of Nonradioactive Multispectral Optoacoustic Tomographic Imaging With Conventional Lymphoscintigraphic Imaging for Sentinel Lymph Node Biopsy in Melanoma. JAMA Netw Open. 2019;2(8):e199020. https://doi.org/10.1001/jamanetworkopen.2019.9020.
- 171. Diot G, Metz S, Noske A, Liapis E, Schroeder B, Ovsepian SV, Meier R, Rummeny E, Ntziachristos V. Multispectral Optoacoustic Tomography (MSOT) of Human Breast Cancer. Clin Cancer Res. 2017;23(22):6912–6922. https://doi.org/10.1158/1078-0432.CCR-16-3200.
- 172. Attia ABE, Chuah SY, Razansky D, Ho CJH, Malempati P, Dinish US, Bi R, Fu CY, Ford SJ, Lee JS-S, Tan MWP, Olivo M, Thng STG. Noninvasive real-time characterization of non-melanoma skin cancers with handheld optoacoustic probes. Photoacoustics. 2017;7:20–26. https://doi.org/10.1016/j.pacs.2017.05.003.
- 173. Nitkunanantharajah S, Haedicke K, Moore TB, Manning JB, Dinsdale G, Berks M, Taylor C, Dickinson MR, Jüstel D, Ntziachristos V, Herrick AL, Murray AK. Three-dimensional optoacoustic imaging of nailfold capillaries in systemic sclerosis and its potential for disease differentiation using deep learning. Sci Rep. 2020;10(1):16444. https://doi.org/10.1038/s41598-020-73319-2.
- 174. Chlis N-K, Karlas A, Fasoula N-A, Kallmayer M, Eckstein H-H, Theis FJ, Ntziachristos V, Marr C. A sparse deep learning approach for automatic segmentation of human vasculature in multispectral optoacoustic tomography. Photoacoustics. 2020;20:100203. https://doi.org/10.1016/j.pacs.2020.100203.
- 175. Haka AS, Volynskaya Z, Gardecki JA, Nazemi J, Lyons J, Hicks D, Fitzmaurice M, Dasari RR, Crowe JP, Feld MS. In vivo margin assessment during partial mastectomy breast surgery using raman spectroscopy. Cancer Res. 2006;66(6):3317–3322. https://doi.org/10.1158/0008-5472.CAN-05-2815.
- 176. Palermo A, Fosca M, Tabacco G, Marini F, Graziani V, Santarsia MC, Longo F, Lauria A, Cesareo R, Giovannoni I, Taffon C, Rocchia M, Manfrini S, Crucitti P, Pozzilli P, Crescenzi A, Rau JV. Raman Spectroscopy Applied to Parathyroid Tissues: A New Diagnostic Tool to Discriminate Normal Tissue from Adenoma. Anal Chem. 2018;90(1):847–854. https://doi.org/10.1021/acs.analchem.7b03617.
- 177. Horsnell J, Stonelake P, Christie-Brown J, Shetty G, Hutchings J, Kendall C, Stone N. Raman spectroscopy—a new method for the intra-operative assessment of axillary lymph nodes. Analyst. 2010;135(12):3042–3047. https://doi.org/10.1039/c0an00527d.
- 178. Barroso EM, Smits RWH, Bakker Schut TC, ten Hove I, Hardillo JA, Wolvius EB, Baatenburg de Jong RJ, Koljenovi S, Puppels GJ. Discrimination between oral cancer and healthy tissue based on water content determined by Raman spectroscopy. Anal Chem. 2015;87(4):2419–2426. https://doi.org/10.1021/ac504362y.
- 179. Barroso EM, Ten Hove I, Bakker Schut TC, Mast H, van Lanschot CGF, Smits RWH, Caspers PJ, Verdijk R, Noordhoek Hegt V, Baatenburg de Jong RJ, Wolvius EB, Puppels GJ, Koljenovi S. Raman spectroscopy for assessment of bone resection margins in mandibulectomy for oral cavity squamous cell carcinoma. Eur J Cancer. 2018;92:77–87. https://doi.org/10.1016/j.ejca.2018.01.068.
