

Pollen-associated microbiome correlates with pollution parameters and the allergenicity of pollen

Andrea Obersteiner, Stefanie Gilles, Ulrike Frank, Isabelle Beck, Franziska Häring, Dietrich Ernst, Michael Rothballer, Anton Hartmann, Claudia Traidl-Hoffmann, Michael Schmid

Angaben zur Veröffentlichung / Publication details:

Obersteiner, Andrea, Stefanie Gilles, Ulrike Frank, Isabelle Beck, Franziska Häring, Dietrich Ernst, Michael Rothballer, Anton Hartmann, Claudia Traidl-Hoffmann, and Michael Schmid. 2016. "Pollen-associated microbiome correlates with pollution parameters and the allergenicity of pollen." *PLoS ONE* 11 (2): e0149545.
<https://doi.org/10.1371/journal.pone.0149545>.

RESEARCH ARTICLE

Pollen-Associated Microbiome Correlates with Pollution Parameters and the Allergenicity of Pollen

Andrea Obersteiner¹, Stefanie Gilles^{2,3}, Ulrike Frank⁴, Isabelle Beck², Franziska Häring², Dietrich Ernst⁴, Michael Rothballer¹, Anton Hartmann¹, Claudia Traidl-Hoffmann^{2,3}, Michael Schmid^{1*}

1 Research Unit Microbe-Plant Interactions, *Helmholtz Zentrum München*—German Research Centre for Environmental Health (GmbH), Neuherberg, Germany, **2** Institute of Environmental Medicine, UNIKA-T, *Technische Universität München*, Augsburg, Germany, **3** CK Care, Christine-Kühne-Center for Allergy Research and Education, Davos, Switzerland, **4** Institute of Biochemical Plant Pathology, *Helmholtz Zentrum München*—German Research Centre for Environmental Health (GmbH), Neuherberg, Germany

* michael.schmid@helmholtz-muenchen.de



OPEN ACCESS

Citation: Obersteiner A, Gilles S, Frank U, Beck I, Häring F, Ernst D, et al. (2016) Pollen-Associated Microbiome Correlates with Pollution Parameters and the Allergenicity of Pollen. PLoS ONE 11(2): e0149545. doi:10.1371/journal.pone.0149545

Editor: Ulrich Melcher, Oklahoma State University, UNITED STATES

Received: November 17, 2015

Accepted: February 2, 2016

Published: February 24, 2016

Copyright: © 2016 Obersteiner et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The authors have no support or funding to report.

Competing Interests: The authors have declared that no competing interests exist.

Abstract

Pollen allergies have been rapidly increasing over the last decades. Many allergenic proteins and non-allergenic adjuvant compounds of pollen are involved in the plant defense against environmental or microbial stress. The first aim of this study was to analyze and compare the colonizing microbes on allergenic pollen. The second aim was to investigate detectable correlations between pollen microbiota and parameters of air pollution or pollen allergenicity. To reach these aims, bacterial and fungal DNA was isolated from pollen samples of timothy grass (*Phleum pratense*, $n = 20$) and birch trees (*Betula pendula*, $n = 55$). With this isolated DNA, a terminal restriction fragment length polymorphism analysis was performed. One result was that the microbial diversity on birch tree and timothy grass pollen samples (Shannon/Simpson diversity indices) was partly significantly correlated to allergenicity parameters (Bet v 1/Phl p 5, pollen-associated lipid mediators). Furthermore, the microbial diversity on birch pollen samples was correlated to on-site air pollution (nitrogen dioxide (NO₂), ammonia (NH₃), and ozone (O₃)). What is more, a significant negative correlation was observed between the microbial diversity on birch pollen and the measured NO₂ concentrations on the corresponding trees. Our results showed that the microbial composition of pollen was correlated to environmental exposure parameters alongside with a differential expression of allergen and pollen-associated lipid mediators. This might translate into altered allergenicity of pollen due to environmental and microbial stress.

Introduction

Over the last decades, incidences of allergic diseases have been increasing worldwide thus causing tremendous health problems and imposing considerable economic burden on the society [1]. A prime causative of allergies are airborne pollen grains from anemochorous plants (like

trees, grasses or weeds) causing diseases like polinosis and hay-fever symptoms [2]. Sensitization rates to pollen allergens are estimated to range from 20% to 30% in Germany [3, 4]. The allergenic potential of pollen depends on the expression of immunogenic proteins—the allergens—as well as on non-allergenic molecules exerting pro-inflammatory or immune-modulatory effects [5]. Among these are NADPH oxidases, proteases, adenosine, or pollen-associated lipid mediators (PALMs) [6–8]. Some plant allergens, such as the birch pollen major allergen Bet v 1, belong to the pathogenesis-related (PR) family of proteins involved in host defense systems [9]. Specifically, Bet v 1 is proposed to confer protection from pathogens during pollen germination on the stigma [10]. Likewise, PALMs are assumed to play a role in the plant's stress response to pathogens or heavy metal exposure [11, 12]. Plants constantly monitor the presence of microbes via specialized pattern recognition receptors. Analogous to immune cells in the animal kingdom, plant cells respond to pathogen associated molecular patterns, such as bacterial lipopolysaccharide, peptidoglycans, flagellin, or fungal chitin and β -glucan, by inducing specific host defense programs [13, 14]. Besides closure of stomata, host defense mechanisms can involve a burst of nitric oxide and reactive oxygen species production [15], which are both suggested to trigger PR-protein expression or synthesis of lipid mediators in tobacco cultures [16, 17]. Our previous data imply that those birch trees exposed to elevated ozone levels produce pollen grains with elevated levels of Bet v 1 and an altered PALMs composition, eventually leading to higher pro-inflammatory potential [18]. Furthermore, abiotic stress might alter a plant's susceptibility to microbial pathogens [19]. In turn, pathogen-induced defense mechanisms might change the expression of allergens and adjuvant substances.

Most of the knowledge about plant-microbe interaction results from well characterized model organisms like *Arabidopsis thaliana* and studies of its rhizosphere. In contrast, little is known about microbial colonization of pollen and their influences on the allergen expression in pollen. In this study, we aimed to analyze and compare the microbial community on pollen from birch trees (*Betula pendula*) and timothy grass (*Phleum pratense*) which are both characterized by a high allergenic potential [2]. In addition, we aimed at investigating the correlations between pollen microbiota and parameters of air pollution and pollen allergenicity. For this reason, we collected pollen in urban and rural areas, analyzed their pollen-associated bacterial and fungal communities by performing a tRFLP analysis (terminal restricted fragment length polymorphism), and also did correlation (Spearman) and comparison analyses (PCoA, Cluster). Pollen obtained from birch and timothy grass showed a specific microbial pattern and, for both plant species, we detected an interrelationship between their microbial load and allergen content. Furthermore, we showed that the microbial diversity on birch pollen is correlated to environmental factors typical for urbanized areas. Our findings point out to previously unappreciated environment-plant-microbe interactions as being possible influencing factors of pollen allergenicity.

Material and Methods

Pollen Sampling and Preparation

In the flowering season of 2014, birch (*B. pendula*) catkins were sampled in the city and greater region of Augsburg, Germany. The number of pollen samples is equal to the number of birch trees ($n = 20$ urban, $n = 35$ rural). 100 catkins were sampled at each tree, at approximately 2 m height above the ground, and pollen were sieved from these catkins and pooled for each tree.

Timothy grass (*P. pratense*) spikes were collected from meadows in urban and suburban Munich as well as from rural meadows in Bavaria during the flowering season of 2014. For grass pollen samples, the number of pollen samples was equal to the number of meadows ($n = 20$). Pollen grains were extracted from the aforementioned birch catkins and grass spikes

as it was described in Beck *et al.* [18]. Pollen grains were stored at -80°C . Aqueous pollen extracts of these samples were prepared as it was described in Gilles *et al.* [20].

On-Site Measurements of Air Pollution Parameters

As to the air pollutant measurements, NO_2 , O_3 , NH_3 were all measured on-site at each birch tree by means of passive samplers for the duration of eight weeks, beginning before flowering in march and spanning the whole pollen season as described in Beck *et al.* [18]. So there is one value for each measurement site/birch pollen sample ($n = 55$).

No on-site air pollutant measurements were performed at the grass pollen sampling-sites, only an urban index was calculated.

Urbanization Index

The Urbanization Index (UI) was calculated on the basis of CORINE Land Cover data as described previously in Beck *et al.* [18]. Briefly, CORINE classifies land use in the area of the European Union into 44 categories. Areas with continuous urban fabric, discontinuous urban fabric, industrial or commercial units, road and rail networks and associated land, port areas, airports, mineral extraction sites, dump sites and construction sites are all classified as urban. The Urbanization Index shows the share of urban land use in the surrounding of 1km around the site. If $\text{UI} = 1$, the land use in the surrounding of the site is strictly urban, if $\text{UI} = 0$, the surrounding is completely of non-urban land use.

