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**Impact of pollen associated lipid mediators (PALMs) from grass pollen on human mast cells**

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We recently demonstrated that pollen do not only function as allergen carriers but are also a rich source of bioactive lipids. These pollen-associated lipid mediators (PALMs) act as immunostimulators and -modulators on cells of the innate and adaptive immune system. Herein we aimed to investigate the impact of water-soluble factors from grass pollen (*Phleum pratense* L.), their Hexane isopropanol total lipid extracts (HIP), RP-HPLC fractions (RP) from HIP and associated phytoprostanes (PPE1) on human mast cells# effector functions such as degranulation. IgE- and Calcium-Ionophore-mediated mast cell degranulation was documented by  $\beta$ -hexosaminidase release. The human mast cell line LAD2 as well as primary mast cells (PMCs) isolated from human fore-skin were sensitized with or without human myeloma IgE for 16 h. After centrifugation, they were stimulated with anti-human IgE or Calcium-Ionophore following pretreatment with *Phleum pratense* L. aqueous pollen extracts (Phl.-APE), HIP, RP and PPE1. The  $\beta$ -hexosaminidase content in supernatants and cell pellets was measured by p-nitrophenyl-acetyl-glucosaminide formation. Water-soluble factors from pollen (Phl.-APE) dose-dependently enhanced the Calcium-Ionophore and IgE/ $\alpha$ -IgE-mediated degranulation. Additionally, pollen derived lipids such as HIP-extracts and RP-HPLC-fractions also synergistically increased specific and unspecific mast cell degranulation. A similar potentiation of IgE/ $\alpha$ -IgE-mediated degranulation was observed by PPE1. In the absence of IgE-receptor crosslinking only Phl.-APE was able to induce mast cell degranulation. All these outcomes were observed in LAD2 as well as in PMCs. Our results suggest that pollen-associated lipid mediators (PALMs) such as PPE1 may profoundly influence mast cell degranulation. The mechanisms leading to this effect are currently under