- 180. Jermyn M, Mok K, Mercier J, Desroches J, Pichette J, Saint-Arnaud K, Bernstein L, Guiot M-C, Petrecca K, Leblond F. Intraoperative brain cancer detection with Raman spectroscopy in humans. Sci Transl Med. 2015;7(274):274ra19. https://doi.org/10.1126/scitranslmed.aaa2384.

- 181. Desroches J, Jermyn M, Mok K, Lemieux-Leduc C, Mercier J, St-Arnaud K, Urmey K, Guiot M-C, Marple E, Petrecca K, Leblond F. Characterization of a Raman spectroscopy probe system for intraoperative brain tissue classification. Biomed Opt Express. 2015;6(7):2380–2397. https://doi.org/10.1364/BOE.6.002380.
- 182. Rau JV, Fosca M, Graziani V, Taffon C, Rocchia M, Caricato M, Pozzilli P, Onetti Muda A, Crescenzi A. Proof-of-concept Raman spectroscopy study aimed to differentiate thyroid follicular patterned lesions. Sci Rep. 2017;7(1):14970. https://doi.org/10.1038/s41598-017-14872-1.
- 183. Jermyn M, Desroches J, Mercier J, Tremblay M-A, St-Arnaud K, Guiot M-C, Petrecca K, Leblond F. Neural networks improve brain cancer detection with Raman spectroscopy in the presence of operating room light artifacts. J Biomed Opt. 2016;21(9):94002. https://doi.org/10.1117/1.JBO.21.9.094002.
- 184. Yu M, Yan H, Xia J, Zhu L, Zhang T, Zhu Z, Lou X, Sun G, Dong M. Deep convolutional neural networks for tongue squamous cell carcinoma classification using Raman spectroscopy. Photodiagnosis Photodyn Ther. 2019;26:430–435. https://doi.org/10.1016/j.pdpdt.2019.05.008.
- 185. Hollon TC, Pandian B, Urias E, Save AV, Adapa AR, Srinivasan S, Jairath NK, Farooq Z, Marie T, Al-Holou WN, Eddy K, Heth JA, Khalsa SSS, Conway K, Sagher O, Bruce JN, Canoll P, Freudiger CW, Camelo-Piragua S, Lee H, Orringer DA. Rapid, label-free detection of diffuse glioma recurrence using intraoperative stimulated Raman histology and deep neural networks. Neuro Oncol. 2020. https://doi.org/10.1093/neuonc/noaa162.
- 186. Livermore LJ, Isabelle M, Bell IM, Edgar O, Voets NL, Stacey R, Ansorge O, Vallance C, Plaha P. Raman spectroscopy to differentiate between fresh tissue samples of glioma and normal brain: a comparison with 5-ALA-induced fluorescence-guided surgery. J Neurosurg:1–11. 2020. https://doi.org/10.3171/2020.5.JNS20376.
- 187. Pouw B, de Wit-van der Veen LJ, van der Hage JA, Vrancken Peeters M-JTFD, Wesseling J, Stokkel MPM, Valdés Olmos RA. Radio-guided seed localization for breast cancer excision: an ex-vivo specimen-based study to establish the accuracy of a freehand-SPECT device in predicting resection margins. Nucl Med Commun. 2014;35(9):961–966. https://doi.org/10.1097/MNM.0000000000000159.
- 188. Bluemel C, Cramer A, Grossmann C, Kajdi GW, Malzahn U, Lamp N, Langen H-J, Schmid J, Buck AK, Grimminger H-J, Herrmann K. iROLL: does 3-D radioguided occult lesion localization improve surgical management in early-stage breast cancer? Eur. J. Nucl. Med. Mol. Imaging. 2015. https://doi.org/10.1007/s00259-015-3121-7.
- 189. van Oosterom MN, Meershoek P, Welling MM, Pinto F, Matthies P, Simon H, Wendler T, Navab N, van de Velde CJH, van der Poel HG, van Leeuwen FWB. Extending the Hybrid Surgical Guidance Concept With Freehand Fluorescence Tomography. IEEE Trans Med Imaging. 2020;39(1):226–235. https://doi.org/10.1109/TMI.2019.2924254.