Determination of Allergenicity Parameters

Levels of Bet v 1 and PALMs in aqueous pollen extracts were determined by ELISA as described in Beck *et al.* [18]. Group 5 grass pollen allergen (Phl p 5) was determined in aqueous grass pollen extracts by ELISA as described in Buters *et al.* [21].

Determination of NADPH Oxidase Activity

NADPH oxidase activity measurement was adapted from Wang *et al.* [22]. In brief, pollen grains (1mg/assay) were hydrated in PBS and mixed with or without 2 mM nitroblue tetrazolium (NBT) (Sigma-Aldrich) and 1 mM nicotinamide adenine dinucleotide phosphate (reduced) (NADPH) (Roth) solved in pure water. The reaction (240 μl /assay) was started by adding the NBT. The pollen was afterwards incubated for one hour at 37°C under rotation. The obtained suspension was centrifuged and the NBT residual was removed by washing twice with 1 ml PBS. The produced formazan sediment was then resolved in 300 μl methanol under rotation for 30 minutes at room temperature and centrifuged again. Supernatants (200 μl) were measured at 530 nm in a Tecan infinite M1000 PRO.

tRFLP-Analysis of Bacterial Communities

Bacterial and fungal DNA from pollen was extracted with a Fast DNA Spin kit for soil (MP Biomedicals), according to the manufacturer's instructions including an additional enzymatic lysis (10 μg of lysozyme, 30 min, 37°C). For the following steps each sample of extracted microbial DNA was divided in two parts, one for the determination of fungal pattern and one for the determination of bacterial pattern. In case of bacterial pattern firstly the 16S rRNA coding genes were amplified with a TopTaq DNA polymerase kit (Qiagen), a FAM-labeled forward primer 27f [23] and the reverse primer 907r [24]. Amplification was processed as follows: initial denaturation (94°C , 5 min), 30 cycles of denaturation (94°C , 45 sec), annealing (59°C , 45 sec), and elongation (72°C , 45 sec). A final extension at 72°C for five minutes completed the

reaction. For the amplification of fungal ITS1-ITS2-region FAM-labeled primer ITS1F with the reverse primer ITS4 [25, 26] were used. After horizontal agarose gel-electrophoresis (120 V, 45 min), the amplicons were purified with a NucleoSpin Gel and PCR Clean up kit (Macherey & Nagel) according to the manufacturer's instruction. For a rapid DNA digest, 16S rRNA gene products were restricted using the enzyme MspI (HpaII; 5'-C[^]CGG-3'; [27]) for 2h incubation at 37°C in recommended buffer. Amplified ITS-products, however, were restricted using the enzyme TaiI (MaeII; 5'-ACGT-3') for 2h at 65°C in recommended buffer (Thermo Fisher Scientific). In pre-experiments regarding the efficiency and obtained fungal diversity the restriction enzymes TaiI performed best (Weikl F. 2015, pers. communication). After transferring the obtained fragments to HiDi Formamid (Applied Biosystems) including MapMarker ROX1000 (1:400; Bioventures), the denatured fragments were separated by an electrophoresis based on a capillary system on an automated DNA sequencer (ABI 3730, Applied Biosystems, Applied Biosystems GmbH, Darmstadt, Germany). The validation of data quality was done by using the online software package Peak Scanner Software v1.0 (Life technologies). True peaks (size < 50bp, height < 50 fluorescent units) were imported into t-REX (<http://trex.biohpc.org/>) for improving the data's quality. Settings were adjusted according to Culman *et al.* [27] and Chowdhury *et al.* [28]. The data matrix of a bacterial pattern was first exported and then statistically analyzed in PAST ver. 2.17c [29]. In the following statistics of microbial pattern analysis the fragments (tRFs) were assumed to be derived from the same group of organisms in each sample.

Exclusion of plastidic DNA

During amplification of bacterial 16S-rRNA coding genes, sequences of plastidic DNA from plant cells were co-amplified. For exclusion, the length of plastid-fragments was determined by tRFLP using the fluorescently labeled forward primer 27f [23], paired with the plastid-specific reverse primer CYA781r [30] for amplification. The obtained fragments were ligated and cloned by using the StrataClone cloning kit (Agilent technologies). For the colony PCR the forward primer T3 was used in combination with the reverse primer T7, both recommended in the manual of the StrataClone Cloning kit. Cloned fragments were sequenced at SequiServe GmbH (Vaterstetten, Munich, Germany). The hereby obtained sequences were aligned with the aligner tool implemented into the ARB software package [31]. For the case that a cloning procedure wants to be avoided, another well working opportunity for the exclusion of plastid DNA would be a 16S rRNA gene amplification with the primer pair 799f and 1492r. According to Chelius & Triplett 2001 [32] the 799f primer shows a different binding position in bacterial and plastid DNA. Hence, bands showing different length can be separated electrophoretically.

Statistics

To compare the microbial composition of two groups the R-value and p-value of a one-way analysis of similarity (ANOSIM) was calculated as described in Clarke [33]. The ANOSIM R-value describes similarity-ranges from completely overlapping features (R = 0) to distinctly separated features (R = 1) of the compared communities [28]. For descriptive statistics, a principal coordinate analysis (PCoA, Bray-Curtis-Index) and a cluster-dendrogram (Bray-Curtis-Index) were performed. Additionally two indices referring to the diversity of the communities were calculated in Rstudio (Version 0.98.953, 2009–2013 RStudio, Inc.; 2014) [34]. To assign the alpha-diversity of the microbial composition the Simpson-Index 1-D (weighing high abundant fragments) and Shannon-Index H (weighing less abundant fragments) was assessed [35]. For comparison of numeric data, the significance level p was calculated by using the Man U Whitney-test in case of non-Gaussian distributed data. For Gaussian distributed data, the t-test for equal / unequal data was used to obtain the significance level. For correlations of measured

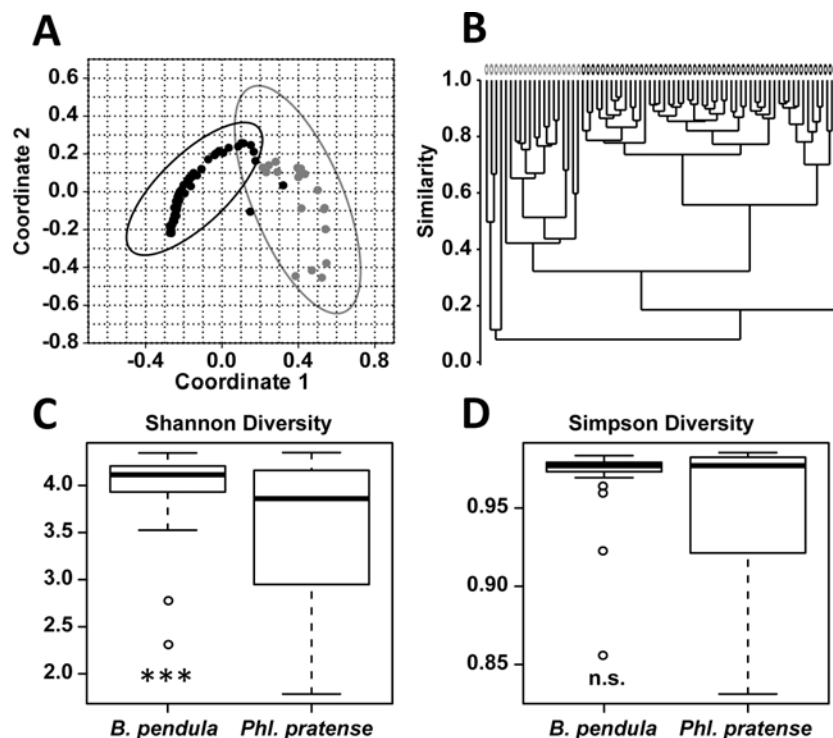


Fig 1. Comparison of bacterial pattern on pollen from birch and timothy grass. **A)** PCoA (Bray-Curtis-Index) of bacterial pattern on birch pollen (*Betula pendula*; $n = 55$; black) and timothy grass pollen (*Phleum pratense*; $n = 20$; grey) show hardly overlapping 95%-confidence intervals. The significant differences were confirmed by ANOSIM ($R = 0.81$, $p = 0.0001$). **B)** In the cluster dendrogram (Bray-Curtis) pollen from both species are forming separated groups also attesting the significant difference. **C,D)** Boxplots of indices weighing species due to their extent of abundance in bacterial community on birch- (*Betula pendula*; $n = 55$) and timothy grass-pollen (*Phleum pratense*, $n = 20$). In comparison the two groups differs significantly due to low abundant species (Shannon-index, $p = 0.009$), but not significantly due to high abundant species of bacterial composition (Simpson-index, $p = 0.78$).

doi:10.1371/journal.pone.0149545.g001

pollution and allergen parameter to microbial diversity-indices, Spearman's r_s and permutation p-value was calculated for non-Gaussian distributed samples [35]. For descriptive and univariate statistics of mean / median comparison as well as for correlations analyses, the statistical program PAST ver. 2.17c was used [29].