- 190. Ankersmit M, Hoekstra OS, van Lingen A, Bloemena E, Jacobs MJM, Vugts DJ, Bonjer HJ, van Dongen GMS, Meijerink WJHJ. Perioperative PET/CT lymphoscintigraphy and fluorescent real-time imaging for sentinel lymph node mapping in early staged colon cancer. Eur J Nucl Med Mol Imaging. 2019;46(7):1495–1505. https://doi.org/10.1007/s00259-019-04284-w.
- 191. Povoski SP, Hall NC, Murrey DA, Chow AZ, Gaglani JR, Bahnson EE, Mojzisik CM, Kuhrt MP, Hitchcock CL, Knopp MV, Martin EW. Multimodal imaging and detection



- approach to 18F-FDG-directed surgery for patients with known or suspected malignancies: a comprehensive description of the specific methodology utilized in a single-institution cumulative retrospective experience. World J Surg Oncol. 2011;9:152. https://doi.org/10.1186/1477-7819-9-152.
- Alsadoun N, Devouassoux-Shisheboran M. Pathological process for sentinel lymph node. Bull Cancer. 2020;107(6):642–652 (fre). https://doi.org/10.1016/j.bulcan.2019.11.003.
- 193. Kwon S-Y, Miller SJ. Mohs surgery for melanoma in situ. Dermatol Clin. 2011;29(2):175–183, vii–viii. https://doi.org/10.1016/j.det.2011.01.001.
- 194. St John ER, Al-Khudairi R, Ashrafian H, Athanasiou T, Takats Z, Hadjiminas DJ, Darzi A, Leff DR. Diagnostic Accuracy of Intraoperative Techniques for Margin Assessment in Breast Cancer Surgery: A Meta-analysis. Ann Surg. 2017;265(2):300–310. https://doi.org/10.1097/SLA.0000000000001897.
- Mannelli G, Comini LV, Piazza C. Surgical margins in oral squamous cell cancer: intraoperative evaluation and prognostic impact. Curr Opin Otolaryngol Head Neck Surg. 2019;27(2):98– 103. https://doi.org/10.1097/MOO.0000000000000516.
- 196. Sighinolfi MC, Eissa A, Spandri V, Puliatti S, Micali S, Reggiani Bonetti L, Bertoni L, Bianchi G, Rocco B. Positive surgical margin during radical prostatectomy: overview of sampling methods for frozen sections and techniques for the secondary resection of the neurovascular bundles. BJU Int. 2020;125(5):656–663. https://doi.org/10.1111/bju.15024.
- 197. Dinneen E, Haider A, Grierson J, Freeman A, Oxley J, Briggs T, Nathan S, Williams NR, Brew-Graves C, Persad R, Aning J, Jamieson C, Ratynska M, Ben-Salha I, Ball R, Clow R, Allen C, Heffernan-Ho D, Kelly J, Shaw G. NeuroSAFE frozen section during robot-assisted radical prostatectomy (RARP): Peri-operative and Histopathological Outcomes from the NeuroSAFE PROOF Feasibility Randomised Controlled Trial. BJU Int. 2020. https://doi.org/10.1111/bju.15256.
- 198. Kim Y-G, Kim S, Cho CE, Song IH, Lee HJ, Ahn S, Park SY, Gong G, Kim N. Effectiveness of transfer learning for enhancing tumor classification with a convolutional neural network on frozen sections. Sci Rep. 2020;10(1):21899. https://doi.org/10.1038/s41598-020-78129-0.
- 199. Marsh JN, Matlock MK, Kudose S, Liu T-C, Stappenbeck TS, Gaut JP, Swamidass SJ. Deep Learning Global Glomerulosclerosis in Transplant Kidney Frozen Sections. IEEE Trans Med Imaging. 2018;37(12):2718–2728. https://doi.org/10.1109/TMI.2018.2851150.
- 200. Shi F, Liang Z, Zhang Q, Wang C, Liu X. The performance of one-step nucleic acid amplification assay for intraoperative detection of sentinel lymph node macrometastasis in breast cancer: An updated meta-analysis. Breast. 2018;39:39–45. https://doi.org/10.1016/j.breast.2018.03.005.
- 201. Zhou M, Wang X, Jiang L, Chen X, Bao X, Chen X. The diagnostic value of one step nucleic acid amplification (OSNA) in differentiating lymph node metastasis of tumors: A systematic review and meta-analysis. Int J Surg. 2018;56:49–56. https://doi.org/10.1016/j.ijsu.2018.05.010.
- 202. Rauscher I, Horn T, Eiber M, Gschwend JE, Maurer T. Novel technology of molecular radio-guidance for lymph node dissection in recurrent prostate cancer by PSMA-ligands. World J Urol. 2018;36(4):603–608. https://doi.org/10.1007/s00345-018-2200-3.
- 203. Metser U, McVey R, Ferguson SE, Halankar J, Bernardini MQ. Intraoperative lymph node evaluation using 18F-FDG and a hand-held gamma probe in endometrial cancer surgery–a pilot study. Eur J Gynaecol Oncol. 2016;37(3):362–366.

- 204. Gola S, Doyle-Lindrud S. The MarginProbe® System: An Innovative Approach to Reduce the Incidence of Positive Margins Found After Lumpectomy. Clin J Oncol Nurs. 2016;20(6):598–599. https://doi.org/10.1188/16.CJON.598-599.
- Fanjul-Vélez F, Pampín-SuÁrez S, Arce-Diego JL. Application of Classification Algorithms to Diffuse Reflectance Spectroscopy Measurements for Ex Vivo Characterization of Biological Tissues. Entropy (Basel). 2020;22(7). https://doi.org/10.3390/e22070736.
- 206. Paredes P, Vidal-Sicart S, Zanón G, Roé N, Rubí S, Lafuente S, Pavía J, Pons F. Radioguided occult lesion localisation in breast cancer using an intraoperative portable gamma camera: first results. Eur J Nucl Med Mol Imaging. 2008;35(2):230–235. https://doi.org/10.1007/s00259-007-0640-x.
- Vidal-Sicart S, Rioja ME, Paredes P, Keshtgar MR, Valdés Olmos RA. Contribution of perioperative imaging to radioguided surgery. Q J Nucl Med Mol Imaging. 2014;58(2):140–160.
- 208. Ciarrocchi E, Belcari N. Cerenkov luminescence imaging: physics principles and potential applications in biomedical sciences. EJNMMI Phys. 2017;4(1):14. https://doi.org/10.1186/s40658-017-0181-8.
- Das S, Thorek DLJ, Grimm J. Cerenkov imaging. Adv Cancer Res. 2014;124:213–234. https://doi.org/10.1016/B978-0-12-411638-2.00006-9.
- Spinelli AE, Boschi F. Novel biomedical applications of Cerenkov radiation and radioluminescence imaging. Phys Med. 2015;31(2):120–129. https://doi.org/10.1016/j.ejmp.2014. 12.003.
- 211. Glaser AK, Zhang R, Andreozzi JM, Gladstone DJ, Pogue BW. Cherenkov radiation fluence estimates in tissue for molecular imaging and therapy applications. Phys Med Biol. 2015;60(17):6701–6718. https://doi.org/10.1088/0031-9155/60/17/6701.
- 212. Thorek DL, Robertson R, Bacchus WA, Hahn J, Rothberg J, Beattie BJ, Grimm J. Cerenkov imaging a new modality for molecular imaging. Am J Nucl Med Mol Imaging. 2012;2(2):163–173.
- 213. Darr C, Harke NN, Radtke JP, Yirga L, Kesch C, Grootendorst MR, Fendler WP, Costa PF, Rischpler C, Praus C, Haubold J, Reis H, Hager T, Herrmann K, Binse I, Hadaschik B. Intraoperative 68Ga-PSMA Cerenkov Luminescence Imaging for Surgical Margins in Radical Prostatectomy: A Feasibility Study. J Nucl Med. 2020;61(10):1500–1506. https://doi.org/10.2967/jnumed.119.240424.