Results

Species Dependent Microbial Patterns on Pollen

The bacterial pattern distribution of pollen from birch trees ($n_{\text{birch}} = 55$) and timothy grass ($n_{\text{grass}} = 20$) was illustrated in a PCoA (Principle Coordinate Analysis). The therein created 95%-confidence intervals were not overlapping. In an appending cluster dendrogram two separate clusters were formed ($R = 0.81$, $p = 0.001$). The diversity of the bacterial composition on birch tree and timothy grass pollen differed significantly due to low abundant bacterial species (Shannon index, $p = 0.009$) but not due to the dominant species (Simpson index, $p = 0.78$) (Fig 1).

When comparing the fungal community composition on pollen obtained from birch trees ($n_{\text{birch}} = 31$) and timothy grass ($n_{\text{grass}} = 16$) it differed significantly ($R = 0.51$; $p = 0.0001$) between the two of them. Similar to the bacterial community the 95%-confidence interval of the PCoA did not completely overlapped in this case, too. In the cluster dendrogram the separation of the two groups was less distinct than for bacterial patterns, but still was statistically significant. The

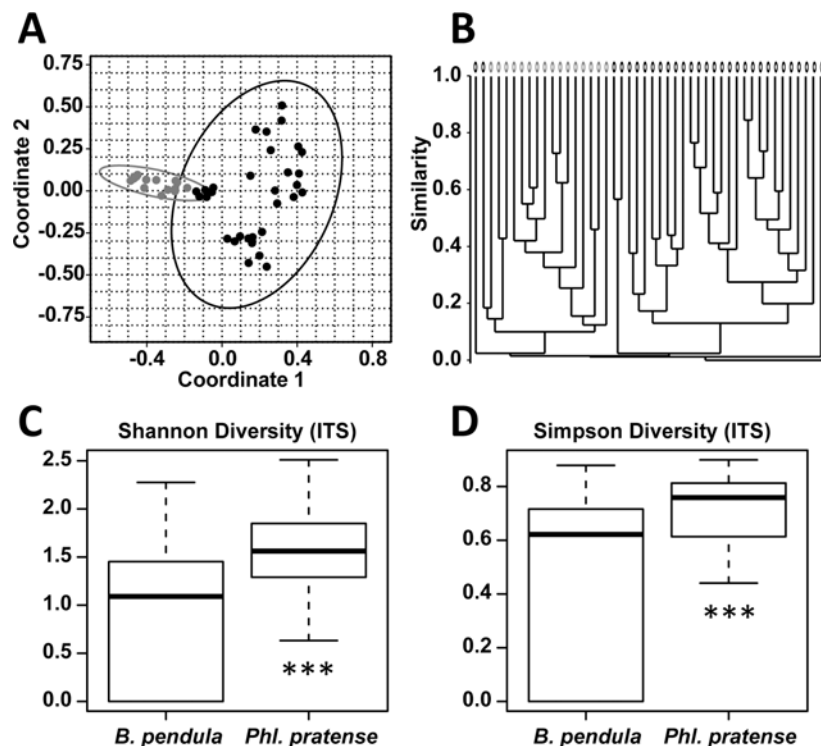


Fig 2. Comparison of fungal pattern on pollen from birch and timothy grass. **A)** PCoA (Bray-Curtis-Index) of fungal pattern on birch pollen (*Betula pendula*; $n = 31$; black) and timothy grass pollen (*Phleum pratense*; $n = 16$; grey) show hardly overlapping 95%-confidence. The significant differences were confirmed by ANOSIM ($R = 0.51$, $p = 0.0001$). **B)** The cluster dendrogram (Bray-Curtis-Index) demonstrates the separation of the two pollen species due to fungal pattern. **C,D)** Boxplots of indices weighing species due to their extent of abundance in fungal community on birch- and timothy grass-pollen. In comparison the two groups differed significantly due to low abundant species (Shannon-index, $p = 0.006$), and also due to high abundant species of fungal composition (Simpson-index, $p = 0.005$).

doi:10.1371/journal.pone.0149545.g002

diversity indices calculated, weighing either high abundant (Simpson-Index) or low abundant fungal species (Shannon-Index), showed that birch pollen had a significantly higher fungal diversity than timothy grass pollen ($p_{\text{Shannon}} = 0.006$, $p_{\text{Simpson}} = 0.004$) (Fig 2).

Urban Sample Sites Show Different Bacterial Composition on Birch Pollen, but Not on Timothy Grass Pollen

The bacterial composition on the gathered pollen from urban sample sites partly show significant differences compared to the pollen collected in rural sample sites. Significant differences were detected for the bacterial pattern on birch pollen ($n_{\text{urban}} = 20$, $n_{\text{rural}} = 35$; $R = 0.23$, $p = 0.003$). Rural and urban timothy grass pollen samples ($n_{\text{urban}} = 13$, $n_{\text{rural}} = 4$) did not show statistically significant differences in bacterial composition ($R = 0.02$; $p = 0.36$). However, the fungal composition on birch tree ($n_{\text{urban}} = 9$, $n_{\text{rural}} = 22$) and timothy grass pollen ($n_{\text{urban}} = 14$, $n_{\text{rural}} = 4$) did not differ in urban or rural samples ($R_{\text{birch}} = 0.04$, $p_{\text{birch}} = 0.62$; $R_{\text{grass}} = 0.09$, $p_{\text{grass}} = 0.69$; Fig 3).

Bacterial Diversity on Birch Pollen Was Correlated with NO_2 Levels and Urbanization Index

Pollen was sampled from 55 birch trees and on-site air pollutant measurements were performed by means of passive samplers at each tree. Due to loss of passive samplers at a few sites,

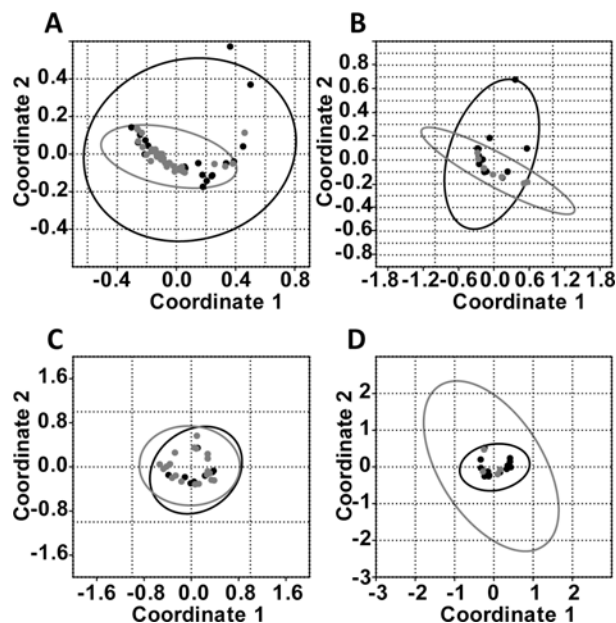


Fig 3. Comparison of bacterial and fungal pattern on pollen collected in rural and urban sample sites. PCoA performed to compare the microbial composition on pollen obtained from rural (grey) and urban areas (black). **A)** Bacterial composition on birch pollen. **B)** Bacterial composition on timothy grass pollen. **C)** Fungal composition on birch pollen. **D)** Fungal composition on timothy grass pollen.

doi:10.1371/journal.pone.0149545.g003

air pollutant measurements could be obtained for the following number of sampling sites: $n(\text{NO}_2) = 51$, $n(\text{O}_3) = 52$, $n(\text{NH}_3) = 49$.

Diversity indices of bacterial community on birch pollen were significantly correlated with pollution parameters of pollen sampling sites (Fig 4). Specifically, a significant negative correlation between the number of species (= total number of fragments; $n(\text{tRFs})$) and ambient levels of NO_2 ($r_s = -0.31$, $p = 0.027$; Fig 4B) was observed. In addition, a significant negative correlation between $n(\text{tRFs})$ and calculated UI ($r_s = -0.36$, $p < 0.001$; Fig 4D), but not to O_3 - ($r_s = 0.21$, $p = 0.13$; Fig 4A) or ammonia NH_3 -levels ($r_s = 0.14$, $p = 0.34$; Fig 4C) was observed. In contrast, the diversity of high abundant species (Simpson-Index) correlated significantly positive with O_3 -concentrations ($r_s = 0.39$, $p = 0.0039$; Fig 4E), but significantly negative to the calculated UI ($r_s = -0.44$, $p < 0.001$; Fig 4H) and NO_2 -level ($r_s = -0.54$, $p < 0.0001$; Fig 4F). There was no correlation of bacterial Simpson-diversity to NH_3 -levels ($r_s = 0.01$, $p = 0.94$; Fig 4G). The diversity of low abundant species (Shannon-Index) did not show significant correlations to any of the measured pollution parameters ($r_s(\text{O}_3) = 0.19$, $p(\text{O}_3) = 0.16$, Fig 3I; $r_s(\text{NO}_2) = 0.14$, $p(\text{NO}_2) = 0.31$, Fig 4J; $r_s(\text{NH}_3) = 0.01$, $p(\text{NH}_3) = 0.89$, Fig 4K; $r_s(\text{UI}) = 0.06$, $p(\text{UI}) = 0.67$, Fig 4L).

Bacterial Diversity on Birch Pollen Correlated with Content of Bet v 1 and PALMs

Diversity indices (Shannon, Simpson, $n(\text{tRFs})$) calculated for the bacterial community on birch pollen were plotted against parameters defining the pollen's allergenic potential (content of Bet v 1, PALMs, NADPH oxidase). Significant correlations were observed (Fig 5), specifically, the total number of different species ($n(\text{tRFs})$) was positively correlated to the concentration of Bet v 1 ($r_s = 0.28$, $p = 0.038$; Fig 5A) but not to the concentration of $\text{PALM}_{\text{LTB4}}$ ($r_s = 0.14$, $p = 0.33$; Fig 5B), $\text{PALM}_{\text{PGE2}}$ ($r_s = 0.17$, $p = 0.22$; Fig 5C) or NADPH oxidase ($r_s = -0.04$,

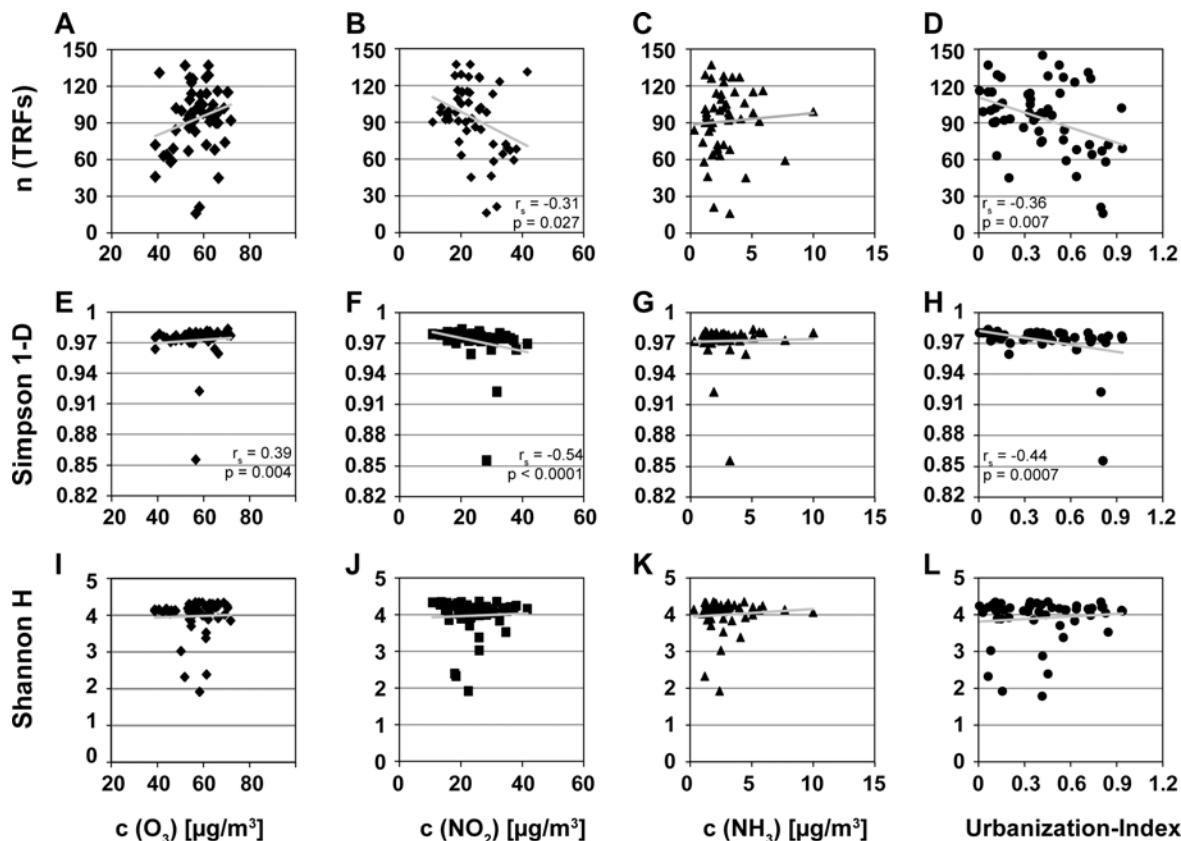


Fig 4. Spearman correlations of bacterial diversity on birch pollen to pollution parameter. Scatterplot including linear trend line, spearman correlation values rho and significance level p of diversity indices (Shannon H/Simpson 1-D) and parameter (number of fragments n(trFs)) of bacterial pattern on birch pollen (*Betula pendula* 2014, n = 55) with pollution measurements. **A-D** Correlation of n(trFs) and pollution parameters ozone O₃ ($r_s = 0.21$, $p = 0.13$), nitrogendioxid NO₂ ($r_s = -0.31$, $p = 0.027$), ammonia NH₃ ($r_s = 0.14$, $p = 0.34$) and the urbanization-index ($r_s = -0.36$, $p < 0.01$). **E-H** Correlation of Simpson diversity Index 1-D and pollution parameters O₃ ($r_s = 0.39$, $p = 0.004$), NO₂ ($r_s = -0.54$, $p < 0.0001$), NH₃ ($r_s = 0.01$, $p = 0.94$) and the urbanization-index ($r_s = -0.44$, $p = 0.0007$). **I-L** Correlation of Shannon diversity Index H and pollution parameters O₃ ($r_s = 0.19$, $p = 0.17$), NO₂ ($r_s = -0.14$, $p = 0.31$), NH₃ ($r_s = 0.01$, $p = 0.89$) and the urbanization-index ($r_s = 0.06$, $p = 0.67$).

doi:10.1371/journal.pone.0149545.g004

$p = 0.79$, Fig 5D). The diversity of high abundant species (Simpson-Index) showed no significant correlation with any of the measured parameters of the respective allergenic potential (r_s (Bet v 1) = 0.25, p (Bet v 1) = 0.068, Fig 5E; r_s (PALM_{LTB4}) = 0.08, p (PALM_{LTB4}) = 0.53, Fig 5F; r_s (PALM_{PGE2}) = 0.12, p (PALM_{PGE2}) = 0.366, Fig 5G; r_s (NADPH oxidase) = 0.05, p (NADPH oxidase) = 0.75, Fig 5H). The diversity of low abundant species (Shannon-Index), however, was significantly negative correlated to the concentration of PALM_{LTB4} ($r_s = -0.36$, $p = 0.006$; Fig 5I) and PALM_{PGE2} ($r_s = -0.04$, $p = 0.001$; Fig 5K), despite a low r_s -value. Shannon diversity did not correlate to the concentration of Bet v 1 ($r_s = 0.14$, $p = 0.317$; Fig 5J) or NADPH oxidase ($r_s = 0.2$, $p = 0.19$; Fig 5L).

No Correlation between Fungal Diversity on Birch Pollen with Pollution or Allergenicity Parameters

In contrast to the bacterial patterns the composition of fungi on birch pollen grains were not significantly correlated to any of the pollution (NO₂, O₃, NH₃) or allergenicity parameters (Bet v 1, PALM_{PGE2}, PALM_{LTB4}, NADPH oxidase) tested (S1 Table).