- 214. Olde Heuvel J, de Wit-van der Veen BJ, van der Poel HG, Bekers EM, Grootendorst MR, Vyas KN, Slump CH, Stokkel MPM. 68Ga-PSMA Cerenkov luminescence imaging in primary prostate cancer: first-in-man series. Eur J Nucl Med Mol Imaging. 2020;47(11):2624–2632. https://doi.org/10.1007/s00259-020-04783-1.
- 215. Chin PTK, Welling MM, Meskers SCJ, Valdes Olmos RA, Tanke H, van Leeuwen FWB. Optical imaging as an expansion of nuclear medicine: Cerenkov-based luminescence vs fluorescence-based luminescence. Eur J Nucl Med Mol Imaging. 2013;40(8):1283–1291. https://doi.org/10.1007/s00259-013-2408-9.
- 216. Yu X, Hu C, Zhang W, Zhou J, Ding Q, Sadiq MT, Fan Z, Yuan Z, Liu L. Feasibility evaluation of micro-optical coherence tomography (μOCT) for rapid brain tumor type and grade discriminations: μOCT images versus pathology. BMC Med Imaging. 2019;19(1):102. https://doi.org/10.1186/s12880-019-0405-6.
- 217. Valdés Olmos RA, Vidal-Sicart S, Giammarile F, Zaknun JJ, Van Leeuwen FW, Mariani G. The GOSTT concept and



- hybrid mixed/virtual/augmented reality environment radioguided surgery. Q J Nucl Med Mol Imaging. 2014;58(2):207–215.
- 218. Wester H-J, Schottelius M. PSMA-Targeted Radiophar-maceuticals for Imaging and Therapy. Semin Nucl Med. 2019;49(4):302–312. https://doi.org/10.1053/j.semnuclmed. 2019.02.008.
- Lei Y, Fu Y, Wang T, Qiu RLJ, Curran WJ, Liu T, Yang X. Deep Learning in Multi-organ Segmentation. 2020. arXiv:2001.10619.
- 220. Alam IS, Steinberg I, Vermesh O, van den Berg NS, Rosenthal EL, van Dam GM, Ntziachristos V, Gambhir SS, Hernot S, Rogalla S. Emerging Intraoperative Imaging Modalities to Improve Surgical Precision. Mol Imaging Biol. 2018;20(5):705–715. https://doi.org/10.1007/s11307-018-1227-6.
- 221. Herrmann K, Nieweg OE, Povoski SP (eds.) Radioguided Surgery: Current Applications and Innovative Directions in Clinical Practice; 2016. Springer International Publishing.
- 222. Azargoshasb S, van Alphen S, Slof LJ, Rosiello G, Puliatti S, van Leeuwen SI, Houwing KM, Boonekamp M, Verhart J, Dell'Oglio P, van der Hage J, van Oosterom MN, van Leeuwen FWB. The click-on gamma probe, a second-generation tethered robotic gamma probe that improves dexterity and

- surgical decision-making. Eur J Nucl Med Molec Imaging. 2021. (in press).
- 223. Leal Ghezzi T, Campos Corleta O. 30 Years of Robotic Surgery. World J Surg. 2016;40(10):2550–2557. https://doi.org/10.1007/s00268-016-3543-9.
- 224. Jiang B, Azad TD, Cottrill E, Zygourakis CC, Zhu AM, Crawford N, Theodore N. New spinal robotic technologies. Front Med. 2019;13(6):723–729. https://doi.org/10.1007/s11684-019-0716-6.
- 225. Smith JA, Jivraj J, Wong R, Yang V. 30 Years of Neurosurgical Robots: Review and Trends for Manipulators and Associated Navigational Systems. Ann Biomed Eng. 2016;44(4):836–846. https://doi.org/10.1007/s10439-015-1475-4.
- 226. Esteban J, Simson W, Requena Witzig S, Rienmüller A, Virga S, Frisch B, Zettinig O, Sakara D, Ryang Y-M, Navab N, Hennersperger C. Robotic ultrasound-guided facet joint insertion. Int J Comput Assist Radiol Surg. 2018;13(6):895–904. https://doi.org/10.1007/s11548-018-1759-x.

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