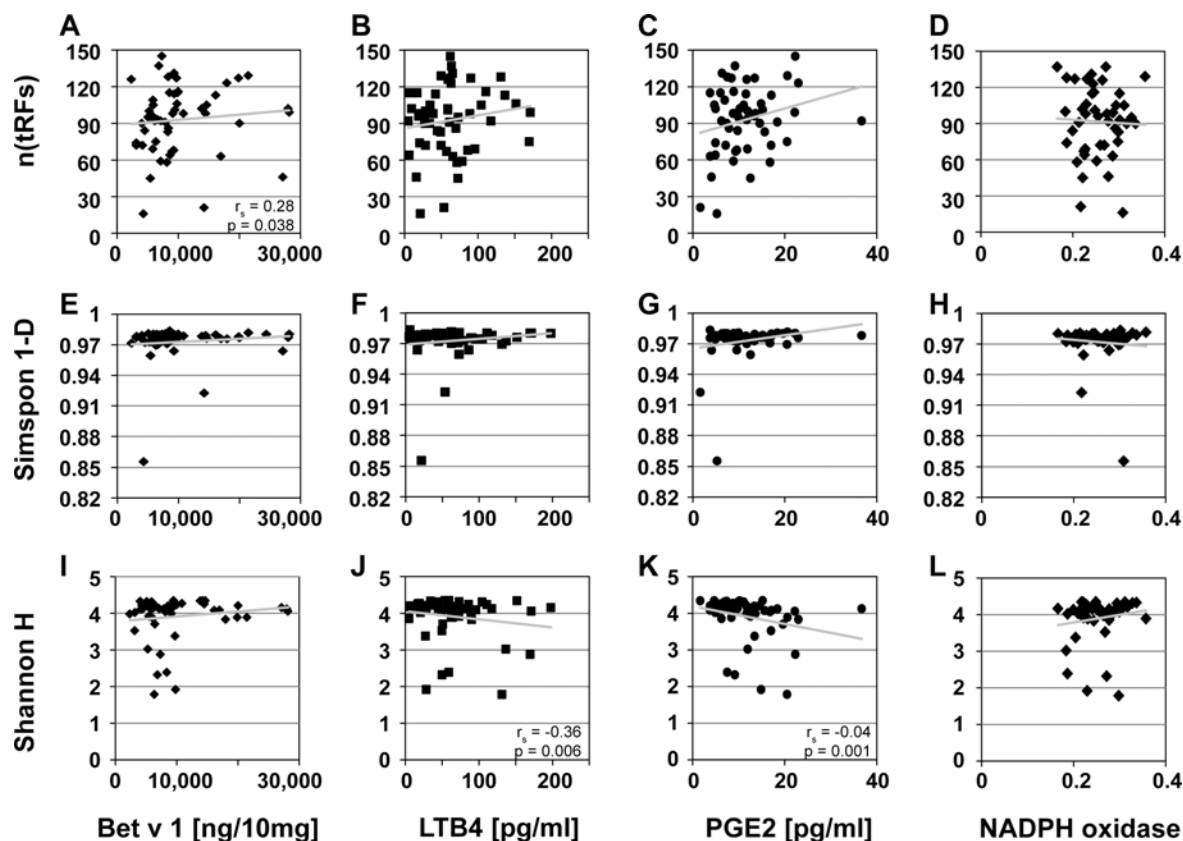


Fig 5. Spearman correlations of bacterial diversity on birch pollen to allergenicity parameter. Scatterplot including linear trend line, spearman correlation values rho and significance level p of diversity indices (Shannon H/Simpson 1-D) and parameter (number of fragments n(tRFs)) of bacterial pattern on birch pollen (*Betula pendula* 2014, n = 55) with allergen concentration in pollen. **A-D)** Correlation of n(tRFs) and allergenicity parameters Bet v 1 ($r_s = 0.28$, $p = 0.038$), $PALM_{LTB4}$ ($r_s = 0.14$, $p = 0.33$), $PALM_{PGE2}$ ($r_s = 0.17$, $p = 0.22$) and NADPH oxidase ($r_s = -0.04$, $p = 0.79$). **E-H)** Correlation of Simpson diversity Index 1-D and allergenicity parameters Bet v 1 ($r_s = 0.25$, $p = 0.068$), $PALM_{LTB4}$ ($r_s = 0.08$, $p = 0.53$), $PALM_{PGE2}$ ($r_s = 0.12$, $p = 0.366$) and NADPH oxidase ($r_s = 0.2$, $p = 0.19$). **I-L)** Correlation of Shannon diversity Index H and allergenicity parameters Bet v 1 ($r_s = 0.14$, $p = 0.317$), $PALM_{LTB4}$ ($r_s = -0.36$, $p = 0.006$), $PALM_{PGE2}$ ($r_s = -0.04$, $p = 0.0015$) and NADPH oxidase ($r_s = 0.2$, $p = 0.19$).

doi:10.1371/journal.pone.0149545.g005

Bacterial, but Not Fungal Diversity on Timothy Grass Pollen Correlated with the Content of $PALM_{PGE2}$ and Urbanization Index

The statistical analysis of bacterial patterns obtained from timothy grass pollen resulted in a significantly negative correlation ($r_s = -0.51$, $p = 0.039$) between the diversity of high abundant species (Simpson-Index) and the $PALM_{PGE2}$ content of pollen. Moreover, there was a significantly negative correlation between highly abundant species (Simpson-Index) ($r_s = -0.57$, $p = 0.04$) and the Urbanization Index (UI) and also between the total number of species (n-tRFs); $r_s = -0.58$, $p = 0.04$) and the Urbanization Index (UI). No further significant interrelationships were observed between bacterial diversity indices and parameters of allergenic potential (S2 Table). Similarly, no significant correlations were observed between fungal diversity on timothy grass pollen and allergenicity-related parameters or the Urbanization Index (S3 Table).

Discussion

Plant-microbe interactions govern diverse physiological processes in plants such as growth regulation, nutrient supply, and host defense [36]. So far, however, only a few previous studies

investigated the presence of microbes on plant pollen [37]. A more recent paper described the identification of gram-positive *Bacillus* species on timothy grass pollen. This paper also demonstrated that bacterial isolates can promote the maturation of human monocyte-derived dendritic cells [38]. To our knowledge, the present study is the first which systematically characterizes communities of bacteria and fungi on allergenic pollen grains and which also correlates the microbial colonization of pollen with parameters of environmental pollution and allergenicity. It has been known that anthropogenic pollutants, such as ozone and ultra-fine particulate matter, exert direct adverse effects on the human health [39, 40]. However, not only humans are exposed to air pollutants, but also the plants producing allergenic pollen. In recent years, it has become increasingly clear [5] that pollen allergenicity is not a constant parameter, but is readily influenced by biogenic and anthropogenic stress factors—among them are drought stress and such pollutions which cause stress to the allergenic plant [41, 42].

Pollen Associated Microbiome Depends on Plant Species

In our present study, we firstly compared the microbial pattern on pollen grains of two different species, birch tree and timothy grass, and were thereby able to detect species-specific patterns due to bacterial and fungal colonization. The composition of microbial patterns as well as the extent of the microbial diversity revealed significant differences between birch tree and timothy grass pollen. Birch pollen showed a higher diversity in dominant bacterial species, but a lower fungal diversity than pollen obtained from timothy grass.

The microbial load of natural samples is influenced by several biotic factors (insect occurrence, contact to other plants) and abiotic factors (temperature, wind, humidity) which both characterize an environment [43, 44]. Birch catkins and timothy grass inflorescence are exposed to different environments caused e.g. by growth height or blooming period. Beside the presence of surface colonizing microbes in our microbial communities, also endophytes must be considered because of the mechanical and enzymatic cell-lysis during DNA-extraction. Amongst other bacteria, a strain of *Enterobacter cloacae* was previously identified in surface-sterilized pollen, roots, and shoots of several Pine species [45]. Translocational processes within the plants, like it is known for nutrients or heavy metals [46], may promote the existence of endophytic microbes in pollen. Other plant-specific parameters influencing the soil microbiome in the rhizosphere (e.g. root exudates altering pH-milieu or antimicrobial compounds [47]) would further impact the composition of endophytic microbes and might then result in diverse microbial pattern in pollen of different plant species. Another important factor influencing pollen-associated microbial colonization is the plant's genotype. In experiments, mutants from *A. thaliana* which contain an altered cuticular wax biosynthesis presented a significantly different microbial composition on leaves compared to the plant's wildtype [48]. Pollen of different plant species also shows a high phenotypical variability, thus influencing the occurrence of microbes, for instance a diverse structure of the exine [49] as well as various chemical and physical signals delivered by the exine in order to avoid inappropriate fertilization [50].

Effect of Urbanization on the Pollen-Microbiome

Bacterial community patterns on timothy grass pollen did not differ between urban and rural sampling sites, whereas differences in bacterial patterns were observed between rural and urban birch pollen. The bacterial diversity (n(tRFs), Simpson-Index) correlated significantly negative with ambient NO₂ levels and the calculated Urbanization Index. Ozone levels, in contrast, were positively correlated to the bacterial diversity (Simpson-Index), which was also visible by the trend of enhanced total species' diversity (ntRFs).

A study analyzing the impact of O_3 to soil microbiome in the rhizosphere showed that O_3 influences bacterial communities not directly but indirectly by stressing the plant [51, 52]. Normal atmospheric concentrations of O_3 already cause damage in crop plants [53]. NO_2 , in contrast, at normal atmospheric concentrations supports the plant by promoting nutrient uptake, plant growth, and metabolism [54]. Only artificial high amounts of NO_2 would lead to negative effects on the plant [55]. Our data suggest that alterations of ambient air pollutants, whether favorable or unfavorable for the general growth condition, are accompanied by changes in bacterial composition colonizing the plant.

In general, higher NO_2 and lower O_3 concentrations are typically associated with high traffic urban areas. This gradient from rural to urban locations is distinctly shown by the significant positive correlation of NO_2 and the significant negative correlation of O_3 values to an increasing UI-value (S1 Fig). In detail, urban environments expose plants to a wide range of abiotic stressors, among them polluted soil, heat, drought stress and nutrient shortage [56]. All these factors, in case they occur in high intensity, can stress plants [19]. We did not analyze the influence of one of these additional stress factors to the plants. Therefore, we cannot suggest a direct relation of NO_2 to the stress level in plants, because it could also be possible that another stress factor, which correlates positively with NO_2 is the main stressor for the plants.

For measuring the stress level of the plants, we determined the activity of pollen NADPH oxidases. Besides the impact on allergic inflammation in the airways of animals and humans NADPH oxidases are involved in several processes of the plant system such as the response to biotic and/or abiotic stimuli (summarized in [57]). Our data show no correlation of NADPH oxidase activity to any of the measured pollution parameters. Neither was there any significant correlation of NADPH oxidase activity to microbial diversity. This suggests that NADPH oxidase activity in birch tree and timothy grass pollen is irrelevant for responses to abiotic stresses measured in this study.

A study analyzing the communities of dust-associated microbes found a declined microbial diversity in urban areas compared to rural areas [58]. We can confirm these investigations with regard to our results which show a declined bacterial diversity on birch pollen in urbanized areas. It is difficult to say what urban factors exactly might affect bacterial communities to the point of homogenization. There are studies showing high effects [59] and other studies showing little effect of urbanization to the atmospheric bacterial diversity [60]. Lower bacterial diversity might result from a higher homogenization of environmental conditions, land-use types or homogenized plant community in cities [58, 61]. Furthermore, there might be a link to the hypothesis that people living in urban areas often suffer from allergic diseases caused by an exposition to lower level of bacterial diversity [58].

Correlation of Pollen Microbiome and Allergen/PALM Contents

In further correlation analyses, we investigated interrelationships between the microbial diversity and the pollen's content of allergens and PALMs in birch tree and timothy grass pollen. Higher bacterial diversity (n(tRFs)) on birch pollen was found to be associated with higher concentrations of Bet v 1. Higher contents of PALMs were associated with a lower Shannon diversity on birch tree and timothy grass pollen.

Plants are able to recognize bacterial colonization via pattern recognition receptors (PRRs) on the plant cell surface [13, 14]. Microbe-recognition by PRRs induces a wide spectrum of defense responses in the plant [62]. The protein Bet v 1 belongs to the plant pathogenesis-related protein family 10 (PR-10). This protein is suggested to play a role in the response against pathogens or abiotic stress. In birch cell culture approaches it was detected that the gene expression encoding Bet v 1 isoforms is transcriptionally activated during pollen

maturation and rapidly induced in the presence of microbes. Other stress factors like heat or heavy metals did not induce such an effect. An antibiotic function of enhanced Bet v 1 concentration seem to be likely, but tests with pollen washes including high amounts of Bet v 1 protein do not show any antibiotic effects [9]. In our study, we analyzed the interrelationship between microbes and Bet v 1 in natural habitats. In combination with the information we received from the cell culture approach explained above [9], the hypothesis of microbes influencing the expression of the allergen Bet v 1 might be supported. Future studies should address the effect of microbial species on the expression of Bet v 1 in pollen in more detail.

Finally, the diversity of low abundant bacteria on birch pollen was negatively correlated to the PALM content of pollen. Despite a low r_s -value in case of PALM_{PGE2} the correlation was significant because of the high number of analyzed samples. The physiological roles of most PALMs are unknown to date. Some PALMs, e.g. the B₁-Phytosteranes are involved in the plant defense response against heavy metal and oxidants [11, 12]. High diversity of low abundance microbes, as measured by a high Shannon index, could reflect the presence of plant pathogens. In this case, PALMs production could be elevated in the course of pathogen-induced stress-responses. On the other hand, PALMs were measured by means of cross-reactivity to PGE₂- and LTB₄- antibodies in commercial ELISAs. It is therefore also possible that some lipids measured are not of pollen but of microbial origin. Detailed studies on pollen, from metabolome to function, will be required for dissecting the contribution of different PALMs *in planta* as well as their impact on the human immune system.

Conclusion

In this study we characterized systematically bacterial and fungal communities colonizing allergenic pollen from birch tree and timothy grass. Summing up the gained results we showed that in pollen of different plant species significantly different microbial communities are represented. Moreover we found that the bacterial diversity on birch pollen decreases in more urbanized locations. Furthermore we were able to show a relationship between high bacterial diversity on pollen and the amounts of allergens and PALMs produced by pollen.

Based on our results and in accordance with other studies we suggest that pollution / urbanization influences the microbial load associated with pollen, which then probably impacts the expression of allergenic proteins and immune-modulatory and stimulatory PALMs in the plant pollen. Up to now, the bacterial colonization of pollen was treated as a factor stressing the plant. We point out that the well-known and important beneficial and symbiotic effects of bacteria must not be neglected, too. With regard to the increasing number of incidences of allergic diseases, it is necessary to reveal to what extent anthropogenic factors influence the allergenic potential of pollen. Further studies should shed light on the molecular interplay on and in pollen grains. These studies should also clear whether there are certain bacterial isolates that might harbor key-functions in terms of modulating the expression of immune-related compounds.

Supporting Information

S1 Fig. Correlation of pollution parameters NO₂ and O₃ to the Urbanization Index (UI).

Ambient NO₂ concentration is significantly positive correlated to the Urbanization Index (UI; $\rho = 0.68$, $p < 0.0001$), whereas ambient O₃ concentration is significantly negative correlated to the Urbanization Index ($\rho = -0.46$, $p < 0.001$). (TIF)

S1 Table. Correlation of fungal diversity on birch pollen to the content of PALMs and allergens, to the activity of NADPH oxidase as well as to measured pollution parameters.

Spearman-Correlation of fungal diversity-indices (Simpson, Shannon) and the absolute number of different fragments (n(tRFs)) analyzed from birch pollen (*Betula pendula*, 2014) to air pollution concentration (NO₂, O₃, NH₃; n = 31), the produced amount of allergens and PALMs (bet v 1, PALM_{PGE2}, PALM_{LTB4}, n = 31) and the activity of NADPH oxidase (n = 18). p = significance level.

(PDF)

S2 Table. Correlation of bacterial diversity on timothy grass pollen to the content of PALMs and allergens and to the Urbanization Index (UI). Spearman-Correlation of bacterial diversity-indices (Simpson, Shannon) and the absolute number of different fragments (n(tRFs)) analyzed from timothy grass pollen (*Phleum pratense*, n = 20; 2014) to the produced amount of allergens (Phl p 5), the amount of PALMs (PALM_{PGE2}, PALM_{LTB4}) and the Urbanization Index (UI). p = significance level.

(PDF)

S3 Table. Correlation of fungal diversity on timothy grass pollen to the content of PALMs and allergens and to the Urbanization Index (UI). Spearman-Correlation of fungal diversity-indices (Simpson, Shannon) and the absolute number of different fragments (n(tRFs)) analyzed from timothy grass pollen (*Phleum pratense*, 2014) to the produced amount of allergens (Phl p 5; n = 20) and PALMs (PALM_{PGE2}, PALM_{LTB4}, n = 20) and also to the Urbanization Index (UI; n = 18). p = significance level.

(PDF)

Acknowledgments

The authors would like to thank Angelo Weiss (Helmholtz center Munich, research group “Plant Microbe Interaction”) for his excellent technical assistance and Fabian Weigl (Helmholtz Center Munich, Institute of Biochemical Plant Pathology) for giving us substantial methodological advices.

Author Contributions

Conceived and designed the experiments: AO SG UF CTH DE AH MR MS. Performed the experiments: AO SG UF. Analyzed the data: AO SG UF. Contributed reagents/materials/analysis tools: AO SG UF. Wrote the paper: AO SG UF MR MS. Supervision: MR MS. Calculation of Urbanization Index: FH. Determination of pollen collection day: IB.

References

1. Ring J, Akdis C, Lauener R, Schappi G, Traidl-Hoffmann C, Akdis M, et al. Global allergy forum and second Davos Declaration 2013 allergy: barriers to cure—challenges and actions to be taken. *Allergy*. 2014; 69(8):978–82. PMID: [25041525](#). doi: [10.1111/all.12406](#)
2. D'Amato G, Cecchi L, Bonini S, Nunes C, Annesi-Maesano I, Behrendt H, et al. Allergenic pollen and pollen allergy in Europe. *Allergy*. 2007; 62(9):976–90. PMID: [17521313](#).
3. Boehme MW, Kompauer I, Weidner U, Piechotowski I, Gabrio T, Behrendt H. Respiratory symptoms and sensitization to airborne pollen of ragweed and mugwort of adults in Southwest Germany. *Deutsche Medizinische Wochenschrift* (1946). 2013; 138(33):1651–8. doi: [10.1055/s-0033-1343330](#) PMID: [23913351](#)
4. Nowak D, Heinrich J, Jorres R, Wassmer G, Berger J, Beck E, et al. Prevalence of respiratory symptoms, bronchial hyperresponsiveness and atopy among adults: west and east Germany. *Eur Respir J*. 1996; 9(12):2541–52. PMID: [8980967](#).
5. Traidl-Hoffmann C, Jakob T, Behrendt H. Determinants of allergenicity. *J Allergy Clin Immunol*. 2009; 123(3):558–66. PMID: [19152966](#). doi: [10.1016/j.jaci.2008.12.003](#)

6. Oeder S, Alessandrini F, Wirz OF, Braun A, Wimmer M, Frank U, et al. Pollen-derived nonallergenic substances enhance Th2-induced IgE production in B cells. *Allergy*. 2015. PMID: [26214762](#).
7. Wimmer M, Alessandrini F, Gilles S, Frank U, Oeder S, Hauser M, et al. Pollen-derived adenosine is a necessary cofactor for ragweed allergy. *Allergy*. 2015. PMID: [25939785](#).
8. Gilles S, Beck I, Lange S, Ring J, Behrendt H, Traidl-Hoffmann C. Non-allergenic factors from pollen modulate T helper cell instructing notch ligands on dendritic cells. *World Allergy Organ J*. 2015; 8(1):2. PMID: [25610519](#). Pubmed Central PMCID: 4300172. doi: [10.1186/s40413-014-0054-8](#)
9. Swoboda I, Scheiner O, Kraft D, Breitenbach M, Heberle-Bors E, Vicente O. A birch gene family encoding pollen allergens and pathogenesis-related proteins. *Biochim Biophys Acta*. 1994; 1219:457–64. PMID: [7918643](#)
10. Swoboda I, Dang TCH, Heberle-Bors E, Vicente O. Expression of Bet v 1, the major birch pollen allergen, during anther development. *Protoplasma*. 1995; 187:103–10.
11. Loeffler C, Berger S, Guy A, Durand T, Bringmann G, Dreyer M, et al. B1-phytoprostanes trigger plant defense and detoxification responses. *J Plant physiol*. 2005; 137(1):328–40. PMID: [15618427](#). Pubmed Central PMCID: 548863.
12. Thoma I, Kruschke M, Loeffler C, Mueller MJ. The isoprostanoic pathway in plants. *Chem Phys Lipids*. 2004; 128(1–2):135–48. PMID: [15037159](#).
13. Newman MA, Sundelin T, Nielsen JT, Erbs G. MAMP (microbe-associated molecular pattern) triggered immunity in plants. *Front Plant Sci*. 2013; 4(139):1–14. PMID: [23720666](#). Pubmed Central PMCID: 3655273.
14. Trda L, Boutrot F, Claverie J, Brule D, Dorey S, Poinssot B. Perception of pathogenic or beneficial bacteria and their evasion of host immunity: pattern recognition receptors in the frontline. *Front Plant Sci*. 2015; 6(219). PMID: [25904927](#). Pubmed Central PMCID: 4389352.
15. Khodai-Kalaki M, Andrade A, Fathy Mohamed Y, Valvano MA. *Burkholderia cenocepacia* lipopolysaccharide modification and flagellin glycosylation affect virulence but not innate immune recognition in plants. *MBio*. 2015; 6(3):e00679–15. PMID: [26045541](#). Pubmed Central PMCID: 4462625. doi: [10.1128/mBio.00679-15](#)
16. Thoma I, Loeffler C, Sinha AK, Gupta M, Kruschke M, Steffan B, et al. Cyclopentenone isoprostanes induced by reactive oxygen species trigger defense gene activation and phytoalexin accumulation in plants. *The Plant Journal*. 2003; 34:363–75. PMID: [12713542](#)
17. Jacquard C, Mazeyrat-Gourbeyre F, Devaux P, Boutilier K, Baillieu F, Clement C. Microspore embryogenesis in barley: anther pre-treatment stimulates plant defence gene expression. *Planta*. 2009; 229(2):393–402. PMID: [18974997](#). doi: [10.1007/s00425-008-0838-6](#)
18. Beck I, Jochner S, Gilles S, McIntyre M, Buters JT, Schmidt-Weber C, et al. High environmental ozone levels lead to enhanced allergenicity of birch pollen. *PLoS One*. 2013; 8(11). PMID: [24278250](#). Pubmed Central PMCID: 3835901.
19. Bostock RM, Pye MF, Roubtsova TV. Predisposition in plant disease: exploiting the nexus in abiotic and biotic stress perception and response. *Annu Rev Phytopathol*. 2014; 52:517–49. PMID: [25001451](#). doi: [10.1146/annurev-phyto-081211-172902](#)
20. Gilles S, Fekete A, Zhang X, Beck I, Blume C, Ring J, et al. Pollen metabolome analysis reveals adenosine as a major regulator of dendritic cell-primed T(H) cell responses. *J Allergy Clin Immunol*. 2011; 127(2):454–61. PMID: [21281872](#). doi: [10.1016/j.jaci.2010.12.1082](#)
21. Buters J, Prank M, Sofiev M, Pusch G, Albertini R, Annesi-Maesano I, et al. Variation of the group 5 grass pollen allergen content of airborne pollen in relation to geographic location and time in season. *J Allergy Clin Immunol*. 2015; 136(1):87–95. PMID: [25956508](#). doi: [10.1016/j.jaci.2015.01.049](#)
22. Wang X-L, Takai T, Kamijo S, Gunawan H, Ogawa H, Okumura K. NADPH oxidase activity in allergenic pollen grains of different plant species. *Biochem Biophys Res Commun*. 2009; 387(3):430–4. doi: [10.1016/j.bbrc.2009.07.020](#) PMID: [19595671](#)
23. Edwards U, Rogall T, Blocker H, Emde M, Bottger EC. Isolation and direct complete nucleotide determination of entire genes. Characterization of a gene coding for 16S ribosomal RNA. *Nucleic Acids Res*. 1989; 17(19):7843–53. PMID: [2798131](#). Pubmed Central PMCID: 334891.
24. Lane DJ, Pace B, Olsen GJ, Stahl DA, Sogin ML, Pace NR. Rapid determination of 16S ribosomal RNA sequences for phylogenetic analyses. *PNAS*. 1985; 82:6955–9. PMID: [2413450](#)
25. Lord NS, Kaplan CW, Shank P, Kitts CL, Elrod SL. Assessment of fungal diversity. *FEMS Microbiol Ecol*. 2002; 42:327–37. doi: [10.1111/j.1574-6941.2002.tb01022.x](#) PMID: [19709292](#)
26. Anderson IC, Campbell CD, Prosser JI. Potential bias of fungal 18S rDNA and internal transcribed spacer polymerase chain reaction primers for estimating fungal biodiversity in soil. *Environ Microbiol*. 2003b; 5(1):36–47.

27. Chowdhury SP, Dietel K, Rändler M, Schmid M, Junge H, Borris R, et al. Effects of *Bacillus amyloliquefaciens* FZB42 on lettuce growth and health under pathogen pressure and its impact on the rhizosphere bacterial community. PLoS ONE. 2013; 8(7):e68818. doi: [10.1371/journal.pone.0068818](https://doi.org/10.1371/journal.pone.0068818) PMID: [23935892](https://pubmed.ncbi.nlm.nih.gov/23935892/)
28. Culman SW, Gauch HG, Blackwood CB, Thies JE. Analysis of tRFLP data using analysis of variance and ordination methods: A comparative study. J Microbiol Methods. 2008; 75:55–63. doi: [10.1016/j.mimet.2008.04.011](https://doi.org/10.1016/j.mimet.2008.04.011) PMID: [18584903](https://pubmed.ncbi.nlm.nih.gov/18584903/)
29. Hammer O, Harper DAT, Ryan PD. PAST: Paleontological statistics software package for education and data analysis. Palaeontol Electronica. 2001; 4(1).
30. Nubel U, Garcia-Pichel F, Muyzer G. PCR primers to amplify 16S rRNA genes from cyanobacteria. Appl Environ Microbiol. 1997; 63(8):3327–32. PMID: [9251225](https://pubmed.ncbi.nlm.nih.gov/9251225/). Pubmed Central PMCID: 168636.
31. Strunk O, Ludwig W. ARB software program package 1997. Available: <http://www.biol.chemie.tu-muenchen.de/pub/ARB/>.
32. Chelius MK, Triplett EW. The Diversity of Archaea and Bacteria in Association with the Roots of Zea mays L. Microbial ecology. 2001; 41(3):252–63. PMID: [11391463](https://pubmed.ncbi.nlm.nih.gov/11391463/).
33. Clarke KR. Non-parametric multivariate analyses of changes in community structure. Austral Ecol. 1993; 18:117–43.
34. Team RDC. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing,; 2014.
35. Köhler W, Schachtel G, Voleske P. Biostatistik. Einführung für Biologen und Agrarwissenschaftler. Heidelberg: Springer-Verlag Berlin 2007.
36. Compant S, Clément C, Sessitsch A. Plant growth-promoting bacteria in the rhizo- and endosphere of plants: Their role, colonization, mechanisms involved and prospects for utilization. Soil Biol Biochem. 2010; 42(5):669–78.
37. Colldahl H, Carlsson G. Allergens in pollen. Acta Allergologica. 1968; XXIII:387–95.
38. Heydenreich B, Bellinghausen I, König B, Becker WM, Grabbe S, Petersen A, et al. Gram-positive bacteria on grass pollen exhibit adjuvant activity inducing inflammatory T cell responses. Clin Exp Allergy. 2012; 42(1):76–84. PMID: [22092824](https://pubmed.ncbi.nlm.nih.gov/22092824/). doi: [10.1111/j.1365-2222.2011.03888.x](https://doi.org/10.1111/j.1365-2222.2011.03888.x)
39. Heinrich J, Wichmann H-E. Traffic related pollutants in Europe and their effect on allergic disease. Curr Opin Allergy Clin Immunol. 2004; 4(5):341–8. PMID: [15349031](https://pubmed.ncbi.nlm.nih.gov/15349031/)
40. Diaz-Sanchez D, Riedl M. Diesel effects on human health: a question of stress? Am J Physiol Lung Cell Mol Physiol. 2005; 289(5):L722–L3. PMID: [16214822](https://pubmed.ncbi.nlm.nih.gov/16214822/).
41. El Kelish A, Zhao F, Heller W, Durner J, Winkler JB, Behrendt H, et al. Ragweed (*Ambrosia artemisiifolia*) pollen allergenicity: SuperSAGE transcriptomic analysis upon elevated CO2 and drought stress. BMC Plant Biol. 2014; 14(176):1–16. PMID: [24972689](https://pubmed.ncbi.nlm.nih.gov/24972689/). Pubmed Central PMCID: 4084800.
42. Zhao F, Elkelish A, Durner J, Lindermayr C, Winkler JB, Ruëff F, et al. Common ragweed (*Ambrosia artemisiifolia* L.): Allergenicity and molecular characterisation of pollen after plant exposure to elevated NO2. Plant Cell Environ. 2015.
43. Rastogi G, Sbodio A, Tech JJ, Suslow TV, Coaker GL, Leveau JH. Leaf microbiota in an agroecosystem: spatiotemporal variation in bacterial community composition on field-grown lettuce. ISME J. 2012; 6(10):1812–22. PMID: [22534606](https://pubmed.ncbi.nlm.nih.gov/22534606/). Pubmed Central PMCID: 3446804. doi: [10.1038/ismej.2012.32](https://doi.org/10.1038/ismej.2012.32)
44. Finkel OM, Burch AY, Elad T, Huse SM, Lindow SE, Post AF, et al. Distance-decay relationships partially determine diversity patterns of phyllosphere bacteria on *Tamarix* trees across the Sonoran Desert. Appl Environ Microbiol. 2012; 78(17):6187–93. PMID: [22752165](https://pubmed.ncbi.nlm.nih.gov/22752165/). Pubmed Central PMCID: 3416633. doi: [10.1128/AEM.00888-12](https://doi.org/10.1128/AEM.00888-12)
45. Madmony A, Chernin L, Pleban S, Peleg E, Riov J. *Enterobacter cloacae*, an obligatory endophyte of pollen grains of mediterranean pines. Folia Microbiol. 2005; 50(3):209–16.
46. Mattina MI, Lannucci-Berger W, Musante C, White JC. Concurrent plant uptake of heavy metals and persistent organic pollutants from soil. Environ Pollut. 2003; 124(3):375–8. PMID: [12758018](https://pubmed.ncbi.nlm.nih.gov/12758018/)
47. Hartmann A, Schmid M, van Tuinen D, Berg G. Plant-driven selection of microbes. Plant Soil. 2008; 321(1–2):235–57.
48. Reisberg EE, Hildebrandt U, Riederer M, Hentschel U. Distinct phyllosphere bacterial communities on Arabidopsis wax mutant leaves. PLoS One. 2013; 8(11):e78613. PMID: [24223831](https://pubmed.ncbi.nlm.nih.gov/24223831/). Pubmed Central PMCID: 3818481. doi: [10.1371/journal.pone.0078613](https://doi.org/10.1371/journal.pone.0078613)
49. Blackmore S, Wortley AH, Skvarla JJ, Rowley JR. Pollen wall development in flowering plants. New Phytol. 2007; 174(3):483–98. PMID: [17447905](https://pubmed.ncbi.nlm.nih.gov/17447905/).

50. Zinkl GM, Zwiebel BI, Grier DG, Preuss D. Pollen-stigma adhesion in *Arabidopsis*: a species-specific interaction mediated by lipophilic molecules in the pollen exine. *Development*. 1999; 126:5431–40. PMID: [10556067](#)
51. Scagel CF, Andersen CP. Seasonal changes in root and soil respiration of ozone-exposed ponderosa pine (*Pinus ponderosa*) grown in different substrates. *New Phytol*. 1997; 136:627–43.
52. Chen Z, Wang X, Feng Z, Xiao Q, Duan X. Impact of elevated O₃ on soil microbial community function under wheat crop. *Water Air Soil Pollut*. 2008; 198(1–4):189–98.
53. Avnery S, Mauzerall DL, Fiore AM. Increasing global agricultural production by reducing ozone damages via methane emission controls and ozone-resistant cultivar selection. *Glob Chang Biol*. 2013; 19(4):1285–99. PMID: [23504903](#). Pubmed Central PMCID: 3627305. doi: [10.1111/gcb.12118](#)
54. Takahashi M, Nakagawa M, Sakamoto A, Ohsumi C, Matsubara T, Morikawa H. Atmospheric nitrogen dioxide gas is a plant vitalization signal to increase plant size and the contents of cell constituents. *New Phytol*. 2005; 168(1):149–54. PMID: [16159329](#).
55. Hu Y, Bellaloui N, Tigabu M, Wang J, Diao J, Wang K, et al. Gaseous NO₂ effects on stomatal behavior, photosynthesis and respiration of hybrid poplar leaves. *Acta Physiol Plant*. 2015; 37(39):1–8.
56. Tiwari S, Agrawal M, Marshall FM. Evaluation of ambient air pollution impact on carrot plants at a suburban site using open top chambers. *Environ Monit Assess*. 2006; 119(1–3):15–30. PMID: [16736274](#).
57. Baxter A, Mittler R, Suzuki N. ROS as key players in plant stress signalling. *J Exp Bot*. 2014; 65(5):1229–40. PMID: [24253197](#). doi: [10.1093/jxb/ert375](#)
58. Barberán A, Ladau J, Leff JW, Pollard KS, Menninger HL, Dunn RR, et al. Continental-scale distributions of dust-associated bacteria and fungi. *PNAS*. 2015; 112(18):5756–61. doi: [10.1073/pnas.1420815112](#) PMID: [25902536](#)
59. Hanski I, von Hertzen L, Fyhrquist N, Koskinen K, Torppa K, Laatikainen T, et al. Environmental biodiversity, human microbiota, and allergy are interrelated. *PNAS*. 2012; 109(21):8334–9. doi: [10.1073/pnas.1205624109](#) PMID: [22566627](#)
60. Bowers RM, McLetchie S, Knight R, Fierer N. Spatial variability in airborne bacterial communities across land-use types and their relationship to the bacterial communities of potential source environments. *ISME J*. 2011; 5(4):601–12. PMID: [21048802](#). Pubmed Central PMCID: 3105744. doi: [10.1038/ismej.2010.167](#)
61. McKinney ML. Urbanization as a major cause of biotic homogenization. *Biol Conserv*. 2006; 127(3):247–60.
62. Zipfel C. Plant pattern-recognition receptors. *Trends Immunol*. 2014; 35(7):345–51. PMID: [24946686](#). doi: [10.1016/j.it.2014.05.004](#)